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(54) Title: METHODS TO ENGINEER MAMMALIAN-TYPE CARBOHYDRATE STRUCTURES

(57) **Abstract:** The present invention relates to host cells having modified lipid-linked oligosaccharides which may be modified further by heterologous expression of a set of glycosyltransferases, sugar transporters and mannosidases to become host-strains for the production of mammalian, e.g., human therapeutic glycoproteins. The process provides an engineered host cell which can be used to express and target any desirable gene(s) involved in glycosylation. Host cells with modified lipid-linked oligosaccharides are created or selected. N-glycans made in the engineered host cells have a GlcNAcMan₃GlcNAc₂ core structure which may then be modified further by heterologous expression of one or more enzymes, e.g., glycosyl-transferases, sugar transporters and mannosidases, to yield human-like glycoproteins. For the production of therapeutic proteins, this method may be adapted to engineer cell lines in which any desired glycosylation structure may be obtained.

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METHODS TO ENGINEER MAMMALIAN-TYPE CARBOHYDRATE
STRUCTURES

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims priority to U. S. provisional application Ser. No.
5 60/344,169, Dec. 27, 2001, which is incorporated by reference herein in its
entirety.

FIELD OF THE INVENTION

[0002] The present invention generally relates to modifying the glycosylation
structures of recombinant proteins expressed in fungi or other lower eukaryotes, to
10 more closely resemble the glycosylation of proteins of higher mammals, in
particular humans.

BACKGROUND OF THE INVENTION

[0003] After DNA is transcribed and translated into a protein, further post
15 translational processing involves the attachment of sugar residues, a process known
as glycosylation. Different organisms produce different glycosylation enzymes
(glycosyltransferases and glycosidases), and have different substrates (nucleotide
sugars) available, so that the glycosylation patterns as well as composition of the
individual oligosaccharides, even of one and the same protein, will be different
20 depending on the host system in which the particular protein is being expressed.
Bacteria typically do not glycosylate proteins, and if so only in a very unspecific
manner (Moens, 1997). Lower eukaryotes such as filamentous fungi and yeast add

primarily mannose and mannosylphosphate sugars, whereas insect cells such as Sf9 cells glycosylate proteins in yet another way. See for example (Bretthauer, 1999; Martinet, 1998; Weikert, 1999; Malissard, 2000; Jarvis, 1998; and Takeuchi, 1997).

5 [0004] Synthesis of a mammalian-type oligosaccharide structure consists of a series of reactions in the course of which sugar-residues are added and removed while the protein moves along the secretory pathway in the host organism. The enzymes which reside along the glycosylation pathway of the host organism or cell determine what the resulting glycosylation patterns of secreted proteins.

10 Unfortunately, the resulting glycosylation pattern of proteins expressed in lower eukaryotic host cells differs substantially from the glycosylation found in higher eukaryotes such as humans and other mammals (Bretthauer, 1999). Moreover, the vastly different glycosylation pattern has, in some cases, been shown to increase the immunogenicity of these proteins in humans and reduce their half-life
15 (Takeuchi, 1997). It would be desirable to produce human-like glycoproteins in non-human host cells, especially lower eukaryotic cells.

[0005] The early steps of human glycosylation can be divided into at least two different phases: (i) lipid-linked $\text{Glc}_3\text{Man}_9\text{GlcNAc}_2$ oligosaccharides are assembled by a sequential set of reactions at the membrane of the endoplasmic reticulum (ER)
20 and (ii) the transfer of this oligosaccharide from the lipid anchor dolichyl pyrophosphate onto *de novo* synthesized protein. The site of the specific transfer is defined by an asparagine (Asn) residue in the sequence Asn-Xaa-Ser/Thr (see Fig. 1), where Xaa can be any amino acid except proline (Gavel, 1990). Further processing by glucosidases and mannosidases occurs in the ER before the nascent
25 glycoprotein is transferred to the early Golgi apparatus, where additional mannose residues are removed by Golgi specific alpha (α)-1,2-mannosidases. Processing continues as the protein proceeds through the Golgi. In the medial Golgi, a number of modifying enzymes, including N-acetylglucosaminyltransferases (GnT I, GnT II, GnT III, GnT IV GnT V GnT VI), mannosidase II and
30 fucosyltransferases, add and remove specific sugar residues (see, e.g., Figs. 2 and 3). Finally, in the trans-Golgi, galactosyltransferases and sialyltransferases produce a glycoprotein structure that is released from the Golgi. It is this structure,

characterized by bi-, tri- and tetra-antennary structures, containing galactose, fucose, N-acetylglucosamine and a high degree of terminal sialic acid, that gives glycoproteins their human characteristics.

[0006] In nearly all eukaryotes, glycoproteins are derived from the common core oligosaccharide precursor $\text{Glc}_3\text{Man}_9\text{GlcNAc}_2\text{-PP-Dol}$, where PP-Dol stands for dolichol-pyrophosphate (**Fig. 1**). Within the endoplasmic reticulum, synthesis and processing of dolichol pyrophosphate bound oligosaccharides are identical between all known eukaryotes. However, further processing of the core oligosaccharide by yeast, once it has been transferred to a peptide leaving the ER and entering the Golgi, differs significantly from humans as it moves along the secretory pathway and involves the addition of several mannose sugars.

[0007] In yeast, these steps are catalyzed by Golgi residing mannosyltransferases, like Och1p, Mnt1p and Mnn1p, which sequentially add mannose sugars to the core oligosaccharide. The resulting structure is undesirable for the production of humanoid proteins and it is thus desirable to reduce or eliminate mannosyltransferase activity. Mutants of *S. cerevisiae*, deficient in mannosyltransferase activity (for example *och1* or *mnn9* mutants) have been shown to be non-lethal and display a reduced mannose content in the oligosaccharide of yeast glycoproteins. Other oligosaccharide processing enzymes, such as mannosylphosphate transferase may also have to be eliminated depending on the host's particular endogenous glycosylation pattern.

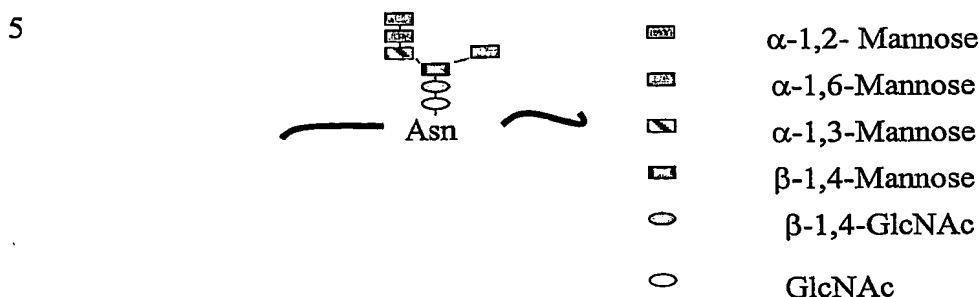
Lipid-Linked Oligosaccharide Precursors

[0008] Of particular interest for this invention are the early steps of N-glycosylation (**Figs. 1 and 2**). The study of *alg* (asparagine-linked glycosylation) mutants defective in the biosynthesis of the $\text{Glc}_3\text{Man}_9\text{GlcNAc}_2\text{-PP-Dol}$ has helped to elucidate the initial steps of N-glycosylation.

[0009] The *ALG3* gene of *S. cerevisiae* has been successfully cloned and knocked out by deletion (Aebi, 1996). *ALG3* has been shown to encode the enzyme Dol-P-Man:Man₅GlcNAc₂-PP-Dol Mannosyltransferase, which is involved in the first Dol-P-Man dependent mannosylation step from Man₅GlcNAc₂-PP-Dol to Man₆GlcNAc₂-PP-Dol at the luminal side of the ER (Sharma, 2001) (**Figs 1 and**

2). *S. cerevisiae* cells harboring a leaky *alg3-1* mutation accumulate Man₅GlcNAc₂-PP-Dol (structure I) (Huffaker, 1983).

Structure I: Man₅GlcNAc₂



Man₅GlcNAc₂ (Structure I) and Man₈GlcNAc₂ accumulate in total cell mannoprotein of an *och1 mnn1 alg3* mutant (Nakanishi-Shindo, 1993). This *S.cerevisiae och1, mnn1, alg3* mutant was shown to be viable, but temperature-sensitive, and to lack α -1,6 polymannose outer chains.

[0010] In another study, secretory proteins expressed in a strain deleted for *alg3* (Δ *alg3* background) were studied for their resistance to Endo- β -N-acetylglucosaminidase H (Endo H) (Aebi, 1996). Previous observations have indicated that only those oligosaccharides larger than Man₅GlcNAc₂ are susceptible to cleavage by Endo H (Hubbard, 1980). In the *alg3-1* phenotype, some glycoforms were sensitive to Endo H cleavage, confirming its leakiness, whereas in the Δ *alg3* mutant all glycoforms appeared to be resistant and of the Man₅-type (Aebi, 1996), suggesting a tight phenotype and transfer of Man₅GlcNAc₂ oligosaccharide structures onto the nascent polypeptide chain. No obvious phenotype was connected with the inactivation of the *ALG3* gene (Aebi, 1996). Secreted exoglucanase produced in a *Saccharomyces cerevisiae alg3* mutant was found to contain between 35-44% underglycosylated and unglycosylated forms and only about 50% of the transferred oligosaccharides remained resistant to Endo H treatment (Cueva, 1996). Exoglucanase (Exg), an enzyme that contains two potential N-glycosylation sites at Asn₁₆₅ and Asn₃₂₅, was analyzed in more detail. For Exg molecules that received two oligosaccharides it was shown that the first N-glycosylation site (Asn₁₆₅) was enriched in truncated

residues, whereas the second (Asn₃₂₅) was enriched in regular oligosaccharides. 35-44% of secreted exoglucanase was non- or underglycosylated and about 73 - 78 % of all available N-glycosylation sites were occupied with either truncated or regular oligosaccharides (Cueva, 1996).

5 **Transfer of Glucosylated Lipid-Linked Oligosaccharides**

- [0011] Evidence suggests that, in mammalian cells, only glucosylated lipid-linked oligosaccharides are transferred to nascent proteins (Turco, 1977), while in yeast *alg5*, *alg6* and *dpg1* mutants, nonglucosylated oligosaccharides can be transferred (Ballou, 1986; Runge, 1984). In a *Saccharomyces cerevisiae alg8* mutant, underglucosylated GlcMan₉GlcNAc₂ is transferred (Runge, 1986). Verostek and co-workers studied an *alg3*, *sec18*, *gls1* mutant and proposed that glucosylation of a Man₅GlcNAc₂ structure (Structure I, above) is relatively slow in comparison to glucosylation of a lipid-linked Man₉ structure. In addition, the transfer of this Man₅GlcNAc₂ structure to protein appears to be about 5-fold more efficient than the glucosylation to Glc₃Man₅GlcNAc₂. The decreased rate of Man₅GlcNAc₂ glucosylation in combination with the comparatively faster rate of Man₅ structure transfer onto nascent protein is believed to be the cause of the observed accumulation of nonglucosylated Man₅ structures in *alg3* mutant yeast (Verostek-a, 1993; Verostek-b, 1993).
- 20 [0012] Studies preceding the above work did not reveal any lipid-linked glucosylated oligosaccharides (Orlean, 1990; Huffaker, 1983) allowing the conclusion that glucosylated oligosaccharides are transferred at a much higher rate than their nonglucosylated counterparts and thus are much harder to isolate. Recent work has allowed the creation and study of yeast strains with un- and hypoglucosylated oligosaccharides and has further confirmed the importance of the addition of glucose to the antenna of lipid-linked oligosaccharides for substrate recognition by the oligosaccharyltransferase complex (Reiss, 1996; Stagljar, 1994; Burda, 1998). The decreased degree of glucosylation of the lipid-linked Man₅-oligosaccharides in an *alg3* mutant negatively impacts the kinetics of the transfer of lipid-linked oligosaccharides onto nascent protein and is believed to be the cause for the strong underglycosylation of secreted proteins in an *alg3* knock-out strain (Aebi, 1996).
- 30

[0013] The assembly of the lipid-linked core oligosaccharide $\text{Man}_9\text{GlcNAc}_2$ occurs, as described above, at the membrane of the endoplasmic reticulum. The additions of three glucose units to the α -1,3-antenna of the lipid-linked oligosaccharides are the final reactions in the oligosaccharide assembly. First an α -1,3 glucose residue is added followed by another α -1,3 glucose residue and a terminal α -1,2 glucose residue. Mutants accumulating dolichol-linked $\text{Man}_9\text{GlcNAc}_2$ have been shown to be defective in the *ALG6* locus, and Alg6p has similarities to Alg8p, the α -1,3-glucosyltransferase catalyzing the addition of the second α -1,3-linked glucose (Reiss, 1996). Cells with a defective *ALG8* locus accumulate dolichol-linked $\text{Glc}_1\text{Man}_9\text{GlcNAc}_2$ (Runge, 1986; Stagljar, 1994). The *ALG10* locus encodes the α -1,2 glucosyltransferase responsible for the addition of a single terminal glucose to $\text{Glc}_2\text{Man}_9\text{GlcNAc}_2\text{-PP-Dol}$ (Burda, 1998).

Sequential Processing of N-glycans by Localized Enzyme Activities

[0014] Sugar transferases and mannosidases line the inner (luminal) surface of the ER and Golgi apparatus and thereby provide a "catalytic" surface that allows for the sequential processing of glycoproteins as they proceed through the ER and Golgi network. In fact the multiple compartments of the cis, medial, and trans Golgi and the trans-Golgi Network (TGN), provide the different localities in which the ordered sequence of glycosylation reactions can take place. As a glycoprotein proceeds from synthesis in the ER to full maturation in the late Golgi or TGN, it is sequentially exposed to different glycosidases, mannosidases and glycosyltransferases such that a specific carbohydrate structure may be synthesized. Much work has been dedicated to revealing the exact mechanism by which these enzymes are retained and anchored to their respective organelle. The evolving picture is complex but evidence suggests that, stem region, membrane spanning region and cytoplasmic tail individually or in concert direct enzymes to the membrane of individual organelles and thereby localize the associated catalytic domain to that locus.

[0015] In some cases these specific interactions were found to function across species. For example the membrane spanning domain of α 2,6-ST from rats, an enzyme known to localize in the trans-Golgi of the animal, was shown to also localize a reporter gene (invertase) in the yeast Golgi (Schwientek, 1995).

However, the very same membrane spanning domain as part of a full-length $\alpha 2,6$ ST was retained in the ER and not further transported to the Golgi of yeast (Krezdorn, 1994). A full length Gal-Tr from humans was not even synthesized in yeast, despite demonstrably high transcription levels. On the other hand the transmembrane region of human the same GalT fused to an invertase reporter was able to direct localization to the yeast Golgi, albeit it at low production levels. Schwientek and co-workers have shown that fusing 28 amino acids of a yeast mannosyltransferase (Mnt1), a region containing a cytoplasmic tail, a transmembrane region and eight amino acids of the stem region, to the catalytic domain of human GalT are sufficient for Golgi localization of an active GalT. Other galactosyltransferases appear to rely on interactions with enzymes resident in particular organelles since after removal of their transmembrane region they are still able to localize properly. To date there exists no reliable way of predicting whether a particular heterologously expressed glycosyltransferase or mannosidase in a lower eukaryote will be (1), sufficiently translated (2), catalytically active or (3) located to the proper organelle within the secretory pathway. Since all three of these are necessary to effect glycosylation patterns in lower eukaryotes, a systematic scheme to achieve the desired catalytic function and proper retention of enzymes in the absence of predictive tools, which are currently not available, has been designed.

Production of Therapeutic Glycoproteins

[0016] A significant number of proteins isolated from humans or animals are post-translationally modified, with glycosylation being one of the most significant modifications. An estimated 70% of all therapeutic proteins are glycosylated and thus currently rely on a production system (i.e., host cell) that is able to glycosylate in a manner similar to humans. To date, most glycoproteins are made in a mammalian host system. Several studies have shown that glycosylation plays an important role in determining the (1) immunogenicity, (2) pharmacokinetic properties, (3) trafficking, and (4) efficacy of therapeutic proteins. It is thus not surprising that substantial efforts by the pharmaceutical industry have been directed at developing processes to obtain glycoproteins that are as "humanoid" or "human-like" as possible. This may involve the genetic engineering of such

mammalian cells to enhance the degree of sialylation (i.e., terminal addition of sialic acid) of proteins expressed by the cells, which is known to improve pharmacokinetic properties of such proteins. Alternatively one may improve the degree of sialylation by *in vitro* addition of such sugars using known glycosyltransferases and their respective nucleotide sugars (e.g., 2,3 sialyltransferase and CMP-Sialic acid).

[0017] Future research may reveal the biological and therapeutic significance of specific glycoforms, thereby rendering the ability to produce such specific glycoforms desirable. To date, efforts have concentrated on making proteins with fairly well characterized glycosylation patterns, and expressing a cDNA encoding such a protein in one of the following higher eukaryotic protein expression systems:

1. Higher eukaryotes such as Chinese hamster ovary cells (CHO), mouse fibroblast cells and mouse myeloma cells (Werner, 1998);
2. Transgenic animals such as goats, sheep, mice and others (Dente, 1988); (Cole, 1994); (McGarvey, 1995); (Bardor, 1999);
3. Plants (*Arabidopsis thaliana*, tobacco etc.) (Staub, 2000); (McGarvey, 1995); (Bardor, 1999);
4. Insect cells (*Spodoptera frugiperda* Sf9, Sf21, *Trichoplusia ni*, etc., in combination with recombinant baculoviruses such as *Autographa californica* multiple nuclear polyhedrosis virus which infects lepidopteran cells (Altmann, 1999).

[0018] While most higher eukaryotes carry out glycosylation reactions that are similar to those found in humans, recombinant human proteins expressed in the above mentioned host systems invariably differ from their “natural” human counterpart (Raju, 2000). Extensive development work has thus been directed at finding ways to improving the “human character” of proteins made in these expression systems. This includes the optimization of fermentation conditions and the genetic modification of protein expression hosts by introducing genes encoding enzymes involved in the formation of human like glycoforms (Werner, 1998); (Weikert, 1999); (Andersen, 1994); (Yang, 2000). Inherent problems associated with all mammalian expression systems have not been solved.

[0019] Fermentation processes based on mammalian cell culture (e.g., CHO, murine, or human cells), for example, tend to be very slow (fermentation times in excess of one week are not uncommon), often yield low product titers, require expensive nutrients and cofactors (e.g., bovine fetal serum), are limited by
5 programmed cell death (apoptosis), and often do not enable expression of particular therapeutically valuable proteins. More importantly, mammalian cells are susceptible to viruses that have the potential to be human pathogens and stringent quality controls are required to assure product safety. This is of particular concern since many such processes require the addition of complex and
10 temperature sensitive media components that are derived from animals (e.g., bovine calf serum), which may carry agents pathogenic to humans such as bovine spongiform encephalopathy (BSE) prions or viruses. Moreover, the production of therapeutic compounds is preferably carried out in a well-controlled sterile environment. An animal farm, no matter how cleanly kept, does not constitute
15 such an environment, thus constituting an additional problem in the use of transgenic animals for manufacturing high volume therapeutic proteins.

[0020] Most, if not all, currently produced therapeutic glycoproteins are therefore expressed in mammalian cells and much effort has been directed at improving (i.e., “humanizing”) the glycosylation pattern of these recombinant proteins. Changes in
20 medium composition as well as the co-expression of genes encoding enzymes involved in human glycosylation have been successfully employed (see, for example, Weikert, 1999).

[0021] While recombinant proteins similar to their human counterparts can be made in mammalian expression systems, it is currently not possible to make
25 proteins with a human-like glycosylation pattern in lower eukaryotes (fungi and yeast). Although the core oligosaccharide structure transferred to a protein in the endoplasmic reticulum is basically identical in mammals and lower eukaryotes, substantial differences have been found in the subsequent processing reactions which occur in in the Golgi apparatus of fungi and mammals. In fact, even
30 amongst different lower eukaryotes there exist a great variety of glycosylation structures. This has prevented the use of lower eukaryotes as hosts for the production of recombinant human glycoproteins despite otherwise notable

advantages over mammalian expression systems, such as: (1) generally higher product titers, (2) shorter fermentation times, (3) having an alternative for proteins that are poorly expressed in mammalian cells, (4) the ability to grow in a chemically defined protein free medium and thus not requiring complex animal derived media components, (5) and the absence of viral, especially retroviral infections of such hosts.

[0022] Various methylotrophic yeasts such as *Pichia pastoris*, *Pichia methanolica*, and *Hansenula polymorpha*, have played particularly important roles as eukaryotic expression systems because they are able to grow to high cell densities and secrete large quantities of recombinant protein. However, as noted above, lower eukaryotes such as yeast do not glycosylate proteins like higher mammals. See for example, Martinet *et al.* (1998) Biotechnol Let. Vol. 20. No.12, which discloses the expression of a heterologous mannosidase in the endoplasmic reticulum (ER).

[0023] Chiba et al. (1998) have shown that *S.cerevisiae* can be engineered to provide structures ranging from $\text{Man}_8\text{GlcNAc}_2$ to $\text{Man}_5\text{GlcNAc}_2$ structures, by eliminating 1,6 mannosyltransferase (*OCH1*), 1,3 mannosyltransferase (*MNN1*) and a regulator of mannosylphosphatetransferase (*MNN4*) and by targeting the catalytic domain of α -1,2-mannosidase I from *Aspergillus saitoi* into the ER of *S.cerevisiae* using an ER retrieval sequence (Chiba, 1998). However, this attempt resulted in little or no production of the desired $\text{Man}_5\text{GlcNAc}_2$, e.g., one that was made *in vivo* and which could function as a substrate for GnT1 (the next step in making human-like glycan structures). Chiba et al. (1998) showed that *P. pastoris* is not inherently able to produce useful quantities (greater than 5%) of GlcNAcTransferase I accepting carbohydrate.

[0024] Maras and co-workers assert that in *T. reesei* "sufficient concentrations of acceptor substrate (i.e. $\text{Man}_5\text{GlcNAc}_2$) are present", however when trying to convert this acceptor substrate to $\text{GlcNAcMan}_5\text{GlcNAc}_2$ *in vitro* less than 2% were converted thereby demonstrating the presence of $\text{Man}_5\text{GlcNAc}_2$ structures that are not suitable precursors for complex N-glycan formation (Maras, 1997; Maras, 1999). To date no enabling disclosure exists, that allows for the production of commercially relevant quantities of $\text{GlcNAcMan}_5\text{GlcNAc}_2$ in lower eukaryotes.

[0025] It is therefore an object of the present invention to provide a system and methods for humanizing glycosylation of recombinant glycoproteins expressed in non-human host cells.

5

SUMMARY OF THE INVENTION

[0026] The present invention relates to host cells such as fungal strains having modified lipid-linked oligosaccharides which may be modified further by heterologous expression of a set of glycosyltransferases, sugar transporters and mannosidases to become host-strains for the production of mammalian, e.g., human therapeutic glycoproteins. A protein production method has been developed using (1) a lower eukaryotic host such as a unicellular or filamentous fungus, or (2) any non-human eukaryotic organism that has a different glycosylation pattern from humans, to modify the glycosylation composition and structures of the proteins made in a host organism ("host cell") so that they resemble more closely carbohydrate structures found in human proteins. The process allows one to obtain an engineered host cell which can be used to express and target any desirable gene(s) involved in glycosylation by methods that are well established in the scientific literature and generally known to the artisan in the field of protein expression. As described herein, host cells with modified lipid-linked oligosaccharides are created or selected. N-glycans made in the engineered host cells have a GlcNAcMan₃GlcNAc₂ core structure which may then be modified further by heterologous expression of one or more enzymes, e.g., glycosyltransferases, sugar transporters and mannosidases, to yield human-like glycoproteins. For the production of therapeutic proteins, this method may be adapted to engineer cell lines in which any desired glycosylation structure may be obtained.

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BRIEF DESCRIPTION OF THE DRAWINGS

[0027] **Figure 1** is a schematic of the structure of the dolichyl pyrophosphate-linked oligosaccharide.

30

[0028] **Figure 2** is a schematic of the generation of GlcNAc₂Man₃GlcNAc₂ N-glycans from fungal host cells which are deficient in *alg3*, *alg9* or *alg12* activities.

[0029] **Figure 3** is a schematic of processing reactions required to produce mammalian-type oligosaccharide structures in a fungal host cell with an *alg3*, *och1* genotype.

[0030] **Figure 4** shows *S. cerevisiae* *Alg3* Sequence Comparisons (Blast)

[0031] **Figure 5** shows *S. cerevisiae* *Alg 3* and *Alg 3p* Sequences

[0032] **Figure 6** shows *P. pastoris* *Alg 3* and *Alg 3p* Sequences

[0033] **Figure 7** shows *P. pastoris* *Alg 3* Sequence Comparisons (Blast)

10 [0034] **Figure 8** shows *K. lactis* *Alg 3* and *Alg 3p* Sequences

[0035] **Figure 9** shows *K. lactis* *Alg 3* Sequence Comparisons (Blast)

[0036] **Figure 10** shows *S. cerevisiae* *Alg 9* and *Alg 9p* Sequences

[0037] **Figure 11** shows *P. pastoris* *Alg 9* and *Alg 9p* Sequences

[0038] **Figure 12** shows *P. pastoris* *Alg 9* Sequence Comparisons (Blast)

15 [0039] **Figure 13** shows *S. cerevisiae* *Alg 12* and *Alg 12p* Sequences

[0040] **Figure 14** shows *P. pastoris* *Alg 12* and *Alg 12p* Sequences

[0041] **Figure 15** shows *P. pastoris* *Alg 12* Sequence Comparisons (Blast)

[0042] **Figure 16** is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P.pastoris* showing that the predominant N-glycan is GlcNAcMan₅GlcNAc₂.

[0043] **Figure 17** is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P.pastoris* (Fig. 16) treated with β -N-hexosaminidase (peak corresponding to Man₅GlcNAc₂) to confirm that the predominant N-glycan of Fig. 16 is GlcNAcMan₅GlcNAc₂.

25 [0044] **Figure 18** is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P.pastoris* *alg3* deletion mutant showing that the predominant N-glycans are GlcNAcMan₃GlcNAc₂ and GlcNAcMan₄GlcNAc₂.

[0045] **Figure 19** is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P.pastoris* *alg3* deletion mutant treated with α 1,2 mannosidase, showing that the GlcNAcMan₄GlcNAc₂ of Fig. 18 is converted to GlcNAcMan₃GlcNAc₂.

30

[0046] Figure 20 is a MALDI-TOF-MS analysis of N-glycans of Fig. 19 treated with β -N-hexosaminidase (peak corresponding to $\text{Man}_3\text{GlcNAc}_2$) to confirm that the N-glycan of Fig. 19 is $\text{GlcNAcMan}_3\text{GlcNAc}_2$.

5 [0047] Figure 21 is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P.pastoris alg3* deletion mutant treated with α 1,2 mannosidase and GnTII, showing that the $\text{GlcNAcMan}_3\text{GlcNAc}_2$ of Fig. 19 is converted to $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$.

[0048] Figure 22 is a MALDI-TOF-MS analysis of N-glycans of Fig. 21 treated with β -N-hexosaminidase (peak corresponding to $\text{Man}_3\text{GlcNAc}_2$) to confirm that
10 the N-glycan of Fig. 21 is $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$.

[0049] Figure 23 is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P.pastoris alg3* deletion mutant treated with α 1,2 mannosidase and GnTII in the presence of UDP-galactose and β 1,4-galactosyltransferase, showing that the $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$ of Fig. 21 is
15 converted to $\text{Gal}_2\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$.

[0050] Figure 24 is a MALDI-TOF-MS analysis of N-glycans isolated from a kringle 3 glycoprotein produced in a *P.pastoris alg3* deletion mutant treated with α 1,2 mannosidase and GnTII in the presence of UDP-galactose and β 1,4-galactosyltransferase, and further treated with CMP-N-acetylneuraminic acid and
20 sialyltransferase, showing that the $\text{Gal}_2\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$ is converted to $\text{NANA}_2\text{Gal}_2\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$.

[0051] Figure 25 shows *S. cerevisiae Alg6* and Alg 6p Sequences

[0052] Figure 26 shows *P. pastoris Alg6* and Alg 6p Sequences

[0053] Figure 27 shows *P. pastoris Alg 6* Sequence Comparisons (Blast)

25 [0054] Figure 28 shows *K.lactis Alg6* and Alg 6p Sequences

[0055] Figure 29 shows *K.lactis Alg 6* Sequence Comparisons (Blast)

[0056] Figure 30 Model of an IgG immunoglobulin. Heavy chain and light chain can be, based on similar secondary and tertiary structure, subdivided into domains. The two heavy chains (domains V_H , C_{H1} , C_{H2} and C_{H3}) are linked
30 through three disulfide bridges. The light chains (domains V_L and C_L) are linked by another disulfide bridge to the C_{H1} portion of the heavy chain and, together with the C_{H1} and V_H fragments, make up the Fab region. Antigens bind to the terminal

portion of the Fab region. Effector-functions, such as Fc-gamma-Receptor binding have been localized to the C_H2 domain, just downstream of the hinge region and are influenced by N-glycosylation of asparagine 297 in the heavy chain.

[0057] **Figure 31** Schematic overview of a modular IgG1 expression vector.

5 [0058] **Figure 32** shows *M. musculus GnT III* Nucleic Acid And Amino Acid Sequences

[0059] **Figure 33** shows *H. sapiens GnT IV* Nucleic Acid And Amino Acid Sequences

[0060] **Figure 34** shows *M. musculus GnT V* Nucleic Acid And Amino Acid
10 Sequences

DETAILED DESCRIPTION OF THE INVENTION

[0061] Unless otherwise defined herein, scientific and technical terms used in connection with the present invention shall have the meanings that are commonly
15 understood by those of ordinary skill in the art. Further, unless otherwise required by context, singular terms shall include pluralities and plural terms shall include the singular. The methods and techniques of the present invention are generally performed according to conventional methods well known in the art. Generally, nomenclatures used in connection with, and techniques of biochemistry,
20 enzymology, molecular and cellular biology, microbiology, genetics and protein and nucleic acid chemistry and hybridization described herein are those well known and commonly used in the art. The methods and techniques of the present invention are generally performed according to conventional methods well known in the art and as described in various general and more specific references that are
25 cited and discussed throughout the present specification unless otherwise indicated. See, e.g., Sambrook et al. Molecular Cloning: A Laboratory Manual, 2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1989); Ausubel et al., Current Protocols in Molecular Biology, Greene Publishing Associates (1992, and Supplements to 2002); Harlow and Lane Antibodies: A Laboratory Manual Cold
30 Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1990); Introduction to Glycobiology, Maureen E. Taylor, Kurt Drickamer, Oxford Univ. Press (2003); Worthington Enzyme Manual, Worthington Biochemical Corp. Freehold, NJ;

Handbook of Biochemistry: Section A Proteins Vol I 1976 CRC Press; Handbook of Biochemistry: Section A Proteins Vol II 1976 CRC Press; Essentials of Glycobiology, Cold Spring Harbor Laboratory Press (1999). The nomenclatures used in connection with, and the laboratory procedures and techniques of, biochemistry and molecular biology described herein are those well known and commonly used in the art.

[0062] All publications, patents and other references mentioned herein are incorporated by reference.

[0063] The following terms, unless otherwise indicated, shall be understood to have the following meanings:

[0064] As used herein, the term "N-glycan" refers to an N-linked oligosaccharide, e.g., one that is attached by an asparagine-N-acetylglucosamine linkage to an asparagine residue of a polypeptide. N-glycans have a common pentasaccharide core of $\text{Man}_3\text{GlcNAc}_2$ ("Man" refers to mannose; "Glc" refers to glucose; and "NAc" refers to N-acetyl; GlcNAc refers to N-acetylglucosamine). N-glycans differ with respect to the number of branches (antennae) comprising peripheral sugars (e.g., fucose and sialic acid) that are added to the $\text{Man}_3\text{GlcNAc}_2$ ("Man3") core structure. N-glycans are classified according to their branched constituents (e.g., high mannose, complex or hybrid). A "high mannose" type N-glycan has five or more mannose residues. A "complex" type N-glycan typically has at least one GlcNAc attached to the 1,3 mannose arm and at least one GlcNAc attached to the 1,6 mannose arm of a "trimannose" core. The "trimannose core" is the pentasaccharide core having a Man3 structure. Complex N-glycans may also have galactose ("Gal") residues that are optionally modified with sialic acid or derivatives ("NeuAc", where "Neu" refers to neuraminic acid and "Ac" refers to acetyl). Complex N-glycans may also have intrachain substitutions comprising "bisecting" GlcNAc and core fucose ("Fuc"). A "hybrid" N-glycan has at least one GlcNAc on the terminal of the 1,3 mannose arm of the trimannose core and zero or more mannoses on the 1,6 mannose arm of the trimannose core.

[0065] Abbreviations used herein are of common usage in the art, see, e.g., abbreviations of sugars, above. Other common abbreviations include "PNGase", which refers to peptide N-glycosidase F (EC 3.2.2.18); "GlcNAc Tr (I – III)",

which refers to one of three N-acetylglucosaminyltransferase enzymes; "NANA" refers to N-acetylneuraminic acid.

[0066] As used herein, the term "secretion pathway" refers to the assembly line of various glycosylation enzymes to which a lipid-linked oligosaccharide precursor and an N-glycan substrate are sequentially exposed, following the molecular flow of a nascent polypeptide chain from the cytoplasm to the endoplasmic reticulum (ER) and the compartments of the Golgi apparatus. Enzymes are said to be localized along this pathway. An enzyme X that acts on a lipid-linked glycan or an N-glycan before enzyme Y is said to be or to act "upstream" to enzyme Y; similarly, enzyme Y is or acts "downstream" from enzyme X.

[0067] As used herein, the term "alg X activity" refers to the enzymatic activity encoded by the "alg X" gene, and to an enzyme having that enzymatic activity encoded by a homologous gene or gene product (see below) or by an unrelated gene or gene product.

[0068] As used herein, the term "antibody" refers to a full antibody (consisting of two heavy chains and two light chains) or a fragment thereof. Such fragments include, but are not limited to, those produced by digestion with various proteases, those produced by chemical cleavage and/or chemical dissociation, and those produced recombinantly, so long as the fragment remains capable of specific binding to an antigen. Among these fragments are Fab, Fab', F(ab')₂, and single chain Fv (scFv) fragments. Within the scope of the term "antibody" are also antibodies that have been modified in sequence, but remain capable of specific binding to an antigen. Example of modified antibodies are interspecies chimeric and humanized antibodies; antibody fusions; and heteromeric antibody complexes, such as diabodies (bispecific antibodies), single-chain diabodies, and intrabodies (see, e.g., Marasco (ed.), Intracellular Antibodies: Research and Disease Applications, Springer-Verlag New York, Inc. (1998) (ISBN: 3540641513), the disclosure of which is incorporated herein by reference in its entirety).

[0069] As used herein, the term "mutation" refers to any change in the nucleic acid or amino acid sequence of a gene product, e.g., of a glycosylation-related enzyme.

15 [0070] The term “polynucleotide” or “nucleic acid molecule” refers to a polymeric form of nucleotides of at least 10 bases in length. The term includes DNA molecules (e.g., cDNA or genomic or synthetic DNA) and RNA molecules (e.g., mRNA or synthetic RNA), as well as analogs of DNA or RNA containing non-natural nucleotide analogs, non-native internucleoside bonds, or both. The nucleic acid can be in any topological conformation. For instance, the nucleic acid can be single-stranded, double-stranded, triple-stranded, quadruplexed, partially double-stranded, branched, hairpinned, circular, or in a padlocked conformation. The term includes single and double stranded forms of DNA.

10 [0071] Unless otherwise indicated, a “nucleic acid comprising SEQ ID NO:X” refers to a nucleic acid, at least a portion of which has either (i) the sequence of SEQ ID NO:X, or (ii) a sequence complementary to SEQ ID NO:X. The choice between the two is dictated by the context. For instance, if the nucleic acid is used as a probe, the choice between the two is dictated by the requirement that the probe
15 be complementary to the desired target.

[0072] An “isolated” or “substantially pure” nucleic acid or polynucleotide (e.g., an RNA, DNA or a mixed polymer) is one which is substantially separated from other cellular components that naturally accompany the native polynucleotide in its natural host cell, e.g., ribosomes, polymerases, and genomic sequences with which
20 it is naturally associated. The term embraces a nucleic acid or polynucleotide that (1) has been removed from its naturally occurring environment, (2) is not associated with all or a portion of a polynucleotide in which the “isolated polynucleotide” is found in nature, (3) is operatively linked to a polynucleotide which it is not linked to in nature, or (4) does not occur in nature. The term
25 “isolated” or “substantially pure” also can be used in reference to recombinant or cloned DNA isolates, chemically synthesized polynucleotide analogs, or polynucleotide analogs that are biologically synthesized by heterologous systems.

[0073] However, “isolated” does not necessarily require that the nucleic acid or polynucleotide so described has itself been physically removed from its native
30 environment. For instance, an endogenous nucleic acid sequence in the genome of an organism is deemed “isolated” herein if a heterologous sequence (i.e., a sequence that is not naturally adjacent to this endogenous nucleic acid sequence) is

placed adjacent to the endogenous nucleic acid sequence, such that the expression of this endogenous nucleic acid sequence is altered. By way of example, a non-native promoter sequence can be substituted (e.g., by homologous recombination) for the native promoter of a gene in the genome of a human cell, such that this
5 gene has an altered expression pattern. This gene would now become "isolated" because it is separated from at least some of the sequences that naturally flank it.

[0074] A nucleic acid is also considered "isolated" if it contains any modifications that do not naturally occur to the corresponding nucleic acid in a genome. For instance, an endogenous coding sequence is considered "isolated" if
10 it contains an insertion, deletion or a point mutation introduced artificially, e.g., by human intervention. An "isolated nucleic acid" also includes a nucleic acid integrated into a host cell chromosome at a heterologous site, a nucleic acid construct present as an episome. Moreover, an "isolated nucleic acid" can be substantially free of other cellular material, or substantially free of culture medium
15 when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized.

[0075] As used herein, the phrase "degenerate variant" of a reference nucleic acid sequence encompasses nucleic acid sequences that can be translated, according to the standard genetic code, to provide an amino acid sequence identical
20 to that translated from the reference nucleic acid sequence.

[0076] The term "percent sequence identity" or "identical" in the context of nucleic acid sequences refers to the residues in the two sequences which are the same when aligned for maximum correspondence. The length of sequence identity comparison may be over a stretch of at least about nine nucleotides, usually at least
25 about 20 nucleotides, more usually at least about 24 nucleotides, typically at least about 28 nucleotides, more typically at least about 32 nucleotides, and preferably at least about 36 or more nucleotides. There are a number of different algorithms known in the art which can be used to measure nucleotide sequence identity. For instance, polynucleotide sequences can be compared using FASTA, Gap or Bestfit,
30 which are programs in Wisconsin Package Version 10.0, Genetics Computer Group (GCG), Madison, Wisconsin. FASTA provides alignments and percent sequence identity of the regions of the best overlap between the query and search

sequences (Pearson, 1990, (herein incorporated by reference). For instance, percent sequence identity between nucleic acid sequences can be determined using FASTA with its default parameters (a word size of 6 and the NOPAM factor for the scoring matrix) or using Gap with its default parameters as provided in GCG Version 6.1, herein incorporated by reference.

[0077] The term “substantial homology” or “substantial similarity,” when referring to a nucleic acid or fragment thereof, indicates that, when optimally aligned with appropriate nucleotide insertions or deletions with another nucleic acid (or its complementary strand), there is nucleotide sequence identity in at least about 50%, more preferably 60% of the nucleotide bases, usually at least about 70%, more usually at least about 80%, preferably at least about 90%, and more preferably at least about 95%, 96%, 97%, 98% or 99% of the nucleotide bases, as measured by any well-known algorithm of sequence identity, such as FASTA, BLAST or Gap, as discussed above.

[0078] Alternatively, substantial homology or similarity exists when a nucleic acid or fragment thereof hybridizes to another nucleic acid, to a strand of another nucleic acid, or to the complementary strand thereof, under stringent hybridization conditions. “Stringent hybridization conditions” and “stringent wash conditions” in the context of nucleic acid hybridization experiments depend upon a number of different physical parameters. Nucleic acid hybridization will be affected by such conditions as salt concentration, temperature, solvents, the base composition of the hybridizing species, length of the complementary regions, and the number of nucleotide base mismatches between the hybridizing nucleic acids, as will be readily appreciated by those skilled in the art. One having ordinary skill in the art knows how to vary these parameters to achieve a particular stringency of hybridization.

[0079] In general, “stringent hybridization” is performed at about 25°C below the thermal melting point (T_m) for the specific DNA hybrid under a particular set of conditions. “Stringent washing” is performed at temperatures about 5°C lower than the T_m for the specific DNA hybrid under a particular set of conditions. The T_m is the temperature at which 50% of the target sequence hybridizes to a perfectly matched probe. See Sambrook et al., *supra*, page 9.51, hereby incorporated by

reference. For purposes herein, "high stringency conditions" are defined for solution phase hybridization as aqueous hybridization (i.e., free of formamide) in 6X SSC (where 20X SSC contains 3.0 M NaCl and 0.3 M sodium citrate), 1% SDS at 65°C for 8-12 hours, followed by two washes in 0.2X SSC, 0.1% SDS at 65°C for 20 minutes. It will be appreciated by the skilled worker that hybridization at 65°C will occur at different rates depending on a number of factors including the length and percent identity of the sequences which are hybridizing.

[0080] The nucleic acids (also referred to as polynucleotides) of this invention may include both sense and antisense strands of RNA, cDNA, genomic DNA, and synthetic forms and mixed polymers of the above. They may be modified chemically or biochemically or may contain non-natural or derivatized nucleotide bases, as will be readily appreciated by those of skill in the art. Such modifications include, for example, labels, methylation, substitution of one or more of the naturally occurring nucleotides with an analog, internucleotide modifications such as uncharged linkages (e.g., methyl phosphonates, phosphotriesters, phosphoramidates, carbamates, etc.), charged linkages (e.g., phosphorothioates, phosphorodithioates, etc.), pendent moieties (e.g., polypeptides), intercalators (e.g., acridine, psoralen, etc.), chelators, alkylators, and modified linkages (e.g., alpha anomeric nucleic acids, etc.) Also included are synthetic molecules that mimic polynucleotides in their ability to bind to a designated sequence via hydrogen bonding and other chemical interactions. Such molecules are known in the art and include, for example, those in which peptide linkages substitute for phosphate linkages in the backbone of the molecule.

[0081] The term "mutated" when applied to nucleic acid sequences means that nucleotides in a nucleic acid sequence may be inserted, deleted or changed compared to a reference nucleic acid sequence. A single alteration may be made at a locus (a point mutation) or multiple nucleotides may be inserted, deleted or changed at a single locus. In addition, one or more alterations may be made at any number of loci within a nucleic acid sequence. A nucleic acid sequence may be mutated by any method known in the art including but not limited to mutagenesis techniques such as "error-prone PCR" (a process for performing PCR under conditions where the copying fidelity of the DNA polymerase is low, such that a

high rate of point mutations is obtained along the entire length of the PCR product. See, e.g., Leung, D. W., et al., *Technique*, 1, pp. 11-15 (1989) and Caldwell, R. C. & Joyce G. F., *PCR Methods Applic.*, 2, pp. 28-33 (1992)); and "oligonucleotide-directed mutagenesis" (a process which enables the generation of site-specific mutations in any cloned DNA segment of interest. See, e.g., Reidhaar-Olson, J. F. & Sauer, R. T., et al., *Science*, 241, pp. 53-57 (1988)).

[0082] The term "vector" as used herein is intended to refer to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments may be ligated. Other vectors include cosmids, bacterial artificial chromosomes (BAC) and yeast artificial chromosomes (YAC). Another type of vector is a viral vector, wherein additional DNA segments may be ligated into the viral genome (discussed in more detail below). Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., vectors having an origin of replication which functions in the host cell). Other vectors can be integrated into the genome of a host cell upon introduction into the host cell, and are thereby replicated along with the host genome. Moreover, certain preferred vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "recombinant expression vectors" (or simply, "expression vectors").

[0083] "Operatively linked" expression control sequences refers to a linkage in which the expression control sequence is contiguous with the gene of interest to control the gene of interest, as well as expression control sequences that act in *trans* or at a distance to control the gene of interest.

[0084] The term "expression control sequence" as used herein refers to polynucleotide sequences which are necessary to affect the expression of coding sequences to which they are operatively linked. Expression control sequences are sequences which control the transcription, post-transcriptional events and translation of nucleic acid sequences. Expression control sequences include appropriate transcription initiation, termination, promoter and enhancer sequences; efficient RNA processing signals such as splicing and polyadenylation signals; sequences that stabilize cytoplasmic mRNA; sequences that enhance translation

efficiency (e.g., ribosome binding sites); sequences that enhance protein stability; and when desired, sequences that enhance protein secretion. The nature of such control sequences differs depending upon the host organism; in prokaryotes, such control sequences generally include promoter, ribosomal binding site, and transcription termination sequence. The term “control sequences” is intended to include, at a minimum, all components whose presence is essential for expression, and can also include additional components whose presence is advantageous, for example, leader sequences and fusion partner sequences.

5 [0085] The term “recombinant host cell” (or simply “host cell”), as used herein, is intended to refer to a cell into which a recombinant vector has been introduced. It should be understood that such terms are intended to refer not only to the particular subject cell but to the progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term “host cell” as used herein. A recombinant host cell may be an isolated cell or cell line grown in culture or may be a cell which resides in a living tissue or organism.

15 [0086] The term “peptide” as used herein refers to a short polypeptide, e.g., one that is typically less than about 50 amino acids long and more typically less than about 30 amino acids long. The term as used herein encompasses analogs and mimetics that mimic structural and thus biological function.

20 [0087] The term “polypeptide” encompasses both naturally-occurring and non-naturally-occurring proteins, and fragments, mutants, derivatives and analogs thereof. A polypeptide may be monomeric or polymeric. Further, a polypeptide may comprise a number of different domains each of which has one or more distinct activities.

25 [0088] The term “isolated protein” or “isolated polypeptide” is a protein or polypeptide that by virtue of its origin or source of derivation (1) is not associated with naturally associated components that accompany it in its native state, (2) when it exists in a purity not found in nature, where purity can be adjudged with respect to the presence of other cellular material (e.g., is free of other proteins from the same species) (3) is expressed by a cell from a different species, or (4) does not

occur in nature (e.g., it is a fragment of a polypeptide found in nature or it includes amino acid analogs or derivatives not found in nature or linkages other than standard peptide bonds). Thus, a polypeptide that is chemically synthesized or synthesized in a cellular system different from the cell from which it naturally
5 originates will be “isolated” from its naturally associated components. A polypeptide or protein may also be rendered substantially free of naturally associated components by isolation, using protein purification techniques well known in the art. As thus defined, “isolated” does not necessarily require that the protein, polypeptide, peptide or oligopeptide so described has been physically
10 removed from its native environment.

[0089] The term “polypeptide fragment” as used herein refers to a polypeptide that has an amino-terminal and/or carboxy-terminal deletion compared to a full-length polypeptide. In a preferred embodiment, the polypeptide fragment is a contiguous sequence in which the amino acid sequence of the fragment is identical
15 to the corresponding positions in the naturally-occurring sequence. Fragments typically are at least 5, 6, 7, 8, 9 or 10 amino acids long, preferably at least 12, 14, 16 or 18 amino acids long, more preferably at least 20 amino acids long, more preferably at least 25, 30, 35, 40 or 45, amino acids, even more preferably at least 50 or 60 amino acids long, and even more preferably at least 70 amino acids long.

20 [0090] A “modified derivative” refers to polypeptides or fragments thereof that are substantially homologous in primary structural sequence but which include, e.g., *in vivo* or *in vitro* chemical and biochemical modifications or which incorporate amino acids that are not found in the native polypeptide. Such modifications include, for example, acetylation, carboxylation, phosphorylation,
25 glycosylation, ubiquitination, labeling, e.g., with radionuclides, and various enzymatic modifications, as will be readily appreciated by those well skilled in the art. A variety of methods for labeling polypeptides and of substituents or labels useful for such purposes are well known in the art, and include radioactive isotopes such as ¹²⁵I, ³²P, ³⁵S, and ³H, ligands which bind to labeled antiligands (e.g.,
30 antibodies), fluorophores, chemiluminescent agents, enzymes, and antiligands which can serve as specific binding pair members for a labeled ligand. The choice of label depends on the sensitivity required, ease of conjugation with the primer,

stability requirements, and available instrumentation. Methods for labeling polypeptides are well known in the art. See Ausubel et al., 1992, hereby incorporated by reference.

[0091] The term "fusion protein" refers to a polypeptide comprising a polypeptide or fragment coupled to heterologous amino acid sequences. Fusion proteins are useful because they can be constructed to contain two or more desired functional elements from two or more different proteins. A fusion protein comprises at least 10 contiguous amino acids from a polypeptide of interest, more preferably at least 20 or 30 amino acids, even more preferably at least 40, 50 or 60 amino acids, yet more preferably at least 75, 100 or 125 amino acids. Fusion proteins can be produced recombinantly by constructing a nucleic acid sequence which encodes the polypeptide or a fragment thereof in frame with a nucleic acid sequence encoding a different protein or peptide and then expressing the fusion protein. Alternatively, a fusion protein can be produced chemically by crosslinking the polypeptide or a fragment thereof to another protein.

[0092] The term "non-peptide analog" refers to a compound with properties that are analogous to those of a reference polypeptide. A non-peptide compound may also be termed a "peptide mimetic" or a "peptidomimetic". See, e.g., Jones, (1992) *Amino Acid and Peptide Synthesis*, Oxford University Press; Jung, (1997) *Combinatorial Peptide and Nonpeptide Libraries: A Handbook* John Wiley; Bodanszky et al., (1993) *Peptide Chemistry--A Practical Textbook*, Springer Verlag; "Synthetic Peptides: A Users Guide", G. A. Grant, Ed, W. H. Freeman and Co., 1992; Evans et al. *J. Med. Chem.* 30:1229 (1987); Fauchere, *J. Adv. Drug Res.* 15:29 (1986); Veber and Freidinger *TINS* p.392 (1985); and references cited in each of the above, which are incorporated herein by reference. Such compounds are often developed with the aid of computerized molecular modeling. Peptide mimetics that are structurally similar to useful peptides of the invention may be used to produce an equivalent effect and are therefore envisioned to be part of the invention.

[0093] A "polypeptide mutant" or "mutein" refers to a polypeptide whose sequence contains an insertion, duplication, deletion, rearrangement or substitution of one or more amino acids compared to the amino acid sequence of a native or

wild type protein. A mutein may have one or more amino acid point substitutions, in which a single amino acid at a position has been changed to another amino acid, one or more insertions and/or deletions, in which one or more amino acids are inserted or deleted, respectively, in the sequence of the naturally-occurring protein, and/or truncations of the amino acid sequence at either or both the amino or carboxy termini. A mutein may have the same but preferably has a different biological activity compared to the naturally-occurring protein. For instance, a mutein may have an increased or decreased neuron or NgR binding activity. In a preferred embodiment of the present invention, a MAG derivative that is a mutein (e.g., in MAG Ig-like domain 5) has decreased neuronal growth inhibitory activity compared to endogenous or soluble wild-type MAG.

[0094] A mutein has at least 70% overall sequence homology to its wild-type counterpart. Even more preferred are muteins having 80%, 85% or 90% overall sequence homology to the wild-type protein. In an even more preferred embodiment, a mutein exhibits 95% sequence identity, even more preferably 97%, even more preferably 98% and even more preferably 99% overall sequence identity. Sequence homology may be measured by any common sequence analysis algorithm, such as Gap or Bestfit.

[0095] Preferred amino acid substitutions are those which: (1) reduce susceptibility to proteolysis, (2) reduce susceptibility to oxidation, (3) alter binding affinity for forming protein complexes, (4) alter binding affinity or enzymatic activity, and (5) confer or modify other physicochemical or functional properties of such analogs.

[0096] As used herein, the twenty conventional amino acids and their abbreviations follow conventional usage. See *Immunology - A Synthesis* (2nd Edition, E.S. Golub and D.R. Gren, Eds., Sinauer Associates, Sunderland, Mass. (1991)), which is incorporated herein by reference. Stereoisomers (e.g., D-amino acids) of the twenty conventional amino acids, unnatural amino acids such as α -, α -disubstituted amino acids, N-alkyl amino acids, and other unconventional amino acids may also be suitable components for polypeptides of the present invention. Examples of unconventional amino acids include: 4-hydroxyproline, γ -carboxyglutamate, ϵ -N,N,N-trimethyllysine, ϵ -N-acetyllysine, O-phosphoserine,

N-acetylserine, N-formylmethionine, 3-methylhistidine, 5-hydroxylysine, s-N-methylarginine, and other similar amino acids and imino acids (e.g., 4-hydroxyproline). In the polypeptide notation used herein, the left-hand direction is the amino terminal direction and the right hand direction is the carboxy-terminal direction, in accordance with standard usage and convention.

[0097] A protein has "homology" or is "homologous" to a second protein if the nucleic acid sequence that encodes the protein has a similar sequence to the nucleic acid sequence that encodes the second protein. Alternatively, a protein has homology to a second protein if the two proteins have "similar" amino acid sequences. (Thus, the term "homologous proteins" is defined to mean that the two proteins have similar amino acid sequences). In a preferred embodiment, a homologous protein is one that exhibits 60% sequence homology to the wild type protein, more preferred is 70% sequence homology. Even more preferred are homologous proteins that exhibit 80%, 85% or 90% sequence homology to the wild type protein. In a yet more preferred embodiment, a homologous protein exhibits 95%, 97%, 98% or 99% sequence identity. As used herein, homology between two regions of amino acid sequence (especially with respect to predicted structural similarities) is interpreted as implying similarity in function.

[0098] When "homologous" is used in reference to proteins or peptides, it is recognized that residue positions that are not identical often differ by conservative amino acid substitutions. A "conservative amino acid substitution" is one in which an amino acid residue is substituted by another amino acid residue having a side chain (R group) with similar chemical properties (e.g., charge or hydrophobicity). In general, a conservative amino acid substitution will not substantially change the functional properties of a protein. In cases where two or more amino acid sequences differ from each other by conservative substitutions, the percent sequence identity or degree of homology may be adjusted upwards to correct for the conservative nature of the substitution. Means for making this adjustment are well known to those of skill in the art (see, e.g., Pearson et al., 1994, herein incorporated by reference).

[0099] The following six groups each contain amino acids that are conservative substitutions for one another: 1) Serine (S), Threonine (T); 2) Aspartic Acid (D),

Glutamic Acid (E); 3) Asparagine (N), Glutamine (Q); 4) Arginine (R), Lysine (K); 5) Isoleucine (I), Leucine (L), Methionine (M), Alanine (A), Valine (V), and 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

[0100] Sequence homology for polypeptides, which is also referred to as percent
 5 sequence identity, is typically measured using sequence analysis software. See, e.g., the Sequence Analysis Software Package of the Genetics Computer Group (GCG), University of Wisconsin Biotechnology Center, 910 University Avenue, Madison, Wisconsin 53705. Protein analysis software matches similar sequences using measure of homology assigned to various substitutions, deletions and other
 10 modifications, including conservative amino acid substitutions. For instance, GCG contains programs such as "Gap" and "Bestfit" which can be used with default parameters to determine sequence homology or sequence identity between closely related polypeptides, such as homologous polypeptides from different species of organisms or between a wild type protein and a mutein thereof. See, e.g., GCG
 15 Version 6.1.

[0101] A preferred algorithm when comparing a inhibitory molecule sequence to a database containing a large number of sequences from different organisms is the computer program BLAST (Altschul, S.F. et al. (1990) *J. Mol. Biol.* 215:403-410; Gish and States (1993) *Nature Genet.* 3:266-272; Madden, T.L. et al. (1996) *Meth.*
 20 *Enzymol.* 266:131-141; Altschul, S.F. et al. (1997) *Nucleic Acids Res.* 25:3389-3402; Zhang, J. and Madden, T.L. (1997) *Genome Res.* 7:649-656), especially blastp or tblastn (Altschul et al., 1997). Preferred parameters for BLASTp are:

	Expectation value:	10 (default)
	Filter:	seg (default)
25	Cost to open a gap:	11 (default)
	Cost to extend a gap:	1 (default)
	Max. alignments:	100 (default)
	Word size:	11 (default)
	No. of descriptions:	100 (default)
30	Penalty Matrix:	BLOWSUM62

[0102] The length of polypeptide sequences compared for homology will generally be at least about 16 amino acid residues, usually at least about 20

residues, more usually at least about 24 residues, typically at least about 28 residues, and preferably more than about 35 residues. When searching a database containing sequences from a large number of different organisms, it is preferable to compare amino acid sequences. Database searching using amino acid sequences
5 can be measured by algorithms other than blastp known in the art. For instance, polypeptide sequences can be compared using FASTA, a program in GCG Version 6.1. FASTA provides alignments and percent sequence identity of the regions of the best overlap between the query and search sequences (Pearson, 1990, herein incorporated by reference). For example, percent sequence identity between amino
10 acid sequences can be determined using FASTA with its default parameters (a word size of 2 and the PAM250 scoring matrix), as provided in GCG Version 6.1, herein incorporated by reference.

[0103] "Specific binding" refers to the ability of two molecules to bind to each other in preference to binding to other molecules in the environment. Typically,
15 "specific binding" discriminates over adventitious binding in a reaction by at least two-fold, more typically by at least 10-fold, often at least 100-fold. Typically, the affinity or avidity of a specific binding reaction is at least about 10^{-7} M (e.g., at least about 10^{-8} M or 10^{-9} M).

[0104] The term "region" as used herein refers to a physically contiguous portion
20 of the primary structure of a biomolecule. In the case of proteins, a region is defined by a contiguous portion of the amino acid sequence of that protein.

[0105] The term "domain" as used herein refers to a structure of a biomolecule that contributes to a known or suspected function of the biomolecule. Domains may be co-extensive with regions or portions thereof; domains may also include
25 distinct, non-contiguous regions of a biomolecule. Examples of protein domains include, but are not limited to, an Ig domain, an extracellular domain, a transmembrane domain, and a cytoplasmic domain.

[0106] As used herein, the term "molecule" means any compound, including, but not limited to, a small molecule, peptide, protein, sugar, nucleotide, nucleic acid,
30 lipid, etc., and such a compound can be natural or synthetic.

[0107] Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art

to which this invention pertains. Exemplary methods and materials are described below, although methods and materials similar or equivalent to those described herein can also be used in the practice of the present invention and will be apparent to those of skill in the art. All publications and other references mentioned herein
5 are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. The materials, methods, and examples are illustrative only and not intended to be limiting.

[0108] Throughout this specification and claims, the word "comprise" or variations such as "comprises" or "comprising", will be understood to imply the
10 inclusion of a stated integer or group of integers but not the exclusion of any other integer or group of integers.

Engineering or Selecting Hosts With Modified Lipid-Linked Oligosaccharides For The Generation of Human-like N-Glycans

15 [0109] The invention provides a method for producing a human-like glycoprotein in a non-human eukaryotic host cell. The method involves making or using a non-human eukaryotic host cell diminished or depleted in an *alg* gene activity (i.e., *alg* activities, including equivalent enzymatic activities in non-fungal host cells) and introducing into the host cell at least one glycosidase activity. In a preferred
20 embodiment, the glycosidase activity is introduced by causing expression of one or more mannosidase activities within the host cell, for example, by activation of a mannosidase activity, or by expression from a nucleic acid molecule of a mannosidase activity, in the host cell.

[0110] In another embodiment, the method involves making or using a host cell
25 diminished or depleted in the activity of one or more enzymes that transfer a sugar residue to the 1,6 arm of lipid-linked oligosaccharide precursors (Fig. 1). A host cell of the invention is selected for or is engineered by introducing a mutation in one or more of the genes encoding an enzyme that transfers a sugar residue (e.g., mannosylates) the 1,6 arm of a lipid-linked oligosaccharide precursor. The sugar
30 residue is more preferably mannose, is preferably a glucose, GlcNAc, galactose, sialic acid, fucose or GlcNAc phosphate residue. In a preferred embodiment, the activity of one or more enzymes that mannosylate the 1,6 arm of lipid-linked

oligosaccharide precursors is diminished or depleted. The method may further comprise the step of introducing into the host cell at least one glycosidase activity (see below).

5 [0111] In yet another embodiment, the invention provides a method for producing a human-like glycoprotein in a non-human host, wherein the glycoprotein comprises an N-glycan having at least two GlcNAcs attached to a trimannose core structure.

[0112] In each above embodiment, the method is directed to making a host cell in which the lipid-linked oligosaccharide precursors are enriched in $\text{Man}_X\text{GlcNAc}_2$ structures, where X is 3, 4 or 5 (Fig. 2). These structures are transferred in the ER of the host cell onto nascent polypeptide chains by an oligosaccharyl-transferase and may then be processed by treatment with glycosidases (e.g., α -mannosidases) and glycosyltransferases (e.g., GnT1) to produce N-glycans having $\text{GlcNAcMan}_X\text{GlcNAc}_2$ core structures, wherein X is 3, 4 or 5, and is preferably 3 (Figs. 2 and 3). As shown in Fig. 2, N-glycans having a $\text{GlcNAcMan}_X\text{GlcNAc}_2$ core structure where X is greater than 3 may be converted to $\text{GlcNAcMan}_3\text{GlcNAc}_2$, e.g., by treatment with an α -1,3 and/or α -1,2-1,3 mannosidase activity, where applicable.

[0113] Additional processing of $\text{GlcNAcMan}_3\text{GlcNAc}_2$ by treatment with glycosyltransferases (e.g., GnTII) produces $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$ core structures which may then be modified, as desired, e.g., by *ex vivo* treatment or by heterologous expression in the host cell of a set of glycosylation enzymes, including glycosyltransferases, sugar transporters and mannosidases (see below), to become human-like N-glycans. Preferred human-like glycoproteins which may be produced according to the invention include those which comprise N-glycans having seven or fewer, or three or fewer, mannose residues; comprise one or more sugars selected from the group consisting of galactose, GlcNAc, sialic acid, and fucose; and comprise at least one oligosaccharide branch comprising the structure NeuNAc-Gal-GlcNAc-Man.

30 [0114] In one embodiment, the host cell has diminished or depleted Dol-P-Man:Man₅GlcNAc₂-PP-Dol Mannosyltransferase activity, which is an activity involved in the first mannosylation step from Man₅GlcNAc₂-PP-Dol to

Man₆GlcNAc₂-PP-Dol at the luminal side of the ER (e.g., *ALG3* Fig. 1; Fig. 2). In *S.cerevisiae*, this enzyme is encoded by the *ALG3* gene. As described above, *S.cerevisiae* cells harboring a leaky *alg3-1* mutation accumulate Man₅GlcNAc₂-PP-Dol and cells having a deletion in *alg3* appear to transfer Man₅GlcNAc₂ structures onto nascent polypeptide chains within the ER. Accordingly, in this embodiment, host cells will accumulate N-glycans enriched in Man₅GlcNAc₂ structures which can then be converted to GlcNAc₂Man₃GlcNAc₂ by treatment with glycosidases (e.g., with α -1,2 mannosidase, α -1,3 mannosidase or α -1,2-1,3 mannosidase activities (Fig. 2).

10 [0115] As described in Example 1, degenerate primers were designed based on an alignment of Alg3 protein sequences from *S. cerevisiae*, *D. melanogaster* and humans (*H. sapiens*) (Figs. 4 and 5), and were used to amplify a product from *P. pastoris* genomic DNA. The resulting PCR product was used as a probe to identify and isolate a *P. pastoris* genomic clone comprising an open reading frame (ORF) that encodes a protein having 35% overall sequence identity and 53% sequence similarity to the *S. cerevisiae* *ALG3* gene (Figs. 6 and 7). This *P. pastoris* gene is referred to herein as "*PpALG3*". The *ALG3* gene was similarly identified and isolated from *K. lactis* (Example 1; Figs. 8 and 9).

20 [0116] Thus, in another embodiment, the invention provides an isolated nucleic acid molecule having a nucleic acid sequence comprising or consisting of at least forty-five, preferably at least 50, more preferably at least 60 and most preferably 75 or more nucleotide residues of the *P.pastoris* *ALG 3* gene (Fig. 6) and the *K. lactis* *ALG 3* gene (Fig. 8), and homologs, variants and derivatives thereof. The invention also provides nucleic acid molecules that hybridize under stringent conditions to the above-described nucleic acid molecules. Similarly, isolated polypeptides (including muteins, allelic variants, fragments, derivatives, and analogs) encoded by the nucleic acid molecules of the invention are provided (25 *P.pastoris* and *K. lactis* *ALG 3* gene products are shown in Fig. 6 and 8). In addition, also provided are vectors, including expression vectors, which comprise a nucleic acid molecule of the invention, as described further herein.

30 [0117] Using gene-specific primers, a construct was made to delete the *PpALG3* gene from the genome of *P. pastoris* (Example 1). This strain was used to

generate a host cell depleted in Dol-P-Man:Man₅GlcNAc₂-PP-Dol
Mannosyltransferase activity and produce lipid-linked Man₅GlcNAc₂-PP-Dol
precursors which are transferred onto nascent polypeptide chains to produce N-
glycans having a Man₅GlcNAc₂ carbohydrate structure.

5 [0118] As described in **Example 2**, such a host cell may be engineered by
expression of appropriate mannosidases to produce N-glycans having the desired
Man₃GlcNAc₂ core carbohydrate structure. Expression of GnTs in the host cell
(e.g., by targeting a nucleic acid molecule or a library of nucleic acid molecules as
described below) enables the modified host cell to produce N-glycans having one
10 or two GlcNAc structures attached to each arm of the Man₃ core structure (i.e.,
GlcNAc₁Man₃GlcNAc₂ or GlcNAc₂Man₃GlcNAc₂; see **Fig. 3**). These structures
may be processed further using the methods of the invention to produce human-
like N-glycans on proteins which enter the secretion pathway of the host cell.

[0119] In another embodiment, the host cell has diminished or depleted dolichyl-
15 P-Man:Man₆GlcNAc₂-PP-dolichyl α -1,2 mannosyltransferase activity, which is an
 α -1,2 mannosyltransferase activity involved in the mannosylation step converting
Man₆GlcNAc₂-PP-Dol to Man₇GlcNAc₂-PP-Dol at the luminal side of the ER (see
above and **Figs. 1 and 2**). In *S.cerevisiae*, this enzyme is encoded by the *ALG9*
gene. Cells harboring an *alg9* mutation accumulate Man₆GlcNAc₂-PP-Dol (**Fig. 2**)
20 and transfer Man₆GlcNAc₂ structures onto nascent polypeptide chains within the
ER. Accordingly, in this embodiment, host cells will accumulate N-glycans
enriched in Man₆GlcNAc₂ structures which can then be processed down to core
Man₃ structures by treatment with α -1,2 and α -1,3 mannosidases (see **Fig. 3 and**
Examples 3 and 4).

25 [0120] A host cell in which the *alg9* gene (or gene encoding an equivalent
activity) has been deleted is constructed (see, e.g., **Example 3**). Deletion of *ALG9*
(or *ALG12*; see below) creates a host cell which produces N-glycans with one or
two additional mannoses, respectively, on the 1,6 arm (**Fig. 2**). In order to make
the 1,6 core-mannose accessible to N-acetylglucosaminyltransferase II (GnTII)
30 these mannoses have to be removed by glycosidase(s). ER mannosidase typically
will remove the terminal 1,2 mannose on the 1,6 arm and subsequently
Mannosidase II (alpha 1-3,6 mannosidase) or other mannosidases such as alpha

1,2, α 1,3 or α 1-2,3 mannosidases (e.g., from *Xanthomonas manihotis*; see **Example 4**) can act upon the 1,6 arm and subsequently GnTII can transfer an N-acetylglucosamine, resulting in GlcNAc₂Man₃ (**Fig. 2**).

5 [0121] The resulting host cell, which is depleted for alg9p activity, is engineered to express α -1,2 and α -1,3 mannosidase activity (from one or more enzymes, and preferably, by expression from a nucleic acid molecule introduced into the host cell and which expresses an enzyme targeted to a preferred subcellular compartment (see below). **Example 4** describes the cloning and expression of one such enzyme from *Xanthomonas manihotis*.

10 [0122] In another embodiment, the host cell has diminished or depleted dolichyl-P-Man:Man₇GlcNAc₂-PP-dolichyl α -1,6 mannosyltransferase activity, which is an α -1,6 mannosyltransferase activity involved in the mannosylation step converting Man₇GlcNAc₂-PP-Dol to Man₈GlcNAc₂-PP-Dol (which mannosylates the α -1,6 mannose on the 1,6 arm of the core mannose structure) at the luminal side of the
15 ER (see above and **Figs. 1 and 2**). In *S.cerevisiae*, this enzyme is encoded by the *ALG12* gene. Cells harboring an *alg12* mutation accumulate Man₇GlcNAc₂-PP-Dol (**Fig. 2**) and transfer Man₇GlcNAc₂ structures onto nascent polypeptide chains within the ER. Accordingly, in this embodiment, host cells will accumulate N-glycans enriched in Man₇GlcNAc₂ structures which can then be processed down to
20 core Man₃ structures by treatment with α -1,2 and α -1,3 mannosidases (see **Fig. 3 and Examples 3 and 4**).

[0123] As described above for alg9 mutant hosts, the resulting host cell, which is depleted for alg12p activity, is engineered to express α -1,2 and α -1,3 mannosidase activity (e.g., from one or more enzymes, and preferably, by expression from one
25 or more nucleic acid molecules introduced into the host cell and which express an enzyme activity which is targeted to a preferred subcellular compartment (see below).

[0124]

30 **Engineering or Selecting Hosts Optionally Having Decreased Initiating α -1,6 Mannosyltransferase Activity**

[0125] In a preferred embodiment, the method of the invention involves making or using a host cell which is both (a) diminished or depleted in the activity of an

alg gene or in one or more activities that mannosylate N-glycans on the α -1,6 arm of the $\text{Man}_3\text{GlcNAc}_2$ ("Man3") core carbohydrate structure; and (b) diminished or depleted in the activity of an initiating α -1,6-mannosyltransferase, i.e., an initiation specific enzyme that initiates outer chain mannosylation (on the α -1,3 arm of the Man3 cores structure). In *S.cerevisiae*, this enzyme is encoded by the *OCH1* gene. Disruption of the *och1* gene in *S.cerevisiae* results in a phenotype in which N-linked sugars completely lack the poly-mannose outer chain. Previous approaches for obtaining mammalian-type glycosylation in fungal strains have required inactivation of *OCH1* (see, e.g., Chiba, 1998). Disruption of the initiating α -1,6-mannosyltransferase activity in a host cell of the invention is optional, however (depending on the selected host cell), as the Och1p enzyme requires an intact $\text{Man}_8\text{GlcNAc}$ for efficient mannose outer chain initiation. Thus, the host cells selected or produced according to this invention, which accumulate lipid-linked oligosaccharides having seven or fewer mannose residues will, after transfer, produce hypoglycosylated N-glycans that will likely be poor substrates for Och1p (see, e.g., Nakayama, 1997).

Engineering or Selecting Hosts Having Increased Glucosyltransferase Activity

[0126] As discussed above, glucosylated oligosaccharides are thought to be transferred to nascent polypeptide chains at a much higher rate than their nonglucosylated counterparts. It appears that substrate recognition by the oligosaccharyltransferase complex is enhanced by addition of glucose to the antennae of lipid-linked oligosaccharides. It is thus desirable to create or select host cells capable of optimal glucosylation of the lipid-linked oligosaccharides. In such host cells, underglycosylation will be substantially decreased or even abolished, due to a faster and more efficient transfer of glucosylated Man_5 structures onto the nascent polypeptide chain.

[0127] Accordingly, in another embodiment of the invention, the method is directed to making a host cell in which the lipid-linked N-glycan precursors are transferred efficiently to the nascent polypeptide chain in the ER. In a preferred embodiment, transfer is augmented by increasing the level of glucosylation on the

branches of lipid-linked oligosaccharides which, in turn, will make them better substrates for oligosaccharyltransferase.

[0128] In one preferred embodiment, the invention provides a method for making a human-like glycoprotein which uses a host cell in which one or more enzymes responsible for glucosylation of lipid-linked oligosaccharides in the ER has increased activity. One way to enhance the degree of glucosylation of the lipid-linked oligosaccharides is to overexpress one or more enzymes responsible for the transfer of glucose residues onto the antennae of the lipid-linked oligosaccharide. In particular, increasing α -1,3 glucosyltransferase activity will increase the amount of glucosylated lipid-linked Man₅ structures and will reduce or eliminate the underglycosylation of secreted proteins. In *S.cerevisiae*, this enzyme is encoded by the *ALG6* gene.

[0129] *Saccharomyces cerevisiae* *ALG6* and its human counterpart have been cloned (Imbach, 1999; Reiss, 1996). Due to the evolutionary conservation of the early steps of glycosylation, *ALG6* loci are expected to be homologous between species and may be cloned based on sequence similarities by anyone skilled in the art. (The same holds true for cloning and identification of *ALG8* and *ALG10* loci from different species.) In addition, different glucosyltransferases from different species can then be tested to identify the ones with optimal activities.

[0130] The introduction of additional copies of an *ALG6* gene and/or the expression of *ALG6* under the control of a strong promoter, such as the GAPDH promoter, is one of several ways to increase the degree of glucosylated lipid-linked oligosaccharides. The *ALG6* gene from *P. pastoris* is cloned and expressed (Example 5). *ALG6* nucleic acid and amino acid sequences are shown in Fig. 25 (*S. cerevisiae*) and Fig. 26 (*P. pastoris*). These sequences are compared to other eukaryotic *ALG6* sequences in Fig. 27.

[0131] Accordingly, another embodiment of the invention provides a method to enhance the degree of glucosylation of lipid-linked oligosaccharides comprising the step of increasing α -1,3 glucosyltransferase activity in a host cell. The increase in activity may be achieved by overexpression of nucleic acid sequences encoding the activity, e.g., by operatively linking the nucleic acid encoding the activity with one or more heterologous expression control sequences. Preferred

expression control sequences include transcription initiation, termination, promoter and enhancer sequences; RNA splice donor and polyadenylation signals; mRNA stabilizing sequences; ribosome binding sites; protein stabilizing sequences; and protein secretion sequences.

- 5 **[0132]** In another embodiment, the increase in alpha-1,3 glucosyltransferase activity is achieved by introducing a nucleic acid molecule encoding the activity on a multi-copy plasmid, using techniques well known to the skilled worker. In yet another embodiment, the degree of glucosylation of lipid-linked oligosaccharides comprising decreasing the substrate specificity of oligosaccharyl transferase
- 10 activity in a host cell. This is achieved by, for example, subjecting at least one nucleic acid encoding the activity to a technique such as gene shuffling, *in vitro* mutagenesis, and error-prone polymerase chain reaction, all of which are well-known to one of skill in the art. Naturally, *ALG8* and *ALG10* can be overexpressed in a host cell and tested in a similar fashion.
- 15 **[0133]** Accordingly, in a preferred embodiment, the invention provides a method for making a human-like glycoprotein using a host cell which is engineered or selected so that one or more enzymes responsible for glucosylation of lipid-linked oligosaccharides in the ER has increased activity. In a more preferred
- 20 embodiment, the invention uses a host cell having both (a) diminished or depleted in the activity of one or more alg gene activities or activities that mannosylate N-glycans on the α -1,6 arm of the $\text{Man}_3\text{GlcNAc}_2$ ("Man3") core carbohydrate structure and (b) engineered or selected so that one or more enzymes responsible for glucosylation of lipid-linked oligosaccharides in the ER has increased activity. The lipid-linked Man_5 structure found in an *alg3* mutant background, however, is
- 25 not a preferred substrate for Alg6p. Accordingly, the skilled worker may identify Alg6p, Alg8p and Alg10p with an increased substrate specificity (Gibbs, 2001) e.g., by subjecting nucleic acids encoding such enzymes to one or more rounds of gene shuffling, error prone PCR, or *in vitro* mutagenesis approaches and selecting for increased substrate specificity in a host cell of interest, using molecular biology
- 30 and genetic selection techniques well known to those of skill in the art. It will be appreciated by the skilled worker that such techniques for improving enzyme substrate specificities in a selected host strain are not limited to this particular

embodiment of the invention but rather, may be used in any embodiment to optimize further the production of human-like N-glycans in a non-human host cell.

[0134] As described, once Man₅ is transferred onto the nascent polypeptide chain, expression of suitable α -1,2-mannosidase(s), as provided by the present invention, will further trim Man₅GlcNAc₂ structures to yield the desired core Man₃GlcNAc₂ structures. α -1,2-mannosidases remove only terminal α -1,2-linked mannose residues and are expected to recognize the Man₅GlcNAc₂ – Man₇GlcNAc₂ specific structures made in *alg3*, *9* and *12* mutant host cells and in host cells in which homologs to these genes are mutated.

10 [0135] As schematically presented in Figure 3, co-expression of appropriate UDP-sugar-transporter(s) and –transferase(s) will cap the terminal α -1,6 and α -1,3 residues with GlcNAc, resulting in the necessary precursor for mammalian-type complex and hybrid N-glycosylation: GlcNAc₂Man₅GlcNAc₂. The peptide-bound N-linked oligosaccharide chain GlcNAc₂Man₃GlcNAc₂ (Figure 3) then serves as a precursor for further modification to a mammalian-type oligosaccharide structure. Subsequent expression of galactosyl-transferases and genetically engineering the capacity to transfer sialic acid will produce a mammalian-type (e.g., human-like) N-glycan structure.

20 [0136] A desired host cell according to the invention can be engineered one enzyme or more than one enzyme at a time. In addition, a library of genes encoding potentially useful enzymes can be created, and a strain having one or more enzymes with optimal activities or producing the most “human-like” glycoproteins, selected by transforming target host cells with one or more members of the library. Lower eukaryotes that are able to produce glycoproteins having the core N-glycan Man₃GlcNAc₂ are particularly useful because of the ease of performing genetic manipulations, and safety and efficiency features. In a preferred embodiment, at least one further glycosylation reaction is performed, *ex vivo* or *in vivo*, to produce a human-like N-glycan. In a more preferred embodiment, active forms of glycosylating enzymes are expressed in the endoplasmic reticulum and/or Golgi apparatus of the host cell to produce the desired human-like glycoprotein.

30

Host Cells

[0137] A preferred non-human host cell of the invention is a lower eukaryotic cell, e.g., a unicellular or filamentous fungus, which is diminished or depleted in the activity of one or more *alg* gene activities (including an enzymatic activity which is a homolog or equivalent to an *alg* activity). Another preferred host cell of the invention is diminished or depleted in the activity of one or more enzymes (other than *alg* activities) that mannosylate the α -1,6 arm of a lipid-linked oligosaccharide structure.

[0138] While lower eukaryotic host cells are preferred, a wide variety of host cells having the aforementioned properties are envisioned as being useful in the methods of the invention. Plant cells, for instance, may be engineered to express a human-like glycoprotein according to the invention. Likewise, a variety of non-human, mammalian host cells may be altered to express more human-like glycoproteins using the methods of the invention. An appropriate host cell can be engineered, or one of the many such mutants already described in yeasts may be used. A preferred host cell of the invention, as exemplified herein, is a hypermannosylation-minus (*OCH1*) mutant in *Pichia pastoris* which has further been modified to delete the *alg3* gene. Other preferred hosts are *Pichia pastoris* mutants having *och1* and *alg 9* or *alg12* mutations.

Formation of complex N-glycans

[0139] The sequential addition of sugars to the modified, nascent N-glycan structure involves the successful targeting of glucosyltransferases into the Golgi apparatus and their successful expression. This process requires the functional expression, e.g., of GnT I, in the early or medial Golgi apparatus as well as ensuring a sufficient supply of UDP-GlcNAc (e.g., by expression of a UDP-GlcNAc transporter).

[0140] To characterize the glycoproteins and to confirm the desired glycosylation, the glycoproteins were purified, the N-glycans were PNGase-F released and then analyzed by MALDI-TOF-MS (Example 2). Kringle 3 domain

of human plasminogen was used as the reporter protein. This soluble glycoprotein was produced in *P. pastoris* in an *alg3*, *och1* knockout background (**Example 2**).

[0141] GlcNAcMan₅GlcNAc₂ was produced as the predominant N-glycan after addition of human GnT I, and *K. lactis* UDP-GlcNAc transporter in **Fig. 16**

5 (**Example 2**). The mass of this N-glycan is consistent with the mass of GlcNAcMan₅GlcNAc₂ at 1463 (m/z). To confirm the addition of the GlcNAc onto Man₅GlcNAc₂, a β -N-hexosaminidase digest was performed, which revealed a peak at 1260 (m/z), consistent with the mass of Man₅GlcNAc₂ (**Fig. 17**).

[0142] The N-glycans from the *alg3 och1* deletion in one strain PBP3 (**Example 2**) provided two distinct peaks at 1138 (m/z) and 1300 (m/z), which is consistent with structures GlcNAcMan₃GlcNAc₂ and GlcNAcMan₄GlcNAc₂ (**Fig. 18**). After an *in vitro* α 1,2-mannosidase digestion for redundant mannoses, a peak eluted at 1138 (m/z), which is consistent with GlcNAcMan₃GlcNAc₂ (**Fig. 19**). To confirm the addition of the GlcNAc onto the Man₃GlcNAc₂ structure, a β -N-
15 hexosaminidase digest was performed, which revealed a peak at 934 (m/z), consistent with the mass of Man₃GlcNAc₂ (**Fig. 20**).

[0143] The addition of the second GlcNAc onto GlcNAcMan₃GlcNAc₂ is shown in **Fig. 21**. The peak at 1357 (m/z) corresponds to GlcNAc₂Man₃GlcNAc₂. To confirm the addition of the two GlcNAcs onto the core mannose structure
20 Man₃GlcNAc₂, another β -N-hexosaminidase digest was performed, which revealed a peak at 934 (m/z), consistent with the mass of Man₃GlcNAc₂ (**Fig. 22**). This is conclusive data displaying a complex-type glycoprotein made in yeast cells.

[0144] The *in vitro* addition of UDP-galactose and β 1,4-galactosyltransferase onto the GlcNAc₂Man₃GlcNAc₂ resulted in a peak at 1664 (m/z), which is
25 consistent with the mass of Gal₂GlcNAc₂Man₃GlcNAc₂ (**Fig. 23**). Finally, the *in vitro* addition of CMP-N-acetylneuraminic acid and sialyltransferase resulted in a peak at 2248 (m/z), which is consistent with the mass of
NANA₂Gal₂GlcNAc₂Man₃GlcNAc₂ (**Fig. 24**). The above data supports the use of non-mammalian host cells, which are capable of producing complex human-like
30 glycoproteins.

Targeting of glycosyl- and galactosyl-transferases to specific organelles.

[0145] Much work has been dedicated to revealing the exact mechanism by which these enzymes are retained and anchored to their respective organelle.

Although complex, evidence suggests that, stem region, membrane spanning
5 region and cytoplasmic tail individually or in concert direct enzymes to the membrane of individual organelles and thereby localize the associated catalytic domain to that locus.

[0146] The method by which active glycosyltransferases can be expressed and directed to the appropriate organelle such that a sequential order of reactions may
10 occur, that leads to complex N-glycan formation, is as follows:

(A) Establish a DNA library of regions that are known to encode proteins/peptides that mediate localization to a particular location in the secretory pathway (ER, Golgi and trans Golgi network). A limited selection of such enzymes and their respective location is shown in Table 1. These sequences may be selected from
15 the host to be engineered as well as other related or unrelated organism. Generally such sequences fall into three categories: (1) N-terminal sequences encoding a cytosolic tail (ct), a transmembrane domain (tmd) and part of a somewhat more ambiguously defined stem region (sr), which together or individually anchor proteins to the inner (luminal) membrane of the Golgi, (2) retrieval signals which
20 are generally found at the C-terminus such as the HDEL or KDEL tetrapeptide, and (3) membrane spanning nucleotide sugar transporters, which are known to locate in the Golgi. In the first case, where the localization region consists of various elements (ct, tmd and sr) the library is designed such that the ct, the tmd and various parts of the stem region are represented. This may be accomplished by
25 using PCR primers that bind to the 5' end of the DNA encoding the cytosolic region and employing a series of opposing primers that bind to various parts of the stem region. In addition one would create fusion protein constructs that encode sugar nucleotide transporters and known retrieval signals.

(B) A second step involves the creation of a series of fusion protein constructs,
30 that encode the above mentioned localization sequences and the catalytic domain of a particular glycosyltransferase cloned in frame to such localization sequence (e.g. GnT I, GalT, Fucosyltransferase or ST). In the case of a sugar nucleotide

transporter fused to a catalytic domain one may design such constructs such that the catalytic domain (e.g. GnT I) is either at the N- or the C-terminus of the resulting polypeptide. The catalytic domain, like the localization sequence, may be derived from various different sources. The choice of such a catalytic domains
5 may be guided by the knowledge of the particular environment in which the catalytic domain is to be active. For example, if a particular glycosyltransferase is to be active in the late Golgi, and all known enzymes of the host organism in the late Golgi have a pH optimum of 7.0, or the late Golgi is known to have a particular pH, one would try to select a catalytic domain that has maximum activity
10 at that pH. Existing *in vivo* data on the activity of such enzymes, in particular hosts, may also be of use. For example, Schwientek and coworkers showed that GalT activity can be engineered into the Golgi of *S.cerevisiae* and showed that such activity was present by demonstrating the transfer of some Gal to existing GlcNAc₂ in an *alg* mutant of *S. cerevisiae*. In addition, one may perform several
15 rounds of gene shuffling or error prone PCR to obtain a larger diversity within the pool of fusion constructs, since it has been shown that single amino mutations may drastically alter the activity of glycoprotein processing enzymes (Romero et al., 2000). Full length sequences of glycosyltransferases and their endogenous anchoring sequence may also be used. In a preferred embodiment, such
20 localization/catalytic domain libraries are designed to incorporate existing information on the sequential nature of glycosylation reactions in higher eukaryotes. In other words, reactions known to occur early in the course of glycoprotein processing require the targeting of enzymes that catalyze such reactions to an early part of the Golgi or the ER. For example, the trimming of
25 Man₈GlcNAc₂ to Man₅GlcNAc₂ is an early step in complex N-glycan formation. Since protein processing is initiated in the ER and then proceeds through the early, medial and late Golgi, it is desirable to have this reaction occur in the ER or early Golgi. When designing a library for mannosidase I localization, one thus attempts to match ER and early Golgi targeting signals with the catalytic domain of
30 mannosidase I.

[0147] Upon transformation of the host strain with the fusion construct library a selection process is used to identify which particular combination of localization

sequence and catalytic domain in fact have the maximum effect on the carbohydrate structure found in such host strain. Such selection can be based on any number of assays or detection methods. They may be carried out manually or may be automated through the use of high throughput screening equipment.

5 [0148] In another example, GnT I activity is required for the maturation of complex N-glycans, because only after addition of GlcNAc to the terminal α 1,3 mannose residue may further trimming of such a structure to the subsequent intermediate GlcNAcMan₃GlcNAc₂ structure occur. Mannosidase II is most likely not capable of removing the terminal α 1,3- and α 1,6- mannose residues in the
10 absence of a terminal β 1,2-GlcNAc and thus the formation of complex N-glycans will not proceed in the absence of GnT I activity (Schachter, 1991). Alternatively, one may first engineer or select a strain that makes sufficient quantities of Man₅GlcNAc₂ as described in this invention by engineering or selecting a strain deficient in Alg3P activity. In the presence of sufficient UDP-GlcNAc transporter
15 activity, as may be achieved by engineering or selecting a strain that has such UDP-GlcNAc transporter activity, GlcNAc can be added to the terminal α -1,3 residue by GnTI as *in vitro* a Man₃ structure is recognized by by rat liver GnTI (Moller, 1992).

[0149] In another approach, one may incorporate the expression of a UDP-
20 GlcNAc transporter into the library mentioned above such that the desired construct will contain: (1) a region by which the transformed construct is maintained in the cell (e.g. origin of replication or a region that mediates chromosomal integration), (2) a marker gene that allows for the selection of cells that have been transformed, including counterselectable and recyclable markers
25 such as *ura3* or *T-urf13* (Soderholm, 2001) or other well characterized selection-markers (e.g. *his4*, *bla*, *Sh ble* etc.), (3) a gene encoding a UDP-GlcNAc transporter (e.g. from *K.lactis*, (Abeijon, 1996), or from *H.sapiens* (Ishida, 1996), and (4) a promotor activating the expression of the above mentioned localization/catalytic domain fusion construct library.

30 [0150] After transformation of the host with the library of fusion constructs described above, one may screen for those cells that have the highest concentration of terminal GlcNAc on the cell surface, or secrete the protein with the highest

terminal GlcNAc content. Such a screen may be based on a visual method, like a staining procedure, the ability to bind specific terminal GlcNAc binding antibodies or lectins conjugated to a marker (such lectins are available from E.Y. Laboratories Inc., San Mateo, CA), the reduced ability of specific lectins to bind to terminal
5 mannose residues, the ability to incorporate a radioactively labeled sugar *in vitro*, altered binding to dyes or charged surfaces, or may be accomplished by using a Fluorescence Assisted Cell Sorting (FACS) device in conjunction with a fluorophore labeled lectin or antibody (Guillen, 1998). It may be advantageous to enrich particular phenotypes within the transformed population with cytotoxic
10 lectins. U.S. Patent No. 5,595,900 teaches several methods by which cells with a desired extra-cellular carbohydrate structures may be identified. Repeatedly carrying out this strategy allows for the sequential engineering of more and more complex glycans in lower eukaryotes.

[0151] After transformation, one may select for transformants that allow for the
15 most efficient transfer of GlcNAc by GlcNAc Transferase II from UDP-GlcNAc in an *in vitro* assay. This screen may be carried out by growing cells harboring the transformed library under selective pressure on an agar plate and transferring individual colonies into a 96-well microtiter plate. After growing the cells, the cells are centrifuged, the cells resuspended in buffer, and after addition of UDP-
20 GlcNAc and GnT V, the release of UDP is determined either by HPLC or an enzyme linked assay for UDP. Alternatively, one may use radioactively labeled UDP-GlcNAc and GnT V, wash the cells and then look for the release of radioactive GlcNAc by N-actylglucosaminidase. All this may be carried manually or automated through the use of high throughput screening equipment.

25 [0152] Transformants that release more UDP, in the first assay, or more radioactively labeled GlcNAc in the second assay, are expected to have a higher degree of GlcNAcMan₃GlcNAc₂ (Fig. 3) on their surface and thus constitute the desired phenotype. Alternatively, one may any use any other suitable screen such as a lectin binding assay that is able to reveal altered glycosylation patterns on the
30 surface of transformed cells. In this case the reduced binding of lectins specific to terminal mannoses may be a suitable selection tool. *Galantus nivalis* lectin binds specifically to terminal α -1,3 mannose, which is expected to be reduced if

sufficient mannosidase II activity is present in the Golgi. One may also enrich for desired transformants by carrying out a chromatographic separation step that allows for the removal of cells containing a high terminal mannose content. This separation step would be carried out with a lectin column that specifically binds

5 cells with a high terminal mannose content (e.g. *Galantus nivalis* lectin bound to agarose, Sigma, St. Louis, MO) over those that have a low terminal mannose content. In addition, one may directly create such fusion protein constructs, as additional information on the localization of active carbohydrate modifying enzymes in different lower eukaryotic hosts becomes available in the scientific

10 literature. For example, the prior art teaches us that human β 1,4-GalTr can be fused to the membrane domain of MNT, a mannosyltransferase from *S. cerevisiae*, and localized to the Golgi apparatus while retaining its catalytic activity (Schwientek et al., 1995). If *S. cerevisiae* or a related organism is the host to be engineered one may directly incorporate such findings into the overall strategy to

15 obtain complex N-glycans from such a host. Several such gene fragments in *P. pastoris* have been identified that are related to glycosyltransferases in *S. cerevisiae* and thus could be used for that purpose.

Table 1

<u>Gene or sequence</u>	<u>Organism</u>	<u>Function</u>	<u>Location of gene product</u>
MnsI	<i>S. cerevisiae</i>	mannosidase	ER
Och1	<i>S. cerevisiae</i>	1,6-mannosyltransferase	Golgi (cis)
Mnn2	<i>S. cerevisiae</i>	1,2-mannosyltransferase	Golgi (medial)
Mnn1	<i>S. cerevisiae</i>	1,3-mannosyltransferase	Golgi (trans)
Och1	<i>P. pastoris</i>	1,6-mannosyltransferase	Golgi (cis)
2,6 ST	<i>H. sapiens</i> <i>S. frugiperda</i>	2,6-sialyltransferase	trans-Golgi network
β 1,4 Gal T	<i>bovine milk</i>	UDP-Gal transporter	Golgi
Mnt1	<i>S. cerevisiae</i>	1,2-mannosyltransferase	Golgi (cis)
HDEL at C-terminus	<i>S. cerevisiae</i>	retrieval signal	ER

Integration Sites

[0153] As one ultimate goal of this genetic engineering effort is a robust protein production strain that is able to perform well in an industrial fermentation process, the integration of multiple genes into the host (e.g., fungal) chromosome involves careful planning. The engineered strain will most likely have to be transformed with a range of different genes, and these genes will have to be transformed in a stable fashion to ensure that the desired activity is maintained throughout the fermentation process. Any combination of the following enzyme activities will have to be engineered into the fungal protein expression host: sialyltransferases, mannosidases, fucosyltransferases, galactosyltransferases, glucosyltransferases, GlcNAc transferases, ER and Golgi specific transporters (e.g. syn and antiport transporters for UDP-galactose and other precursors), other enzymes involved in the processing of oligosaccharides, and enzymes involved in the synthesis of activated oligosaccharide precursors such as UDP-galactose, CMP-N-acetylneuraminic acid. At the same time, a number of genes which encode enzymes known to be characteristic of non-human glycosylation reactions, will have to be deleted. Such genes and their corresponding proteins have been extensively characterized in a number of lower eukaryotes (e.g. *S.cerevisiae*, *T.reesei*, *A. nidulans* etc.), thereby providing a list of known glycosyltransferases in lower eukaryotes, their activities and their respective genetic sequence. These genes are likely to be selected from the group of mannosyltransferases e.g. 1,3 mannosyltransferases (e.g. MNN1 in *S.cerevisiae*) (Graham, 1991), 1,2 mannosyltransferases (e.g. KTR/KRE family from *S.cerevisiae*), 1,6 mannosyltransferases (OCH1 from *S.cerevisiae*), mannosylphosphate transferases (MNN4 and MNN6 from *S.cerevisiae*) and additional enzymes that are involved in aberrant i.e. non human glycosylation reactions. Many of these genes have in fact been deleted individually giving rise to viable phenotypes with altered glycosylation profiles. Examples are shown in Table 2:

Table 2.

<u>Strain</u>	<u>Mutant</u>	<u>Structure wild type</u>	<u>Structure mutant</u>	<u>Authors</u>
<i>Schizosaccharomyces pombe</i>	OCH1	Mannan (i.e. Man ₃₋₉ GlcNAc ₂)	Man ₈ GlcNAc ₂	Yoko-o et al., 2001

<i>S.cerevisiae</i>	<i>OCH1</i> , <i>MNN1</i>	Mannan (i.e. Man ₈₋₉ GlcNAc ₂)	Man ₈ GlcNAc ₂	Nakanishi-Shindo et al., 1993
<i>S.cerevisiae</i>	<i>OCH1</i> , <i>MNN1</i> , <i>MNN4</i>	Mannan (i.e. Man ₈₋₉ GlcNAc ₂)	Man ₈ GlcNAc ₂	Chiba et al., 1998

As any strategy to engineer the formation of complex N-glycans into a lower eukaryote involves both the elimination as well as the addition of glycosyltransferase activities, a comprehensive scheme will attempt to coordinate both requirements. Genes that encode enzymes that are undesirable serve as potential integration sites for genes that are desirable. For example, 1,6 mannosyltransferase activity is a hallmark of glycosylation in many known lower eukaryotes. The gene encoding alpha-1,6 mannosyltransferase (*OCH1*) has been cloned from *S.cerevisiae* and mutations in the gene give rise to a viable phenotype with reduced mannosylation. The gene locus encoding alpha-1,6 mannosyltransferase activity therefor is a prime target for the integration of genes encoding glycosyltransferase activity. In a similar manner, one can choose a range of other chromosomal integration sites that, based on a gene disruption event in that locus, are expected to: (1) improve the cells ability to glycosylate in a more human like fashion, (2) improve the cells ability to secrete proteins, (3) reduce proteolysis of foreign proteins and (4) improve other characteristics of the process that facilitate purification or the fermentation process itself.

Providing sugar nucleotide precursors

[0154] A hallmark of higher eukaryotic glycosylation is the presence of galactose, fucose, and a high degree of terminal sialic acid on glycoproteins. These sugars are not generally found on glycoproteins produced in yeast and filamentous fungi and the method discussed above allows for the engineering of strains that localize glycosyltransferase in the desired organelle. Formation of complex N-glycan synthesis is a sequential process by which specific sugar residues are removed and attached to the core oligosaccharide structure. In higher eukaryotes, this is achieved by having the substrate sequentially exposed to various processing enzymes. These enzymes carry out specific reactions depending on their particular location within the entire processing cascade. This "assembly line"

consists of ER, early, medial and late Golgi, and the trans Golgi network all with their specific processing environment. To recreate the processing of human glycoproteins in the Golgi and ER of lower eukaryotes, numerous enzymes (e.g. glycosyltransferases, glycosidases, phosphatases and transporters) have to be expressed and specifically targeted to these organelles, and preferably, in a location so that they function most efficiently in relation to their environment as well as to other enzymes in the pathway. [0155] Several individual glycosyltransferases have been cloned and expressed in *S.cerevisiae* (GalT, GnT I), *Aspergillus nidulans* (GnT I) and other fungi, without however demonstrating the desired outcome of "humanization" on the glycosylation pattern of the organisms (Yoshida, 1995; Schwientek, 1995; Kalsner, 1995). It was speculated that the carbohydrate structure required to accept sugars by the action of such glycosyltransferases was not present in sufficient amounts. While this most likely contributed to the lack of complex N-glycan formation, there are currently no reports of a fungus supplying a Man₅GlcNAc₂ structure, having GnT I activity and having UDP-Gn transporter activity engineered into the fungus. It is the combination of these three biochemical events that are required for hybrid and complex N-glycan formation.

[0156] In humans, the full range of nucleotide sugar precursors (e.g. UDP-N-acetylglucosamine, UDP-N-acetylgalactosamine, CMP-N-acetylneuraminic acid, UDP-galactose, etc.) are generally synthesized in the cytosol and transported into the Golgi, where they are attached to the core oligosaccharide by glycosyltransferases. To replicate this process in lower eukaryotes, sugar nucleoside specific transporters have to be expressed in the Golgi to ensure adequate levels of nucleoside sugar precursors (Sommers, 1981; Sommers, 1982; Perez, 1987). A side product of this reaction is either a nucleoside diphosphate or monophosphate. While monophosphates can be directly exported in exchange for nucleoside triphosphate sugars by an antiport mechanism, diphospho nucleosides (e.g. GDP) have to be cleaved by phosphatases (e.g. GDPase) to yield nucleoside monophosphates and inorganic phosphate prior to being exported. This reaction appears to be important for efficient glycosylation, as GDPase from *S.cerevisiae* has been found to be necessary for mannosylation. However, the enzyme only has

10% of the activity towards UDP (Berninsone, 1994). Lower eukaryotes often do not have UDP specific diphosphatase activity in the Golgi since they do not utilize UDP-sugar precursors for glycoprotein synthesis in the Golgi.

[0157] *Schizosaccharomyces pombe*, a yeast found to add galactose residues to
5 cell wall polysaccharides (from UDP-galactose) was found to have specific
UDPase activity further suggesting the requirement for such an enzyme
(Berninsone et al., 1994). UDP is known to be a potent inhibitor of
glycosyltransferases and the removal of this glycosylation side product is
important in order to prevent glycosyltransferase inhibition in the lumen of the
10 Golgi (Khatarra et al., 1974). Thus, one may need to provide for the removal of
UDP, which is expected to accumulate in the Golgi of such an engineered strains
(Berninsone, 1995; Beaudet, 1998). [0158] In another example, 2,3
sialyltransferase and 2,6 sialyltransferase cap galactose residues with sialic acid in
the trans-Golgi and TGN of humans leading to a mature form of the glycoprotein.
15 To reengineer this processing step into a metabolically engineered yeast or fungus
will require (1) 2,3-sialyltransferase activity and (2) a sufficient supply of CMP-N-
acetyl neuraminic acid, in the late Golgi of yeast. To obtain sufficient 2,3-
sialyltransferase activity in the late Golgi, the catalytic domain of a known
sialyltransferase (e.g. from humans) has to be directed to the late Golgi in fungi
20 (see above). Likewise, transporters have to be engineered to that allow the
transport of CMP-N-acetyl neuraminic acid into the late Golgi. There is currently
no indication that fungi synthesize sufficient amounts of CMP-N-acetyl neuraminic
acid, not to mention the transport of such a sugar-nucleotide into the Golgi.
Consequently, to ensure the adequate supply of substrate for the corresponding
25 glycosyltransferases, one has to metabolically engineer the production of CMP-
sialic acid into the fungus.

Methods for providing sugar nucleotide precursors to the Golgi apparatus:

UDP-N-acetyl-glucosamine

30 [0159] The cDNA of human UDP-N-acetylglucosamine transporter, which was
recognized through a homology search in the expressed sequence tags database
(dbEST) was cloned by Ishida and coworkers (Ishida, 1999). Guillen and

coworkers have cloned the mammalian Golgi membrane transporter for UDP-N-acetylglucosamine by phenotypic correction with cDNA from canine kidney cells (MDCK) of a recently characterized *Kluyveromyces lactis* mutant deficient in Golgi transport of the above nucleotide sugar (Guillen, 1998). Their results demonstrate that the mammalian Golgi UDP-GlcNAc transporter gene has all of the necessary information for the protein to be expressed and targeted functionally to the Golgi apparatus of yeast and that two proteins with very different amino acid sequences may transport the same solute within the same Golgi membrane (Guillen, 1998).

10 *GDP-Fucose*

[0160] The rat liver Golgi membrane GDP-fucose transporter has been identified and purified by Puglielli, L. and C. B. Hirschberg (Puglielli, 1999). The corresponding gene has not been identified however N-terminal sequencing can be used for the design of oligonucleotide probes specific for the corresponding gene. These oligonucleotides can be used as probes to clone the gene encoding for GDP-fucose transporter.

UDP-Galactose

[0161] Two heterologous genes, *gmal2(+)* encoding alpha 1,2-galactosyltransferase (alpha 1,2 GalT) from *Schizosaccharomyces pombe* and (hUGT2) encoding human UDP-galactose (UDP-Gal) transporter, have been functionally expressed in *S.cerevisiae* to examine the intracellular conditions required for galactosylation. Correlation between protein galactosylation and UDP-galactose transport activity indicated that an exogenous supply of UDP-Gal transporter, rather than alpha 1,2 GalT played a key role for efficient galactosylation in *S.cerevisiae* (Kainuma, 1999). Likewise a UDP-galactose transporter from *S. pombe* was cloned (Aoki, 1999; Segawa, 1999).

CMP-N-acetylneuraminic acid (CMP-Sialic acid)

[0162] Human CMP-sialic acid transporter (hCST) has been cloned and expressed in Lec 8 CHO cells (Aoki, 1999; Eckhardt, 1997). The functional expression of the murine CMP-sialic acid transporter was achieved in *Saccharomyces cerevisiae* (Berninsone, 1997). Sialic acid has been found in some fungi, however it is not clear whether the chosen host system will be able to supply

sufficient levels of CMP-Sialic acid. Sialic acid can be either supplied in the medium or alternatively fungal pathways involved in sialic acid synthesis can also be integrated into the host genome.

5 **Diphosphatases**

[0163] When sugars are transferred onto a glycoprotein, either a nucleoside diphosphate or monophosphate, is released from the sugar nucleotide precursors. While monophosphates can be directly exported in exchange for nucleoside triphosphate sugars by an antiport mechanism, diphospho nucleosides (e.g. GDP) have to be cleaved by phosphatases (e.g. GDPase) to yield nucleoside monophosphates and inorganic phosphate prior to being exported. This reaction appears to be important for efficient glycosylation, as GDPase from *S.cerevisiae* has been found to be necessary for mannosylation. However, the enzyme only has 10% of the activity towards UDP (Berninsone, 1994). Lower eukayotes often do not have UDP specific diphosphatase activity in the Golgi since they do not utilize UDP-sugar precursors for glycoprotein synthesis in the Golgi. *Schizosaccharomyces pombe*, a yeast found to add galactose residues to cell wall polysaccharides (from UDP-galactose) was found to have specific UDPase activity further suggesting the requirement for such an enzyme (Berninsone, 1994). UDP is known to be a potent inhibitor of glycosyltransferases and the removal of this glycosylation side product is important in order to prevent glycosyltransferase inhibition in the lumen of the Golgi (Khatara et al. 1974).

25 **Expression Of GnTs To Produce Complex N-glycans**

25 Expression Of GnT-III To Boost Antibody Functionality

[0164] The addition of an N-acetylglucosamine to the $\text{GlcNAc}_1\text{Man}_3\text{GlcNAc}_2$ structure by N-acetylglucosaminyltransferases II and III yields a so-called bisected N-glycan $\text{GlcNAc}_3\text{Man}_3\text{GlcNAc}_2$ (Fig. 3). This structure has been implicated in greater antibody-dependent cellular cytotoxicity (ADCC) (Umana et al. 1999). Re-engineering glycoforms of immunoglobulins expressed by mammalian cells is a tedious and cumbersome task. Especially in the case of GnTIII, where over-

expression of this enzyme has been implicated in growth inhibition, methods involving regulated (inducible) gene expression had to be employed to produce immunoglobulins with bisected N-glycans (Umana et al 1999a, 1999b).

[0165] Accordingly, in another embodiment, the invention provides systems and methods for producing human-like N-glycans having bisecting N-acetylglucosamine (GlcNAcs) on the core mannose structure. In a preferred embodiment, the invention provides a system and method for producing immunoglobulins having bisected N-glycans. The systems and methods described herein will not suffer from previous problems, e.g., cytotoxicity associated with overexpression of GnTIII or ADCC, as the host cells of the invention are engineered and selected to be viable and preferably robust cells which produce N-glycans having substantially modified human-type glycoforms such as $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$. Thus, addition of a bisecting N-acetylglucosamine in a host cell of the invention will have a negligible effect on the growth-phenotype or viability of those host cells.

[0166] In addition, previous work (Umana) has shown that there is no linear correlation between GnTIII expression levels and the degree of ADCC. Finding the optimal expression level in mammalian cells and maintaining it throughout an FDA approved fermentation process seems to be a challenge. However, in cells of the invention, such as fungal cells, finding a promoter of appropriate strength to establish a robust, reliable and optimal GnTIII expression level is a comparatively easy task for one of skill in the art.

[0167] A host cell such as a yeast strain capable of producing glycoproteins with bisecting N-glycans is engineered according to the invention, by introducing into the host cell a GnTIII activity (Example 6). Preferably, the host cell is transformed with a nucleic acid that encodes GnTIII (see, e.g., Fig. 32) or a domain thereof having enzymatic activity, optionally fused to a heterologous cell signal targeting peptide (e.g., using the libraries and associated methods of the invention.) Host cells engineered to express GnTIII will produce higher antibody titers than mammalian cells are capable of. They will also produce antibodies with higher potency with respect to ADCC.

[0168] Antibodies produced by mammalian cell lines transfected with GnTIII have been shown to be as effective as antibodies produced by non-transfected cell-lines, but at a 10-20 fold lower concentration (Davies et al. 2001). An increase of productivity of the production vehicle of the invention over mammalian systems by a factor of twenty, and a ten-fold increase of potency will result in a net-productivity improvement of two hundred. The invention thus provides a system and method for producing high titers of an antibody having high potency (e.g., up to several orders of magnitude more potent than what can currently be produced). The system and method is safe and provides high potency antibodies at low cost in short periods of time. Host cells engineered to express GnT III according to the invention produce immunoglobulins having bisected N-glycans at rates of at least 50 mg/liter/day to at least 500 mg/liter/day. In addition, each immunoglobulin (Ig) molecule (comprising bisecting GlcNAcs) is more potent than the same Ig molecule produced without bisecting GlcNAcs.

Cloning and expression of GnT-IV and GnT-V

[0169] All branching structures in complex N-glycans are synthesized on a common core-pentasaccharide ($\text{Man}_3\text{GlcNAc}_2$ or $\text{Man } \alpha 1-6(\text{Man } \alpha 1-3)\text{Man } \beta 1-4 \text{ GlcNAc } \beta 1-4 \text{ GlcNAc } \beta 1-4$ or $\text{Man}_3\text{GlcNAc}_2$) by N-acetylglucosamine transferases (GnTs) -I to -VI (Schachter H et al. (1989) *Methods Enzymo*;179:351-97). Current understanding of the biosynthesis of more highly branched N-glycans suggests that after the action of GnTII (generation of $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$ structures) GnTIV transfers GlcNAc from UDP-GlcNAc in $\beta 1,4$ linkage to the $\text{Man } \alpha 1,3 \text{ Man } \beta 1,4$ arm of $\text{GlcNAc}_2\text{Man}_3\text{GlcNAc}_2$ N-glycans (Allen SD et al. (1984) *J Biol Chem.* Jun 10;259(11):6984-90; and Gleeson PA and Schachter H.J (1983); *J.Biol Chem* 25;258(10):6162-73) resulting in a triantennary agalacto sugar chain. This N-glycan ($\text{GlcNAc } \beta 1-2 \text{ Man } \alpha 1-6(\text{GlcNAc } \beta 1-2 \text{ Man } \alpha 1-3) \text{ Man } \beta 1-4 \text{ GlcNAc } \beta 1-4 \text{ GlcNAc } \beta 1,4 \text{ Asn}$) is a common substrate for GnT-III and -V, leading to the synthesis of bisected, tri-and tetra-antennary structures. Where the action of GnTIII results in a bisected N-glycan and where GnTV catalyzes the addition of $\beta 1-6\text{GlcNAc}$ to the $\alpha 1-6$ mannosyl core, creating the $\beta 1-6$ branch. Addition of galactose

and sialic acid to these branches leads to the generation of a fully sialylated complex N-glycan.

[0170] Branched complex N-glycans have been implicated in the physiological activity of therapeutic proteins, such as human erythropoietin (hEPO). Human
5 EPO having bi-antennary structures has been shown to have a low activity, whereas hEPO having tetra-antennary structures resulted in slower clearance from the bloodstream and thus in higher activity (Misaizu T et al. (1995) *Blood* Dec 1;86(11):4097-104).

[0171] With DNA sequence information, the skilled worker can clone DNA
10 molecules encoding GnT IV and/or V activities (Example 6; Figs. 33 and 34). Using standard techniques well-known to those of skill in the art, nucleic acid molecules encoding GnT IV or V (or encoding catalytically active fragments thereof) may be inserted into appropriate expression vectors under the transcriptional control of promoters and other expression control sequences
15 capable of driving transcription in a selected host cell of the invention, e.g., a fungal host such as *Pichia sp.*, *Kluyveromyces sp.* and *Aspergillus sp.*, as described herein, such that one or more of these mammalian GnT enzymes may be actively expressed in a host cell of choice for production of a human-like complex glycoprotein.

20

[0172] The following are examples which illustrate the compositions and methods of this invention. These examples should not be construed as limiting: the examples are included for the purposes of illustration only.

25

EXAMPLE 1

Identification, cloning and deletion of the *ALG3* gene in *P.pastoris* and *K.lactis*.

[0173] Degenerate primers were generated based on an alignment of Alg3 protein sequences from *S. cerevisiae*, *H. sapiens*, and *D. melanogaster* and were used to amplify an 83 bp product from *P. pastoris* genomic DNA:
30 5'-GGTGT TTTGTTTCTAGATCTTTGCAYTAYCARTT-3' and
5'-AGAATTTGGTGGGTAAGAATTCCARCACCAYTCRTG-3' The resulting PCR product was cloned into the pCR2.1 vector (Invitrogen, Carlsbad, CA) and

sequence analysis revealed homology to known *ALG3/RHK1/NOT56* homologs (Genbank NC_001134.2, AF309689, NC_003424.1). Subsequently, 1929 bp upstream and 2738 bp downstream of the initial PCR product were amplified from a *P. pastoris* genomic DNA library (Boehm, T. Yeast 1999 May;15(7):563-72)

5 using the internal oligonucleotides
 5'-CCTAAGCTGGTATGCGTTCTCTTTGCCATATC-3' and
 5'-GCGGCATAACAATAATAGATGCTATAAAG-3' along with T3
 (5'-AATTAACCCTCACTAAAGGG-3') and T7 (5'-GTAA
 TACGACTCACTATAGGGC-3') (Integrated DNA Technologies, Coralville, IA)

10 in the backbone of the library bearing plasmid lambda ZAP II (Stratagene, La Jolla, CA). The resulting fragments were cloned into the pCR2.1-TOPO vector (Invitrogen) and sequenced. From this sequence, a 1395 bp ORF was identified that encodes a protein with 35% identity and 53% similarity to the *S. cerevisiae* *ALG3* gene (using BLAST programs). The gene was named *PpALG3*.

15 [0174] The sequence of *PpALG3* was used to create a set of primers to generate a deletion construct of the *PpALG3* gene by PCR overlap (Davidson et al, 2002 Microbiol. 148(Pt 8):2607-15). Primers below were used to amplify 1 kb regions 5' and 3' of the *PpALG3* ORF and the *KAN^R* gene, respectively:
 RCD142 (5'-CCACATCATCCGTGCTACATATAG-3'),
 20 RCD144 (5'-ACGAGGCAAGCTAAACAGATCTCGAAGTATCGAGGGTTAT
 CCAG-3'),
 RCD145 (5'-CCATCCAGTGTCGAAAACGAGCCAATGGTTCATGTCTATA
 AATC-3'),
 RCD147 (5'-AGCCTCAGCGCCAACAAGCGATGG-3'),
 25 RCD143 (5'-CTGGATAACCCTCGATACTTCGAGATCTGTTTAGCTTGCC
 TCGT-3'), and
 RCD146 (5'-GATTATAGACATGAACCATTTGGCTCGTTTTTCGACACTGG
 ATGG-3').

Subsequently, primers RCD142 and RCD147 were used to overlap the three
 30 resulting PCR products into a single 3.6 kb *alg3::KAN^R* deletion allele.

Identification, cloning and deletion of the *ALG3* gene in *K.lactis*.

[0175] The *ALG3p* sequences from *S. cerevisiae*, *Drosophila melanogaster*, *Homo sapiens* etc were aligned with *K. lactis* sequences (PENDANT EST database). Regions of high homology that were in common homologs but distinct in exact sequence from the homologs were used to create pairs of degenerate
5 primers that were directed against genomic DNA from the *K. lactis* strain MG1/2 (Bianchi et al, 1987). In the case of *ALG3*, PCR amplification with primers KAL-1 (5'-ATCCTTTACCGATGCTGTAT-3') and KAL-2 (5'-ATAACAGTATGTGTTACACGCGTGTAG-3') resulted in a product that was cloned and sequenced and the predicted translation was shown to have a high
10 degree of homology to Alg3p proteins (>50% to *S. cerevisiae* Alg3p).
[0176] The PCR product was used to probe a Southern blot of genomic DNA from *K. lactis* strain (MG1/2) with high stringency (Sambrook et al, 1989). Hybridization was observed in a pattern consistent with a single gene. This Southern blot was used to map the genomic loci. Genomic fragments were cloned
15 by digesting genomic DNA and ligating those fragments in the appropriate size-range into pUC19 to create a *K. lactis* subgenomic library. This subgenomic library was transformed into *E. coli* and several hundred clones were tested by colony PCR, using primers KAL-1 and KAL-2. The clones containing the predicted *KIALG3* and *KIALG61* genes were sequenced and open reading frames
20 identified.
[0177] Primers for construction of an *alg3::NAT^R* deletion allele, using a PCR overlap method (Davidson et al, 2002), were designed and the resulting deletion allele was transformed into two *K. lactis* strains and NAT-resistant colonies selected. These colonies were screened by PCR and transformants were obtained
25 in which the *ALG3* ORF was replaced with the *och1::NAT^R* mutant allele.

EXAMPLE 2

Generation of an *alg3/och1* mutant strain expressing an α -1,2-Mannosidase, GnT1 and GnTII for production of a human-like glycoprotein.

[0178] The 1215 bp open reading frame of the *P. pastoris OCH1* gene as well as
30 2685 bp upstream and 1175 bp downstream was amplified by PCR (B. K. Choi et al., submitted to *Proc. Natl. Acad. Sci. USA* 2002; see also WO 02/00879; each of which is incorporated herein by reference), cloned into the pCR2.1-TOPO vector

(Invitrogen) and designated pBK9. To create an *och1* knockout strain containing multiple auxotrophic markers, 100 µg of pJN329, a plasmid containing an *och1::URA3* mutant allele flanked with *SfiI* restriction sites was digested with *SfiI* and used to transform *P. pastoris* strain JC308 (Cereghino et al. *Gene* 263 (2001) 159-169) by electroporation. Following incubation on defined medium lacking uracil for 10 days at room temperature, 1000 colonies were picked and re-streaked. URA^+ clones that were unable to grow at 37°C, but grew at room temperature, were subjected to colony PCR to test for the correct integration of the *och1::URA3* mutant allele. One clone that exhibited the expected PCR pattern was designated YJN153. The Kringle 3 domain of human plasminogen (K3) was used as a model protein. A Neo^R marked plasmid containing the K3 gene was transformed into strain YJN153 and a resulting strain, expressing K3, was named BK64-1 (B. K. Choi et al, submitted to *Proc. Natl. Acad. Sci. USA* 2002).

[0179] Plasmid pPB103, containing the *Kluyveromyces lactis* *MNN2-2* gene, encoding a Golgi UDP-N-acetylglucosamine transporter was constructed by cloning a blunt *BglIII-HindIII* fragment from vector pDL02 (Abeijon et al. (1996) *Proc. Natl. Acad. Sci. U.S.A.* 93:5963-5968) into *BglIII* and *BamHI* digested and blunt ended pBLADE-SX containing the *P. pastoris* *ADE1* gene (Cereghino et al. (2001) *Gene* 263:159-169). This plasmid was linearized with *EcoNI* and transformed into strain BK64-1 by electroporation and one strain confirmed to contain the *MNN2-2* by PCR analysis was named PBP1.

[0180] A library of mannosidase constructs was generated, comprising in-frame fusions of the leader domains of several type I or type II membrane proteins from *S. cerevisiae* and *P. pastoris* fused with the catalytic domains of several α -1,2-mannosidase genes from human, mouse, fly, worm and yeast sources (see, e.g., WO02/00879, incorporated herein by reference). This library was created in a *P. pastoris* *HIS4* integration vector and screened by linearizing with *SalI*, transforming by electroporation into strain PBP1, and analyzing the glycans released from the K3 reporter protein. One active construct chosen was a chimera of the 988-1296 nucleotides (C-terminus) of the yeast *SEC12* gene fused with a N-terminal deletion of the mouse α -1,2-mannosidase IA (MmMannIA) gene, which

was missing the 187 nucleotides. A *P. pastoris* strain expressing this construct was named PBP2.

[0181] A library of GnTI constructs was generated, comprising in-frame fusions of the same leader library with the catalytic domains of GnTI genes from human, worm, frog and fly sources (WO 02/00879). This library was created in a *P. pastoris* ARG4 integration vector and screened by linearizing with *Aat*II, transforming by electroporation into strain PBP2, and analyzing the glycans released from K3. One active construct chosen was a chimera of the first 120 bp of the *S. cerevisiae* MNN9 gene fused to a deletion of the human GnTI gene, which was missing the first 154 bp. A *P. pastoris* strain expressing this construct was named PBP3.

[0182] Subsequently, a *P. pastoris* *alg3::KAN^R* deletion construct was generated as described above. Approximately 5µg of the resulting PCR product was transformed into strain PBP3 and colonies were selected on YPD medium containing 200µg/ml G418. One strain out of 20 screened by PCR was confirmed to contain the correct integration of the *alg3::KAN^R* mutant allele and lack the wild-type allele. This strain was named RDP27.

[0183] Finally, a library of GnTII constructs was generated, which was comprised of in-frame fusions of the leader library with the catalytic domains of GnTII genes from human and rat sources (WO 02/00879). This library was created in a *P. pastoris* integration vector containing the NST^R gene conferring resistance to the drug nourseothricin. The library plasmids were linearized with *Eco*RI, transformed into strain RDP27 by electroporation, and the resulting strains were screened by analysis of the released glycans from purified K3.

Materials

[0184] MOPS, sodium cacodylate, manganese chloride, UDP-galactose and CMP-N-acetylneuraminic acid were from Sigma. TFA was from Aldrich. Recombinant rat α 2,6-sialyltransferase from *Spodoptera frugiperda* and β 1,4-galactosyltransferase from bovine milk were from Calbiochem. Protein N-glycosidase F, mannosidases, and oligosaccharides were from Glyko (San Rafael, CA). DEAE ToyoPearl resin was from TosoHaas. Metal chelating "HisBind"

resin was from Novagen (Madison, WI). 96-well lysate-clearing plates were from Promega (Madison, WI). Protein-binding 96-well plates were from Millipore (Bedford, MA). Salts and buffering agents were from Sigma (St. Louis, MO). MALDI matrices were from Aldrich (Milwaukee, WI).

5

Protein Purification

[0185] Kringle 3 was purified using a 96-well format on a Beckman BioMek 2000 sample-handling robot (Beckman/Coulter Ranch Cucamonga, CA). Kringle 3 was purified from expression media using a C-terminal hexa-histidine tag. The robotic purification is an adaptation of the protocol provided by Novagen for their HisBind resin. Briefly, a 150uL (μ L) settled volume of resin is poured into the wells of a 96-well lysate-binding plate, washed with 3 volumes of water and charged with 5 volumes of 50mM NiSO₄ and washed with 3 volumes of binding buffer (5mM imidazole, 0.5M NaCl, 20mM Tris-HCL pH7.9). The protein expression media is diluted 3:2, media/PBS (60mM PO₄, 16mM KCl, 822mM NaCl pH7.4) and loaded onto the columns. After draining, the columns are washed with 10 volumes of binding buffer and 6 volumes of wash buffer (30mM imidazole, 0.5M NaCl, 20mM Tris-HCl pH7.9) and the protein is eluted with 6 volumes of elution buffer (1M imidazole, 0.5M NaCl, 20mM Tris-HCl pH7.9). The eluted glycoproteins are evaporated to dryness by lyophilization.

20

Release of N-linked Glycans

[0186] The glycans are released and separated from the glycoproteins by a modification of a previously reported method (Papac, et al. A. J. S. (1998) Glycobiology 8, 445-454). The wells of a 96-well MultiScreen IP (Immobilon-P membrane) plate (Millipore) are wetted with 100uL of methanol, washed with 3X150uL of water and 50uL of RCM buffer (8M urea, 360mM Tris, 3.2mM EDTA pH8.6), draining with gentle vacuum after each addition. The dried protein samples are dissolved in 30uL of RCM buffer and transferred to the wells containing 10uL of RCM buffer. The wells are drained and washed twice with RCM buffer. The proteins are reduced by addition of 60uL of 0.1M DTT in RCM buffer for 1hr at 37°C. The wells are washed three times with 300uL of water and

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carboxymethylated by addition of 60uL of 0.1M iodoacetic acid for 30min in the dark at room temperature. The wells are again washed three times with water and the membranes blocked by the addition of 100uL of 1% PVP 360 in water for 1hr at room temperature. The wells are drained and washed three times with 300uL of water and deglycosylated by the addition of 30uL of 10mM NH₄HCO₃ pH 8.3 containing one milliunit of N-glycanase (Glyko). After 16 hours at 37°C, the solution containing the glycans was removed by centrifugation and evaporated to dryness.

10 **Matrix Assisted Laser Desorption Ionization Time of Flight Mass Spectrometry**

[0187] Molecular weights of the glycans were determined using a Voyager DE PRO linear MALDI-TOF (Applied Biosciences) mass spectrometer using delayed extraction. The dried glycans from each well were dissolved in 15uL of water and 0.5uL spotted on stainless steel sample plates and mixed with 0.5uL of S-DHB matrix (9mg/mL of dihydroxybenzoic acid, 1mg/mL of 5-methoxysalicylic acid in 1:1 water/acetonitrile 0.1% TFA) and allowed to dry.

[0188] Ions were generated by irradiation with a pulsed nitrogen laser (337nm) with a 4ns pulse time. The instrument was operated in the delayed extraction mode with a 125ns delay and an accelerating voltage of 20kV. The grid voltage was 93.00%, guide wire voltage was 0.10%, the internal pressure was less than 5 X 10⁻⁷ torr, and the low mass gate was 875Da. Spectra were generated from the sum of 100-200 laser pulses and acquired with a 2 GHz digitizer. Man5 oligosaccharide was used as an external molecular weight standard. All spectra were generated with the instrument in the positive ion mode. The estimated mass accuracy of the spectra was 0.5%.

Materials:

[0189] MOPS, sodium cacodylate, manganese chloride, UDP-galactose and CMP-N-acetylneuraminic acid were from Sigma, Saint Louis, MO. Trifluoroacetic acid (TFA) was from Sigma/Aldrich, Saint Louis, MO. Recombinant rat alpha-2,6-

sialyltransferase from *Spodoptera frugiperda* and beta-1,4-galactosyltransferase from bovine milk were from Calbiochem, San Diego, CA.

β -N-acetylhexosaminidase Digestion

5 [0190] The glycans were released and separated from the glycoproteins by a modification of a previously reported method (Papac, et al. A. J. S. (1998) *Glycobiology* 8, 445-454). After the proteins were reduced and carboxymethylated, and the membranes blocked, the wells were washed three time with water. The protein was deglycosylated by the addition of 30 μ l of 10 mM NH_4HCO_3 pH 8.3
10 containing one milliunit of N-glycanase (Glyko, Novato, CA). After 16 hr at 37°C, the solution containing the glycans was removed by centrifugation and evaporated to dryness. The glycans were then dried in SC210A speed vac (Thermo Savant, Halbrook, NY). The dried glycans were put in 50 mM NH_4Ac pH 5.0 at 37°C overnight and 1mU of hexos (Glyko, Novato, CA) was added.

15

Galactosyltransferase Reaction

[0191] Approximately 2mg of protein (r-K3:hPg [PBP6-5]) was purified by nickel-affinity chromatography, extensively dialyzed against 0.1% TFA, and lyophilized to dryness. The protein was redissolved in 150 μ L of 50mM MOPS,
20 20mM MnCl_2 , pH7.4. After addition of 32.5 μ g (533nmol) of UDP-galactose and 4mU of β 1,4-galactosyltransferase, the sample was incubated at 37°C for 18 hours. The samples were then dialyzed against 0.1% TFA for analysis by MALDI-TOF mass spectrometry.

[0192] The spectrum of the protein reacted with galactosyltransferase showed an
25 increase in mass consistent with the addition of two galactose moieties when compared with the spectrum of a similar protein sample incubated without enzyme. Protein samples were next reduced, carboxymethylated and deglycosylated with PNGase F. The recovered N-glycans were analyzed by MALDI-TOF mass spectrometry. The mass of the predominant glycan from the galactosyltransferase
30 reacted protein was greater than that of the control glycan by a mass consistent with the addition of two galactose moieties (325.4 Da).

Sialyltransferase Reaction

[0193] After resuspending the (galactosyltransferase reacted) proteins in 10 μ L of 50mM sodium cacodylate buffer pH6.0, 300 μ g (488nmol) of CMP-N-acetylneuraminic acid (CMP-NANA) dissolved in 15 μ L of the same buffer, and 5 μ L (2mU) of recombinant α -2,6 sialyltransferase were added. After incubation at 37 $^{\circ}$ C for 15 hours, an additional 200 μ g of CMP-NANA and 1mU of sialyltransferase were added. The protein samples were incubated for an additional 8 hours and then dialyzed and analyzed by MALDI-TOF-MS as above.

[0194] The spectrum of the glycoprotein reacted with sialyltransferase showed an increase in mass when compared with that of the starting material (the protein after galactosyltransferase reaction). The N-glycans were released and analyzed as above. The increase in mass of the two ion-adducts of the predominant glycan was consistent with the addition of two sialic acid residues (580 and 583Da).

15

EXAMPLE 3
Identification, cloning and deletion of the
ALG9* and *ALG 12* genes in *P.pastoris

[0195] Similar to Example 1, the ALG9p and ALG12 sequences, respectively from *S. cerevisiae*, *Drosophila melanogaster*, *Homo sapiens*, etc., is aligned and regions of high homology are used to design degenerate primers. These primers are employed in a PCR reaction on genomic DNA from the *P. pastoris*. The resulting initial PCR product is subcloned, sequenced and used to probe a Southern blot of genomic DNA from *P. pastoris* with high stringency (Sambrook et al., 1989). Hybridization is observed. This Southern blot is used to map the genomic loci. Genomic fragments are cloned by digesting genomic DNA and ligating those fragments in the appropriate size-range into pUC19 to create a *P. pastoris* subgenomic library. This subgenomic library is transformed into *E. coli* and several hundred clones tested by colony PCR, using primers designed based on the sequence of the initial PCR product. The clones containing the predicted genes are sequenced and open reading frames identified. Primers for construction of an *alg9::NAT^R* deletion allele, using a PCR overlap method (Davidson et al., 2002), are designed. The resulting deletion allele is transformed into two *P.pastoris*

strains and NAT resistant colonies are selected. These colonies are screened by PCR and transformants obtained in which the *ALG9* ORF is replaced with the *och1::NAT^R* mutant allele. See generally, Cipollo et al. *Glycobiology* 2002 (12)11:749-762; Chantret et al. *J. Biol. Chem.* Jul. 12, 2002 (277)28:25815-25822; Cipollo et al. *J. Biol. Chem.* Feb. 11, 2000 (275)6:4267-4277; Burda et al. *Proc. Natl. Acad. Sci. U.S.A.* July 1996 (93):7160-7165; Karaoglu et al. *Biochemistry* 2001, 40, 12193-12206; Grimme et al. *J. Biol. Chem.* July 20, 2001 (276)29:27731-27739; Verostek et al. *J. Biol. Chem.* June 5, 1993 (268)16:12095-12103; Huffaker et al. *Proc. Natl. Acad. Sci. U.S.A.* Dec. 1983 (80):7466-7470.

EXAMPLE 4

Identification, cloning and expression of Alpha 1,2-3 Mannosidase From *Xanthomonas Manihotis*

[0196] The alpha 1,2-3 Mannosidase from *Xanthomonas Manihotis* has two activities: an alpha-1,2 and an alpha-1,3 mannosidase. The methods of the invention may also use two independent mannosidases having these activities, which may be similarly identified and cloned from a selected organism of interest.

[0197] As described by Landry et al., alpha-mannosidases can be purified from *Xanthomonas sp.*, such as *Xanthomonas manihotis*. *X. manihotis* can be purchased from the American Type Culture Collection (ATCC catalog number 49764) (*Xanthomonas axonopodis* Starr and Garces pathovar *manihotis* deposited as *Xanthomonas manihotis* (Arthaud-Berthet) Starr). Enzymes are purified from crude cell-extracts as previously described (Wong-Madden, S.T. and Landry, D. (1995) Purification and characterization of novel glycosidases from the bacterial genus *Xanthomonas*; and Landry, D. US Patent US 6,300,113 B1 Isolation and composition of novel Glycosidases). After purification of the mannosidase, one of several methods are used to obtain peptide sequence tags (see, e.g., W. Quadroni M et al. (2000). A method for the chemical generation of N-terminal peptide sequence tags for rapid protein identification. *Anal Chem* (2000) Mar 1;72(5):1006-14; Wilkins MR et al. Rapid protein identification using N-terminal "sequence tag" and amino acid analysis. *Biochem Biophys Res Commun.* (1996)

Apr 25;221(3):609-13; and Tsugita A. (1987) Developments in protein microsequencing. *Adv Biophys* (1987) 23:81-113).

[0198] Sequence tags generated using a method above are then used to generate sets of degenerate primers using methods well-known to the skilled worker.

5 Degenerate primers are used to prime DNA amplification in polymerase chain reactions (e.g., using Taq polymerase kits according to manufacturers' instructions) to amplify DNA fragments. The amplified DNA fragments are used as probes to isolate DNA molecules comprising the gene encoding a desired mannosidase, e.g., using standard Southern DNA hybridization techniques to
10 identify and isolate (clone) genomic pieces encoding the enzyme of interest. The genomic DNA molecules are sequenced and putative open reading frames and coding sequences are identified. A suitable expression construct encoding for the glycosidase of interest can then be generated using methods described herein and well-known in the art.

15 [0199] Nucleic acid fragments comprising sequences encoding alpha 1,2-3 mannosidase activity (or catalytically active fragments thereof) are cloned into appropriate expression vectors for expression, and preferably targeted expression, of these activities in an appropriate host cell according to the methods set forth herein.

20

EXAMPLE 5

Identification, cloning and expression of the *ALG6* gene in *P.pastoris*

[0200] Similar to Example 1, the ALG6p sequences from *S. cerevisiae*, *Drosophila melanogaster*, *Homo sapiens* etc., are aligned and regions of high
25 homology are used to design degenerate primers. These primers are employed in a PCR reaction on genomic DNA from the *P. pastoris*. The resulting initial PCR product is subcloned, sequenced and used to probe a Southern blot of genomic DNA from *P. pastoris* with high stringency (Sambrook et al, 1989). Hybridization is observed. This Southern blot is used to map the genomic loci. Genomic
30 fragments are cloned by digesting genomic DNA and ligating those fragments in the appropriate size-range into pUC19 to create a *P. pastoris* subgenomic library. This subgenomic library is transformed into *E. coli* and several hundred clones are

tested by colony PCR, using primers designed based on the sequence of the initial PCR product. The clones containing the predicted genes are sequenced and open reading frames identified. Primers for construction of an *alg6::NAT^R* deletion allele, using a PCR overlap method (Davidson et al, 2002), are designed and the resulting deletion allele is transformed into two *P. pastoris* strains and NAT resistant colonies selected. These colonies are screened by PCR and transformants are obtained in which the *ALG6* ORF is replaced with the *och1::NAT^R* mutant allele. See, e.g., Imbach et al. *Proc. Natl. Acad. Sci. U.S.A.* June 1999 (96)6982-6987.

10 [0201] Nucleic acid fragments comprising sequences encoding Alg6p (or catalytically active fragments thereof) are cloned into appropriate expression vectors for expression, and preferably targeted expression, of these activities in an appropriate host cell according to the methods set forth herein. The cloned *ALG6* gene can be brought under the control of any suitable promoter to achieve

15 overexpression. Even expression of the gene under the control of its own promoter is possible. Expression from multicopy plasmids will generate high levels of expression ("overexpression").

EXAMPLE 6

20 Cloning and Expression Of GnT III To Produce Bisecting GlcNAcs Which Boost Antibody Functionality

A. Background

25 [0202] The addition of an N-acetylglucosamine to the GlcNAc₂Man₃GlcNAc₂ structure by N-acetylglucosaminyltransferases III yields a so-called bisected N-glycan (see Figure 3). This structure has been implicated in greater antibody-dependent cellular cytotoxicity (ADCC) (Umana et al. 1999).

[0203] A host cell such as a yeast strain capable of producing glycoproteins with bisected N-glycans is engineered according to the invention, by introducing into

30 the host cell a GnTIII activity. Preferably, the host cell is transformed with a nucleic acid that encodes GnTIII (e.g., a mammalian such as the murine GnT III shown in Fig. 32) or a domain thereof having enzymatic activity, optionally fused

to a heterologous cell signal targeting peptide (e.g., using the libraries and associated methods of the invention.)

[0204] IgGs consist of two heavy-chains (V_H , C_{H1} , C_{H2} and C_{H3} in Figure 30), interconnected in the hinge region through three disulfide bridges, and two light chains (V_L , C_L in Figure 30). The light chains (domains V_L and C_L) are linked by another disulfide bridge to the C_{H1} portion of the heavy chain and together with the C_{H1} and V_H fragment make up the so-called Fab region. Antigens bind to the terminal portion of the Fab region. The Fc region of IgGs consists of the C_{H3} , the C_{H2} and the hinge region and is responsible for the exertion of so-called effector functions (see below).

[0205] The primary function of antibodies is binding to an antigen. However, unless binding to the antigen directly inactivates the antigen (such as in the case of bacterial toxins), mere binding is meaningless unless so-called effector-functions are triggered. Antibodies of the IgG subclass exert two major effector-functions: the activation of the complement system and induction of phagocytosis. The complement system consists of a complex group of serum proteins involved in controlling inflammatory events, in the activation of phagocytes and in the lytical destruction of cell membranes. Complement activation starts with binding of the C1 complex to the Fc portion of two IgGs in close proximity. C1 consists of one molecule, C1q, and two molecules, C1r and C1s. Phagocytosis is initiated through an interaction between the IgG's Fc fragment and Fc-gamma-receptors ($Fc\gamma RI$, II and III in Figure 30). Fc receptors are primarily expressed on the surface of effector cells of the immune system, in particular macrophages, monocytes, myeloid cells and dendritic cells.

[0206] The C_{H2} portion harbors a conserved N-glycosylation site at asparagine 297 (Asp297). The Asp297 N-glycans are highly heterogeneous and are known to affect Fc receptor binding and complement activation. Only a minority (i.e., about 15-20%) of IgGs bears a disialylated, and 3-10% have a monosialylated N-glycan (reviewed in Jefferis, R., Glycosylation of human IgG Antibodies. BioPharm, 2001). Interestingly, the minimal N-glycan structure shown to be necessary for fully functional antibodies capable of complement activation and Fc receptor binding is a pentasaccharide with terminal N-acetylglucosamine residues

(GlcNAc₂Man₃) (reviewed in Jefferis, R., Glycosylation of human IgG Antibodies. BioPharm, 2001). Antibodies with less than a GlcNAc₂Man₃ N-glycan or no N-glycosylation at Asp297 might still be able to bind an antigen but most likely will not activate the crucial downstream events such as phagocytosis and complement activation. In addition, antibodies with fungal-type N-glycans attached to Asp297 will in all likelihood solicit an immune-response in a mammalian organism which will render that antibody useless as a therapeutic glycoprotein.

B. Cloning And Expression Of GnTIII

The DNA fragment encoding part of the mouse GnTIII protein lacking the TM domain is PCR amplified from murine (or other mammalian) genomic DNA using forward 5'-TCCTGGCGCGCCTTCCCGAGAGAACTGGCCTCCCTC-3' and 5'-AATTAATTAACCCTAGCCCTCCGCTGTATCCAACCTTG-3' reversed primers. Those primers include AscI and PacI restriction sites that will be used for cloning into the vector suitable for the fusion with leader library.

The nucleic acid and amino acid sequence of murine GnTIII is shown in Fig. 32.

C. Cloning of immunoglobulin encoding sequences

[0207] Protocols for the cloning of the variable regions of antibodies, including primer sequences, have been published previously. Sources of antibodies and encoding genes can be, among others, *in vitro* immunized human B cells (see, e.g., Borreback, C.A. et al. (1988) *Proc. Natl. Acad. Sci. USA* 85, 3995-3999), peripheral blood lymphocytes or single human B cells (see, e.g., Lagerkvist, A.C. et al. (1995) *Biotechniques* 18, 862-869; and Terness, P. et al. (1997) *Hum. Immunol.* 56, 17-27) and transgenic mice containing human immunoglobulin loci, allowing the creation of hybridoma cell-lines.

[0208] Using standard recombinant DNA techniques, antibody-encoding nucleic acid sequences can be cloned. Sources for the genetic information encoding immunoglobulins of interest are typically total RNA preparations from cells of interest, such as blood lymphocytes or hybridoma cell lines. For example, by employing a PCR based protocol with specific primers, variable regions can be cloned via reverse transcription initiated from a sequence-specific primer

hybridizing to the IgG C_{H1} domain site and a second primer encoding amino acids 111-118 of the murine kappa constant region. The V_H and V_K encoding DNAs will then be amplified as previously published (see, e.g., Graziano, R.F. et al. (1995) *J Immunol.* 155(10): p. 4996-5002; Welschof, M. et al. (1995) *J. Immunol. Methods* 179, 203-214; and Orlandi, R. et al. (1988) *Proc. Natl. Acad. Sci. USA* 86: 3833). Cloning procedures for whole immunoglobulins (heavy and light chains have also been published (see, e.g., Buckel, P. et al. (1987) *Gene* 51:13-19; Recinos A 3rd et al. (1994) *Gene* 149: 385-386; (1995) *Gene* Jun 9;158(2):311-2; and Recinos A 3rd et al. (1994) *Gene* Nov 18;149(2):385-6). Additional protocols for the cloning and generation of antibody fragment and antibody expression constructs have been described in Antibody Engineering, R. Kontermann and S. Dübel (2001), Editors, Springer Verlag: Berlin Heidelberg New York.

[0209] Fungal expression plasmids encoding heavy and light chain of immunoglobulins have been described (see, e.g., Abdel-Salam, H.A. et al. (2001) *Appl. Microbiol. Biotechnol.* 56: 157-164; and Ogunjimi, A.A. et al. (1999) *Biotechnology Letters* 21: 561-567). One can thus generate expression plasmids harboring the constant regions of immunoglobulins. To facilitate the cloning of variable regions into these expression vectors, suitable restriction sites can be placed in close proximity to the termini of the variable regions. The constant regions can be constructed in such a way that the variable regions can be easily in-frame fused to them by a simple restriction-digest / ligation experiment. **Figure 31** shows a schematic overview of such an expression construct, designed in a very modular way, allowing easy exchange of promoters, transcriptional terminators, integration targeting domains and even selection markers.

[0210] As shown in **Figure 31**, V_L as well as V_H domains of choice can be easily cloned in-frame with C_L and the C_H regions, respectively. Initial integration is targeted to the *P. pastoris* AOX locus (or homologous locus in another fungal cell) and the methanol-inducible AOX promoter will drive expression. Alternatively, any other desired constitutive or inducible promoter cassette may be used. Thus, if desired, the 5' AOX and 3' AOX regions as well as transcriptional terminator (TT) fragments can be easily replaced with different TT, promoter and integration targeting domains to optimize expression. Initially the alpha-factor secretion

signal with the standard KEX protease site is employed to facilitate secretion of heavy and light chains. The properties of the expression vector may be further refined using standard techniques.

[0211] An Ig expression vector such as the one described above is introduced
5 into a host cell of the invention that expresses GnTIII, preferably in the Golgi apparatus of the host cell. The Ig molecules expressed in such a host cell comprise N-glycans having bisecting GlcNAcs.

EXAMPLE 7

10 **Cloning and expression of GnT-IV (UDP-GlcNAc:alpha-1,3-D-mannoside
beta-1,4-N-Acetylglucosaminyltransferase IV) and
GnT-V (beta 1-6-N-acetylglucosaminyltransferase)**

[0212] GnTIV-encoding cDNAs were isolated from bovine and human cells (Minowa, M.T. et al. (1998) *J. Biol. Chem.* 273 (19), 11556-11562; and
15 Yoshida, A. et al. (1999) *Glycobiology* 9 (3), 303-310. The DNA fragments encoding full length and a part of the human GnT-IV protein (Figure 33) lacking the TM domain are PCR amplified from the cDNA library using forward
5'-AATGAGATGAGGCTCCGCAATGGAACTG-3',
5'-CTGATTGCTTATCAACGAGAATTCCTTG-3', and reverse
20 5'-TGTTGGTTTCTCAGATGATCAGTTGGTG-3' primers, respectively. The resulting PCR products are cloned and sequenced.

[0213] Similarly, genes encoding GnT-V protein have been isolated from several mammalian species, including mouse. (See, e.g., Alvarez, K. et al. *Glycobiology* 12 (7), 389-394 (2002)). The DNA fragments encoding full length and a part of
25 the mouse GnT-V protein (Figure 34) lacking the TM domain are PCR amplified from the cDNA library using forward 5'-
AGAGAGAGATGGCTTTCTTTCTCCCTGG-3', 5'-
AAATCAAGTGGATGAAGGACATGTGGC-3', and reverse
5'-AGCGATGCTATAGGCAGTCTTTGCAGAG-3' primers, respectively. The
30 resulting PCR products are cloned and sequenced.

[0214] Nucleic acid fragments comprising sequences encoding GnT IV or V (or catalytically active fragments thereof) are cloned into appropriate expression vectors for expression, and preferably targeted expression, of these activities in an appropriate host cell according to the methods set forth herein.

[0215] REFERENCES

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15

What is Claimed is:

1. A method for producing a human-like glycoprotein in a non-human eukaryotic host cell comprising the step of diminishing or depleting the activity of one or more enzymes in the host cell that transfers a sugar residue to the 1,6 arm of
5 a lipid-linked oligosaccharide structure.
2. The method of claim 1, further comprising the step of introducing into the host cell at least one glycosidase activity.
3. The method of claim 2, wherein at least one glycosidase activity is a mannosidase activity.
- 10 4. The method of claim 1, further comprising producing an N-glycan.
5. The method of claim 4, wherein the N-glycan has a GlcNAcMan_xGlcNAc₂ structure wherein X is 3, 4 or 5.
6. The method of claim 5, further comprising the step of expressing within the host cell one or more enzyme activities, selected from glycosidase and
15 glycosyltransferase activities, to produce a GlcNAc₂Man₃GlcNAc₂ structure.
7. The method of claim 6, wherein the activity is selected from α -1,2 mannosidase, α -1,3 mannosidase and GnTII activities.
8. The method of claim 1, wherein at least one diminished or depleted enzyme is selected from the group consisting of an enzyme having dolichyl-P-
20 Man:Man₅GlcNAc₂-PP-dolichyl alpha-1,3 mannosyltransferase activity; an enzyme having dolichyl-P-Man:Man₆GlcNAc₂-PP-dolichyl alpha-1,2 mannosyltransferase activity and an enzyme having dolichyl-P-Man:Man₇GlcNAc₂-PP-dolichyl alpha-1,6 mannosyltransferase activity.

9. The method of claim 1, wherein the diminished or depleted enzyme has
25 dolichyl-P-Man:Man₅GlcNAc₂-PP-dolichyl alpha-1,3 mannosyltransferase
activity.
10. The method of claim 1, wherein the enzyme is diminished or depleted by
mutation of a host cell gene encoding the enzymatic activity.
11. The method of claim 10, wherein the mutation is a partial or total deletion
30 of a host cell gene encoding the enzymatic activity.
12. The method of claim 1, wherein the glycoprotein comprises *N*-glycans
having seven or fewer mannose residues.
13. The method of claim 1, wherein the glycoprotein comprises *N*-glycans
having three or fewer mannose residues.
- 35 14. The method of claim 1, wherein the glycoprotein comprises one or more
sugars selected from the group consisting of galactose, GlcNAc, sialic acid, and
fucose.
15. The method of claim 1, wherein the glycoprotein comprises at least one
oligosaccharide branch comprising the structure NeuNAc-Gal-GlcNAc-Man.
- 40 16. The method of claim 1, wherein the host is a lower eukaryotic cell.
17. The method of claim 1, wherein the host cell is selected from the group
consisting of *Pichia pastoris*, *Pichia finlandica*, *Pichia trehalophila*, *Pichia*
koclamae, *Pichia membranaefaciens*, *Pichia opuntiae*, *Pichia thermotolerans*,
Pichia salictaria, *Pichia guercuum*, *Pichia pijperi*, *Pichia stiptis*, *Pichia*
45 *methanolica*, *Pichia sp.*, *Saccharomyces cerevisiae*, *Saccharomyces sp.*, *Hansenula*
polymorpha, *Khuyveromyces sp.*, *Candida albicans*, *Aspergillus nidulans*,
Aspergillus niger, *Aspergillus oryzae*, *Trichoderma reesei*, *Chrysosporium*

lucknowense, *Fusarium sp.*, *Fusarium gramineum*, *Fusarium venenatum* and *Neurospora crassa*.

- 50 18. The method of claim 1, wherein the host cell is further deficient in expression of initiating α -1,6 mannosyltransferase activity.
19. The method of claim 18, wherein the host cell is an OCH1 mutant of *P. pastoris*.
20. The method of claim 1, wherein the host cell expresses GnTI and UDP-
55 GlcNAc transporter activities.
21. The method of claim 1, wherein the host cell expresses a UDP- or GDP-specific diphosphatase activity.
22. The method of claim 1, further comprising the step of isolating the glycoprotein from the host.
- 60 23. The method of claim 22, further comprising the step of subjecting the isolated glycoprotein to at least one further glycosylation reaction *in vitro*, subsequent to its isolation from the host.
24. The method of claim 1, further comprising the step of introducing into the host a nucleic acid molecule encoding one or more enzymes involved in the
65 production of GlcNAcMan₃GlcNAc₂ or GlcNAc₂Man₃GlcNAc₂.
25. The method of claim 24, wherein at least one of the enzymes has mannosidase activity.
26. The method of claim 25, wherein the enzyme has an α -1,2-mannosidase activity and is derived from mouse, human, *Lepidoptera*, *Aspergillus nidulans*, *C. elegans*, *D. melanogaster*, or *Bacillus sp.*
70

27. The method of claim 25, wherein the enzyme has an α -1,3-mannosidase activity.
28. The method of claim 24, wherein at least one enzyme has glycosyltransferase activity.
- 75 29. The method of claim 28, wherein the glycosyltransferase activity is selected from the group consisting of GnTI and GnTII.
30. The method of claim 24, wherein at least one enzyme is localized by forming a fusion protein between a catalytic domain of the enzyme and a cellular targeting signal peptide.
- 80 31. The method of claim 30, wherein the fusion protein is encoded by at least one genetic construct formed by the in-frame ligation of a DNA fragment encoding a cellular targeting signal peptide with a DNA fragment encoding a glycosylation enzyme or catalytically active fragment thereof.
32. The method of claim 31, wherein the encoded targeting signal peptide is
85 derived from a member of the group consisting of mannosyltransferases, diphosphotases, proteases, GnT I, GnT II, GnT III, GnT IV, GnT V, GnT VI, GalT, FT, and ST.
33. The method of claim 31, wherein the catalytic domain encodes a
90 consisting of GnT I, GnT II, GnT III, GnT IV, GnT V, GnT VI, GalT, Fucosyltransferase and ST, and wherein the catalytic domain has a pH optimum within 1.4 pH units of the average pH optimum of other representative enzymes in the organelle in which the enzyme is localized, or has optimal activity at a pH between 5.1 and 8.0.

- 95 34. The method of claim 31, wherein the nucleic acid molecule encodes one or more enzymes selected from the group consisting of UDP-GlcNAc transferase, UDP-galactosyltransferase, GDP-fucosyltransferase, CMP-sialyltransferase, UDP-GlcNAc transporter, UDP-galactose transporter, GDP-fucose transporter, CMP-sialic acid transporter, and nucleotide diphosphatases.
- 100 35. The method of claim 31, wherein the host expresses GnTI and UDP-GlcNAc transporter activities.
36. The method of claim 31, wherein the host expresses a UDP- or GDP-specific diphosphatase activity.
37. The method of claim 1, further comprising the step of introducing into a
105 host that is deficient in dolichyl-P-Man:Man5GlcNAc2-PP-dolichyl alpha-1,3 mannosyltransferase activity a nucleic acid molecule encoding one or more enzymes for production of a GlcNAcMan₄GlcNAc₂ carbohydrate structure.
38. The method of claim 1, further comprising the step of introducing into a host that is deficient in dolichyl-P-Man:Man6GlcNAc2-PP-dolichyl alpha-1,2
110 mannosyltransferase or dolichyl-P-Man:Man7GlcNAc2-PP-dolichyl alpha-1,6 mannosyltransferase activity a nucleic acid molecule encoding one or more enzymes for production of a GlcNAcMan₄GlcNAc₂ carbohydrate structure.
39. The method of claim 37 or 38, wherein the nucleic acid molecule encodes at least one enzyme selected from the group consisting of an α -1,2 mannosidase,
115 UDP GlcNAc transporter and GnT1.
40. The method of claim 39, further comprising the step of introducing into the deficient host cell a nucleic acid molecule encoding an α -1,3 or an α -1,2/ α -1,3

mannosidase activity for the conversion of the GlcNAc₁Man₄GlcNAc₂ structure to a GlcNAc₁Man₃GlcNAc₂ structure.

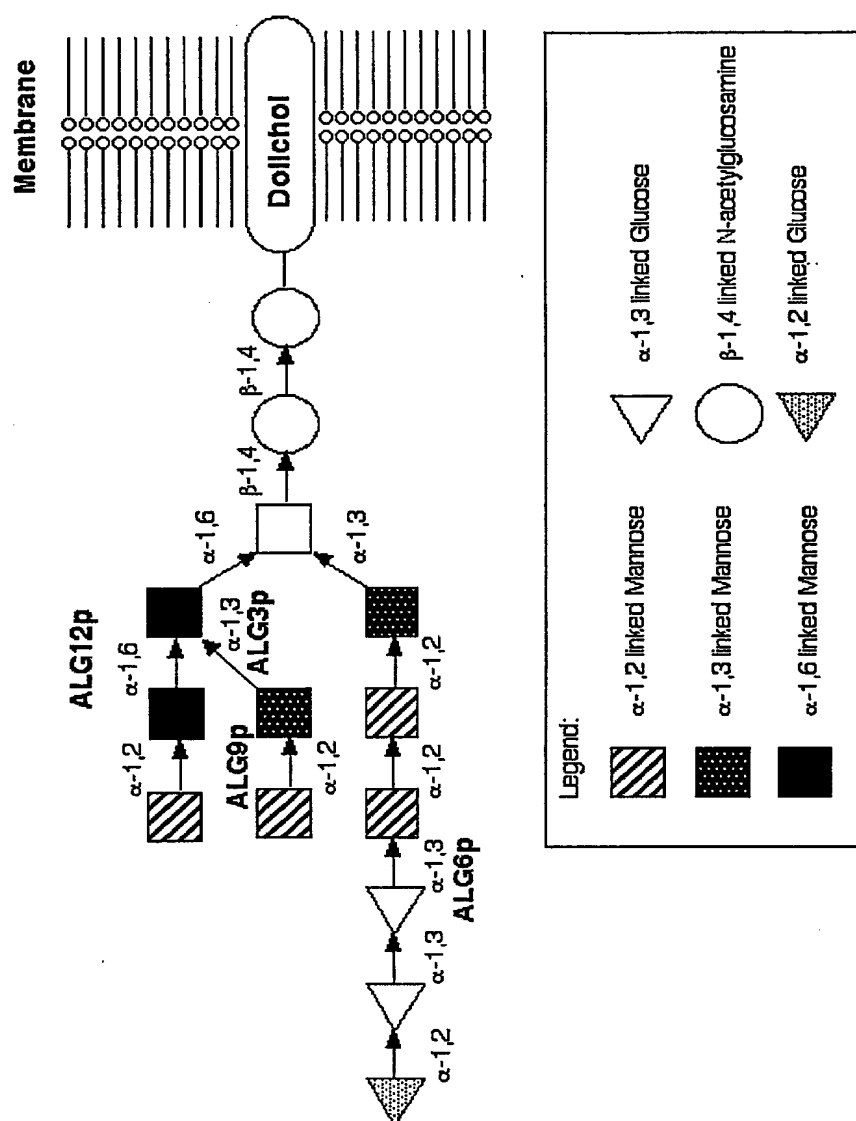
- 120 41. The method of claim 1, further comprising the step of introducing into the host a nucleic acid molecule encoding one or more enzymes for production of a GlcNAc₂Man₃GlcNAc₂ carbohydrate structure.
42. The method of claim 41, wherein at least one enzyme is GnTII.
43. The method of claim 1, further comprising the step of introducing into the
125 host cell at least one nucleic acid molecule encoding at least one mammalian glycosylation enzyme selected from the group consisting of a glycosyltransferase, fucosyltransferase, galactosyltransferase, N-acetylgalactosaminyltransferase, N-acetylglycosaminyltransferase and sulfotransferase.
44. The method of claim 1, comprising the step of transforming host cells with
130 a DNA library to produce a genetically mixed cell population expressing at least one glycosylation enzyme derived from the library, wherein the library comprises at least two different genetic constructs, at least one of which comprises a DNA fragment encoding a cellular targeting signal peptide ligated in-frame with a DNA fragment encoding a glycosylation enzyme or catalytically active fragment thereof.
- 135 45. A host cell produced by the method of claim 1 or 44.
46. A human-like glycoprotein produced by the method of claim 1 or 44.
47. A nucleic acid molecule comprising or consisting of at least **forty-five** consecutive nucleotide residues of Fig. 6 (*P. pastoris* *ALG 3* gene).
48. A vector comprising a nucleic acid molecule of claim 47.
- 140 49. A host cell comprising a nucleic acid molecule of claim 47.

50. A *P.pastoris* cell in which the sequences of Fig. 6 (*P. pastoris* *ALG 3* gene), are mutated whereby the glycosylation pattern of the cell is altered.
51. A method to enhance the degree of glucosylation of lipid-linked oligosaccharides comprising the step of increasing alpha-1,3 glucosyltransferase activity in a host cell.
52. A method to enhance the degree of glucosylation of lipid-linked oligosaccharides comprising decreasing the substrate specificity of oligosaccharyl transferase activity in a host cell.
53. A method for producing in a non-mammalian host cell an immunoglobulin polypeptide having an N-glycan comprising a bisecting GlcNAc, the method comprising the step of expressing in the host cell a GnTIII activity.
54. A non-mammalian host cell that produces an immunoglobulin having an N-glycan comprising a bisecting GlcNAc.
55. An immunoglobulin produced by the host cell of claim 54.
56. A method for producing in a non-human host cell a polypeptide having an N-glycan comprising a bisecting GlcNAc, the method comprising the step of expressing in the host cell a GnTIII activity.
57. A non-human host cell that produces a polypeptide having an N-glycan comprising a bisecting GlcNAc.
58. A polypeptide produced by the host cell of claim 57.
59. A method for producing a human-like glycoprotein in a non-human eukaryotic host cell comprising the step of diminishing or depleting from the host

cell an *alg* gene activity and introducing into the host cell at least one glycosidase activity.

- 165 60. A method for producing a human-like glycoprotein having an N-glycan comprising at least two GlcNAcs attached to a trimannose core.

FIGURE 1



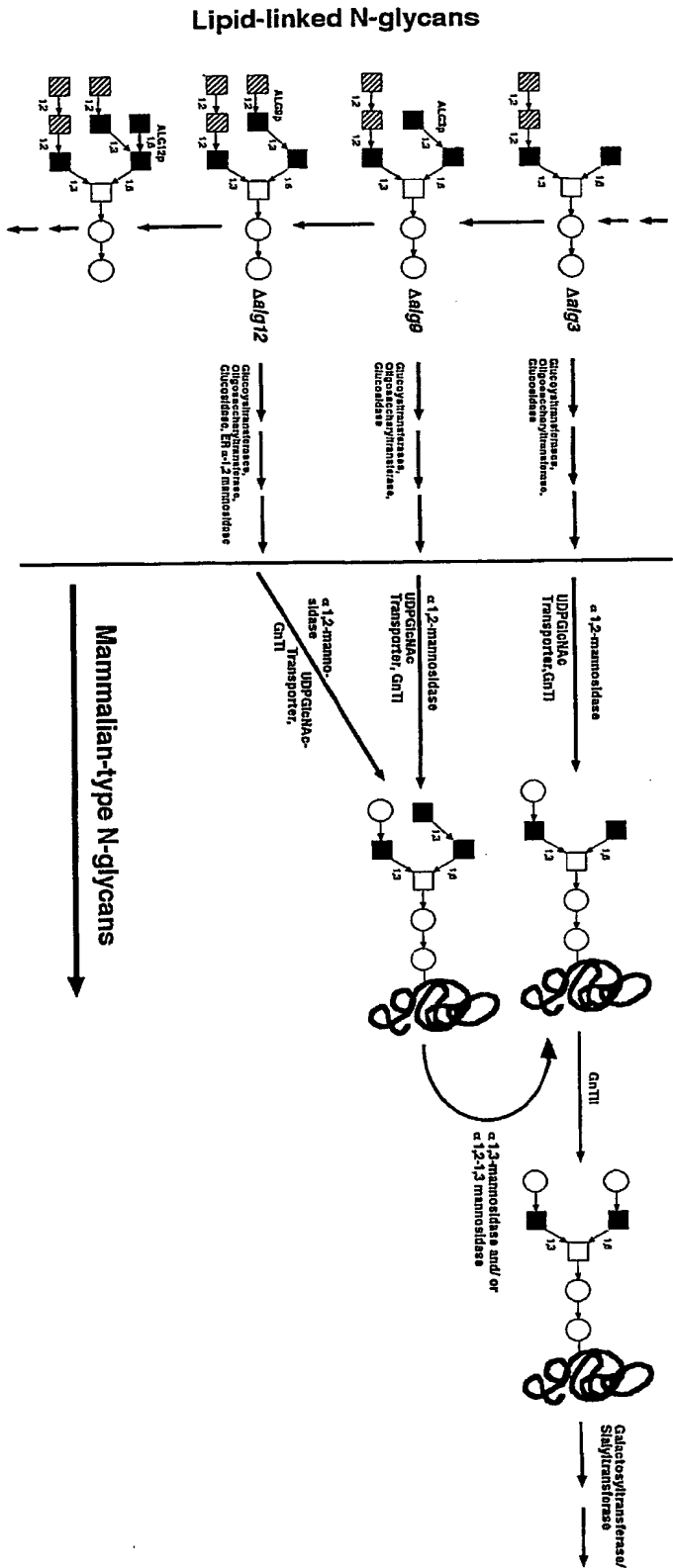


FIGURE 2

FIGURE 3

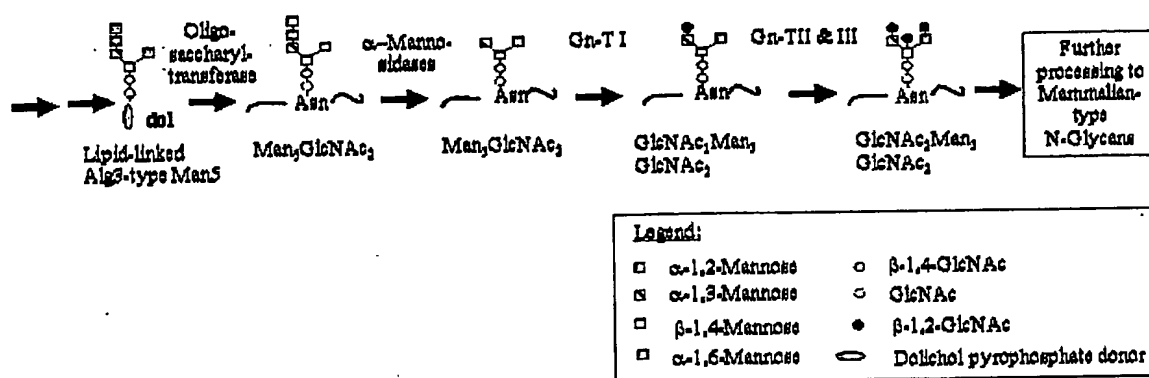


FIGURE 4 (sheet 1)

ALG3 Blast 05-22-01

Sequences producing significant alignments: (bits) Value

gi 586444 sp P38179 ALG3_YEAST	DOLICHYL-P-MAN:MAN(5)GLCNAC(...797	0.0
gi 3024226 sp Q92685 ALG3_HUMAN	DOLICHYL-P-MAN:MAN(5)GLCNAC...173	7e-43
gi 3024221 sp Q24332 NT56_DROVI	LETHAL(2)NEIGHBOUR OF TID P...145	3e-34
gi 3024222 sp Q27333 NT56_DROME	LETHAL(2)NEIGHBOUR OF TID P...121	3e-27
gi 10720153 sp P82149 NT53_DROME	LETHAL(2)NEIGHBOUR OF TID ...121	5e-27
gi 1707982 sp P40989 GLS2_YEAST	1,3-BETA-GLUCAN SYNTHASE CO... 32	2.8
gi 1346146 sp P38631 GLS1_YEAST	1,3-BETA-GLUCAN SYNTHASE CO... 31	6.6

Alignments

Yeast

>gi|586444|sp|P38179|ALG3_YEAST DOLICHYL-P-MAN:MAN(5)GLCNAC(2)-PP-DOLICHYL MANNOSYLTRANSFERASE
 (DOL-P-MAN DEPENDENT ALPHA(1-3)-MANNOSYLTRANSFERASE)
 (HM-1 KILLER TOXIN RESISTANCE PROTEIN)
 Length = 458

Score = 797 bits (2059), Expect = 0.0
 Identities = 422/458 (92%), Positives = 422/458 (92%)

Query: 1	MEGEQSPQGEKSLQRKQFVRPPLDLWQDLKDGVRVYVIFDCRANLIVMPLLLIFESMLCKI	60
Sbjct: 1	MEGEQSPQGEKSLQRKQFVRPPLDLWQDLKDGVRVYVIFDCRANLIVMPLLLIFESMLCKI	60
Query: 61	IIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQVSGGTGPLVYPAGHVLIYKMMYWLTEGM	120
Sbjct: 61	IIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQVSGGTGPLVYPAGHVLIYKMMYWLTEGM	120
Query: 121	DHVERGQVFFRYLYLLTLALQMACYLLHLPWCVVLAACLSKRLHSIYVLRFLNDCFTTL	180
Sbjct: 121	DHVERGQVFFRYLYLLTLALQMACYLLHLPWCVVLAACLSKRLHSIYVLRFLNDCFTTL	180
Query: 181	FMVVTVLGAIVASRCHQRPKLKKSALVISATYSMAVSIKMNALLYFPAMMISLFI LNDA	240
Sbjct: 181	FMVVTVLGAIVASRCHQRPKLKKSALVISATYSMAVSIKMNALLYFPAMMISLFI LNDA	240
Query: 241	NVILTLLDLVAMIAWQVAVAVPFLRSFPQQYLHCAFNFRKFMQWSINWQMMDEEAFND	300
Sbjct: 241	NVILTLLDLVAMIAWQVAVAVPFLRSFPQQYLHCAFNFRKFMQWSINWQMMDEEAFND	300
Query: 301	KRFXXXXXXXXXXXXXXXXX FVTRYPRILPDLWSSLCHPLRKNVNLNANPAKTIPFVLIASN	360
Sbjct: 301	KRFHLALLISHLIALTTLFVTRYPRILPDLWSSLCHPLRKNVNLNANPAKTIPFVLIASN	360
Query: 361	FIGVLFSSRLHYQFLSWYHWTLPILIFWSGMPFFVGPIWYVLHEWCWNSYPPNSQXXXXX	420
Sbjct: 361	FIGVLFSSRLHYQFLSWYHWTLPILIFWSGMPFFVGPIWYVLHEWCWNSYPPNSQ	420
Query: 421	XXXXXXXXXXXXXXXXXSGSVALAKSHLRTTSSMEKKLN	458
Sbjct: 421	LALNTVLLLLLALTQLSGSVALAKSHLRTTSSMEKKLN	458

FIGURE 4 (sheet 2)

Human

>gi|3024226|sp|Q92685|ALG3_HUMAN DOLICHYL-P-MAN:MAN(5)GLCNAC(2)-PP-DOLICHYL
MANNOSYLTRANSFERASE
(DOL-P-MAN DEPENDENT ALPHA(1-3)-MANNOSYLTRANSFERASE)
(NOT56-LIKE PROTEIN)
Length = 438

Score = 173 bits (439), Expect = 7e-43
Identities = 133/396 (33%), Positives = 195/396 (48%), Gaps = 28/396 (7%)

Query: 26 WQDLKDGVRVIFDCRANLIVMPLLLIFESMLCKIIKKVAYTEIDYKAYMEQIEMIQLD 85
WQ+ R ++ + R L+V L L E + +I +VAYTEID+KAYM ++E + ++
Sbjct: 29 WQER----RLLLREPRYTLLVAACLCLAEVGITFWVIHRVAYTEIDWKAYMAEVEGV-IN 83

Query: 86 GMLDYSQVSGGTGPLVYPAGHVLIYKMMYWLTEGMDHVERGQVFFRYLYLLTLALQMACY 145
G DY+Q+ G TGPLVYPAG V I+ +Y+ T + Q F LYL TL L Y
Sbjct: 84 GTYDYTQLOQDGTGPLVYPAGFVYIFMGLYYATSRGTDIRMAQNI FAVLYLATLLLVFLIY 143

Query: 146 Y-LLHLPPWC-VVLACL SKRLHSIYVLRLEFND CFTTFLMVVTVLGAIVASRCHQRPKLKK 203
+ +PP+ + C S R+HSI+VLRLEFND + + +L + QR
Sbjct: 144 HQTCKVPPFVFFFMCCASYRVHSIFVLRLEFNDP-----VAMVLLFLSINLLLAQRWGWG- 197

Query: 204 SLALVISATYSMAVSIKMNALLYFPAMMISLFILNDANVILTLLDLVAMIAWQVAVAVPF 263
+S+AVS+KMN LL+ P ++ L L L + A + QV + +PF
Sbjct: 198 -----CCFFSLAVSVKMNVLFLFAPGLLFLLLTQFGFRGALPKLGICAGL--QVVLGLPF 249

Query: 264 LRSFPQYLHCAFNFGRKFMYQWSINWQMMDEEAFNDKRFXXXXXXXXXXXXXXXXXVTRY 323
L P YL +F+ GR+F++ W++NW+ + E F + F + R+
Sbjct: 250 LLENPSGYLSRSFDLGRQFLFHWTVNWRFLPEALFLHRAFLALLTAHLTLLLLFALCRW 309

Query: 324 PRILPDWSSLCHPLRKNVAVLNANPAKTIPFVLIASNFIGVLFSSRLHYQFLSWYHWTLP 383
R + S L P ++ I L SNFIG+ FSRSLHYQF WY TLP
Sbjct: 310 HRTGESILSLRDPSKRKVPQPPLTPNQIVSTLFTSNFIGICFSRSLHYQFYVWYFHTLP 369

Query: 384 ILIF-----WSGMPFFVGPWIWVYLHEWCWNSYPPNS 414
L++ W + + + E WN+YP S
Sbjct: 370 YLLWAMPARWLTHLLRLLVLGLI--ELSWNTYPSTS 403

Drosophila Vi

>gi|3024221|sp|Q24332|NT56_DROVI LETHAL(2)NEIGHBOUR OF TID PROTEIN (NOT58)
Length = 526

Score = 145 bits (366), Expect = 3e-34
Identities = 103/273 (37%), Positives = 157/273 (56%), Gaps = 17/273 (6%)

Query: 33 VRYVIFDCRANLIVMPLLLIFESMLCKIIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQ 92
++Y+ F+ A IV L++L E+++ ++I++V YTEID+KAYM++ E L+G +YS
Sbjct: 34 IKYLAFEPAALPIVSVLIVLAEAVINVLVIQRPVYTEIDWKAYMQECEGF-LNGTTNYSL 92

Query: 93 VSGGTGPLVYPAGHVLIYKMMYWLTEGMDHVERGQVFFRYLYLLTLALQMACYLLH-LP 151
+ G TGPLVYPA V IY +Y+LT +V Q F +YLL + L + Y +P
Sbjct: 93 LRGDTGPLVYPAAFVYIYSGLYLTGQGTNVRLAQYIFACIYLLQMCILVLRLYTKSRKVP 152

Query: 152 PWCVVLA CL- SKRLHSIYVLRLEFND CFTTFLMVVTVLGAIVASRCHQRPKLKKSALVIS 210
P+ +VL+ S R+HSIYVLRLEFND L +L A + QR L S
Sbjct: 153 PYVLVLSAFTSYRIHSIYVLRLEFNDPVAIL-----LLYAALNLF LDQRWTLG-----S 200

Query: 211 ATYSMAVSIKMNALLYFPAMMISLFILNDANVILTLLDLVAMIAWQVAVAVPFLRSFPQQ 270
YS+AV +KMN + A + LF L + V+ TL+ L Q+ + PFLR+ P +
Sbjct: 201 ICYSLAVGVKMN--ILLFAPALLLFYLANLGVLRTLVQLTICAVLQLFIGAPFLRTHPME 258

FIGURE 4 (sheet 3)

Query: 271 YLHCAFNFGRKFMYQWSINWQMMDEEAFNDKRF 303
 YL +F+ GR F ++W++N++ + +E F + F
 Sbjct: 259 YLRGSFDLGRIFEHKWTVNYRFLSKELFEQREF 291

Score = 53.3 bits (127), Expect = 1e-06
 Identities = 31/62 (50%), Positives = 41/62 (66%), Gaps = 6/62 (9%)

Query: 352 IPFVLIASNFIGVLFSRSLHYQFLSWYHWTLPILIFWSGMPFFVGPWIYVLH--EWCWNS 409
 +PF L NFIGV +RSLHYQF WY +LP L+ WS P+ +G + +L E+CWN+
 Sbjct: 412 LPFFL--CNFIGVACARSLHYQFYIWFHSLPYLV-WS-TPYSLGVRYLILGLIEYCWN 467

Query: 410 YP 411
 YP
 Sbjct: 468 YP 469

Drosophila melanogaster

>gi|3024222|sp|Q27333|NT56_DROME LETHAL(2)NEIGHBOUR OF TID PROTEIN (NOT56)
 (NOT45)

Length = 510

Score = 121 bits (305), Expect = 3e-27
 Identities = 96/272 (35%), Positives = 154/272 (56%), Gaps = 17/272 (6%)

Query: 34 RYVIFDCRANLIVMPLLILFESMLCKIIKKVAYTEIDYKAYMEQIEMIQLDGMLDYSQV 93
 +Y++ + A IV ++L E ++ ++I++V YTEID+ AYM++ E L+G +YS +
 Sbjct: 36 KYLLLEPAALPIVGLFVLLAELVINVVVIQRPVYTEIDWVAYMQECEGF-LNGTTNYSLL 94

Query: 94 SGGTGPLVYPAGHVLIYKMMYWLTEGMDHVERGQVFFRYLYLLTLALQMACYLLH-LPP 152
 G TGPLVYPA V IY +Y++T +V Q F +YL LAL + Y +PP
 Sbjct: 95 RGDTPGLVYPAFVYIYSALYYVTSHTGNVRLAQYIFAGIYLLQLALVRLYSKSRKVPP 154

Query: 153 WCVVLACL-SKRLHSIYVLRLFNDCTTLFMVVTVLGAIVASRCHQRPKLKSLALVISA 211
 + +VL+ S R+HSIYVLRLFND + V +L A + +R L S
 Sbjct: 155 VVLVLSAFTSYRIHSIYVLRLFNDP-----VAVLLLYAALNLFDRRWTLG-----ST 202

Query: 212 TYSMAVSIKMNALLYFPAMMISLFILNDANVILTLLDLVAMIAWQVAVAVPFLRSFPQQY 271
 +S+AV +KMN + A + LF L + ++ T+L L Q+ + PFL + P +Y
 Sbjct: 203 FFS LAVGVKMN--ILLFAPALLLFYLANLGLLRTILQLAVCGVIQQLLGLAPFLLTHPVEY 260

Query: 272 LHCAFNFGRKFMYQWSINWQMMDEEAFNDKRF 303
 L +F+ GR F ++W++N++ + + F ++ F
 Sbjct: 261 LRGSFDLGRIFEHKWTVNYRFLSRDVFENRTF 292

Score = 49.4 bits (117), Expect = 2e-05
 Identities = 27/60 (45%), Positives = 35/60 (58%), Gaps = 2/60 (3%)

Query: 352 IPFVLIASNFIGVLFSRSLHYQFLSWYHWTLPILIFWSGMPFFVGPWIYVLHEWCWNSYP 411
 +PF L N +GV SRSLHYQF WY +LP L + + V + L E+CWN+YP
 Sbjct: 407 LPFFL--CNLVGVACSRSLHYQFYVWYFHSPLPYLAWSTPYSLGVRCLILGLIEYCWN 464

FIGURE 4 (sheet 4)

Matrix: BLOSUM62
Gap Penalties: Existence: 11, Extension: 1
Number of Hits to DB: 28883317
Number of Sequences: 96469
Number of extensions: 1107545
Number of successful extensions: 2870
Number of sequences better than 10.0: 16
Number of HSP's better than 10.0 without gapping: 5
Number of HSP's successfully gapped in prelim test: 11
Number of HSP's that attempted gapping in prelim test: 2839
Number of HSP's gapped (non-prelim): 23

length of query: 458
length of database: 35,174,128
effective HSP length: 45
effective length of query: 413
effective length of database: 30,833,023
effective search space: 12734038499
effective search space used: 12734038499
T: 11
A: 40
X1: 15 (7.1 bits)
X2: 38 (14.6 bits)
X3: 64 (24.7 bits)
S1: 40 (21.8 bits)
S2: 67 (30.4 bits)

FIGURE 5

S. cerevisiae ALG3

ATGGAAGGTGAACAGTCTCCGCAAGGTGAAAAGTCTCTGCAAAGGAAGC
AATTTGTCAGACCTCCGCTGGATCTGTGGCAGGATCTCAAGGACGGTGTG
CGCTACGTGATCTTCGATTGTAGGGCCAATCTTATCGTTATGCCCCTTTTG
ATTTTGTTCGAAAGCATGCTGTGCAAGATTATCATTAAAGAAGGTAGCTTAC
ACAGAGATCGATTACAAGGCGTACATGGAGCAGATCGAGATGATTCAGCT
CGATGGCATGCTGGACTACTCTCAGGTGAGTGGTGGAAACGGGCCCCGCTGG
TGTATCCAGCAGGCCACGTCTTGATCTACAAGATGATGTACTGGCTAACA
GAGGGAATGGACCACGTTGAGCGCGGGCAAGTGTTTTTCAGATACTTGTA
TCTCCTTACACTGGCGTTACAAATGGCGTGTTACTACCTTTTACATCTACC
ACCGTGGTGTGTGGTCTTGGCGTGCCTCTCTAAAAGATTGCACTCTATTTA
CGTGCTACGGTTATTCAATGATTGCTTCACTACTTTGTTTATGGTCGTCACG
GTTTTGGGGGCTATCGTGGCCAGCAGGTGCCATCAGCGCCCCAAATTAAA
GAAGTCCCTTGCGCTGGTGATCTCCGCAACATACAGTATGGCTGTGAGCA
TTAAGATGAATGCGCTGTTGTATTTCCCTGCAATGATGATTTCTCTATTCAT
CCTTAATGACGCGAACGTAATCCTTACTTTGTTGGATCTCGTTGCGATGAT
TGCATGGCAAGTCGCAGTTGCAGTGCCCTTCCTGCGCAGCTTTCCGCAACA
GTACCTGCATTGCGCTTTTAATTTTCGGCAGGAAGTTTATGTACCAATGGAG
TATCAATTGGCAAATGATGGATGAAGAGGCTTTCAATGATAAGAGGTTCC
ACTTGGCCCTTTTAATCAGCCACCTGATAGCGCTCACCACACTGTTCTGTC
CAAGATACCCTCGCATCCTGCCCCGATTATGGTCTTCCCTGTGCCATCCGC
TGAGGAAAAATGCAGTGCTCAATGCCAATCCCGCCAAGACTATTCCATTC
GTTCTAATCGCATCCAACCTTCATCGGCGTCCTATTTTCAAGGTCCCTCCAC
TACCAGTTTCTATCCTGGTATCACTGGACTTTGCCTATACTGATCTTTTGGT
CGGGAATGCCCTTCTTCGTTGGTCCCATTTGGTACGTCTTGCACGAGTGGT
GCTGGAATTCCTATCCACCAAACCTCACAAGCAAGCACGCTATTGTTGGCA
TTGAATACTGTTCTGTTGCTTCTATTGGCCTTGACGCAGCTATCTGGTTCGG
TCGCCCTCGCCAAAAGCCATCTTCGTACCACCAGCTCTATGGAAAAAAG
CTCAACTGA

S. cerevisiae Alg3p

MEGEQSPQGEKSLQRKQFVRPPLDLWQDLKDGVRVYVIFDCRANLIVMPLIL
FESMLCKIIKKVAYTEIDYKAYMEQIEMIQLDGM LDYSQVSGGTGPLVYPAG
HVLIYKMMYWLTEGMDHVERGQVFFRYLYLLTLALQMACYLLHLPPWCV
VLACLSKRLHSIYVLR LFNDCFTT LFMVVTVLGAIVASRCHQRPKLKSLALV
ISATYSMAVSIKMNALLYFPAMMISLFILNDANVILTLLDLVAMIAWQVAVA
VPFLRSFPQQYLHCAFNFGRKFMYQWSINWQMMDEEAFNDKRFHLALLISHL
IALTTLFVTRYPRILPDLWSSLCHPLRKNAVLNANPAKTIPFVLIASNFIGVLFS
RSLHYQFLSWYHWTLPI LFWSGMPFFVGPIWYVLHEWCWNSYPPNSQASTL
LLALNTVLLLLLALTQLSGSVALAKSHLRTTSSMEKKLN

FIGURE 6*P. pastoris* ALG3

ATGCCTCCGATAGAGCCAGCTGAAAGGCCAAAGCTTACGCTGAAAAATGT
TATCGGTGATCTAGTGGCTCTTATTCAAACGTTTTATTAAACCCAGATTTT
AGTGTCTTCGTTGCACCTCTTTTATGGTTAGCTGATTCCATTGTTATCAAGG
TGATCATTGGCACTGTTTCCTACACAGATATTGATTTTTCTTCATATATGCA
ACAAATCTTTAAAATTTCGACAAGGAGAATTAGATTATAGCAACATATTTG
GTGACACCGGTCCATTGGTTTACCCAGCCGGCCATGTTTCATGCTTACTCAG
TACTTTCGTGGTACAGTGATGGTGGAGAAGACGTCAGTTTCGTTCAACAA
GCATTTGGTTGGTTATACCTAGGTTGCTTGTTACTATCCATCAGCTCCTACT
TTTTCTCTGGCTTAGGGAAAATACCTCCGGTTTATTTTGTGGTTGGTAGC
GTCCAAGAGACTGCATTCAATATTTGTATTGAGACTCTTCAATGACTGTTT
AACAAACATTTTTGATGTTGGCAACTATAATCATCCTTCAACAAGCAAGTAG
CTGGAGGAAAGATGGCACAACCTATTCATTATCTGTCCCTGATGCTGCAG
ATACGTACAGTTTAGCCATCTCTGTAAAGATGAATGCGCTGCTATACCTCC
CAGCATTCCTACTACTCATATATCTCATTTGTGACGAAAATTTGATTAAAG
CCTTGGCACCTGTTCTAGTTTTGATATTGGTGCAAGTAGGAGTCGGTTATT
CGTTCATTTTACCGTTGCACTATGATGATCAGGCAAATGAAATTCGTTCTG
CCTACTTTAGACAGGCTTTTGACTTTAGTCGCCAATTTCTTTATAAGTGA
CGGTTAATTGGCGCTTTTGAGCCAAGAACTTTCAACAATGTCCATTTTC
ACCAGCTCCTGTTTGCTCTCCATATTATTACGTTAGTCTTGTTTCATCCTCAA
GTTCTCTCTCCTAAAAACATTGGAAAACCGCTTGGTAGATTTGTGTTGGA
CATTTTCAAATTTTGGGAAGCCAACCTTATCTCCAACCAATATTATCAACGA
CCCAGAAAGAAGCCCAGATTTTGTGTTACACCGTCATGGCTACTACCAACTT
AATAGGGGTGCTTTTTGCAAGATCTTTACACTACCAGTTCCTAAGCTGGTA
TGC GTTCTCTTGCCATATCTCCTTTACAAGGCTCGTCTGAACCTTTATAGCA
TCTATTATTGTTTATGCCGCTCACGAGTATTGCTGGTTGGTTTTCCAGCTA
CAGAACAAAGTTCCGCGTTGTTGGTATCTATCTTACTACTTATCCTGATTC
TCATTTTTACCAACGAACAGTTATTTCTTCTCAATCGGTCCCTGCAGAAA
AAAAGAATACATAA

P. pastoris Alg3p

MPPIEPAERPKLTLKNVIGDLVALIQNVLFNPDFS VFVAPLLWLADSIVIKVIIG
TVSYTDIDFSSYMQQIFKIRQGELDYSNIFGDTGPLVYPAGHVHAYSVLSWYS
DGGEDVSFVQQA FGWLYLGCLLLSISSYFFSGLGKIPPVYFVLLVASKRLHSIF
VLR LFNDCLTTFLMLATIILQQASSWRKDGTIPLSVPDAADTYSLAISVKMN
ALLYLP AFLLLIY LICDENLIKALAPVLVLILVQVGVGYSFILPLHYDDQANEIR
SAYFRQAFDFSRQFLYKWT VNWRFLSQETFN NVHFHQLLFALHIITLVLFILKF
LSPKNIGKPLGRFVL DIFKFWKPTLSPTNIINDPERSPDFVYTVMATTNLIGVLF
ARSLHYQFLSWYAFSLPYLLYKARLNFIASIIIVYAAHEYCWLVFPATEQSSAL
LVSILLILILIFTNEQLFPSQSVP AEK KNT

FIGURE 7 (sheet 1)

P. pastoris ALG3 BLAST

Sequences producing significant alignments: (bits) Value

gi 586444 sp P38179 ALG3 YEAST Dolichyl-P-Man:Man(5)GlcNAc(...228	2e-58
gi 12802365 gb AAK07848.1 AF309689_10 putative NOT-56 manno...212	8e-54
gi 984725 gb AAA75352.1 ORF 1	206
gi 7492702 pir T39084 probable mannosyltransferase - fissi...176	8e-43
gi 16226531 gb AAL16193.1 AF428424_1 At2g47760/F17A22.15 [A...164	2e-39
gi 25367230 pir B84919 Not56-like protein [imported] - Ara...164	3e-39
gi 25814791 emb CAB70171.2 Hypothetical protein K09E4.2 [C...161	2e-38
gi 17535001 ref NP_496950.1 Putative plasma membrane membr...160	3e-38
gi 1654000 emb CAA70220.1 Not56-like protein [Homo sapiens...155	2e-36
gi 13279206 gb AAH04313.1 AAH04313 Unknown (protein for IMA...154	2e-36
gi 22122365 ref NP_666051.1 hypothetical protein MGC36684 ...150	3e-35
gi 21292031 gb EAA04176.1 agCP3388 [Anopheles gambiae str...120	4e-26
gi 1780792 emb CAA71167.1 lethal(2)neighbour of tid [Droso...114	3e-24

Alignments

S. cerevisiae

Score = 228 bits (580), Expect = 2e-58

Identities = 154/429 (35%), Positives = 229/429 (53%), Gaps = 37/429 (8%)

Query: 9 RPKLTLKNVIGDLVALIQNVLFNPDFS VFVAPLLWLADSIKVIIGTVSYTDIDFSSYM 68
 RP L L DL ++ V+F+ ++ V PLL L +S++ K+II V+YT+ID+ +YM
 Sbjct: 20 RPPLDLWQ---DLKDGVRVYIFDCRANLIVMPLLLIFESMLCKIIKKVAYTEIDYKAYM 76

Query: 69 QQIFKIR-QGELDYSNIFGDTGPLVYPAGHVHAYSVLSWYSDGGEDVSFVQQAFGWLYLG 127
 +QI I+ G LDYS + G TGPLVYPAGHV Y ++ W ++G + V Q F +LYL
 Sbjct: 77 EQIEMIQLDGM LDYSQVSGGTGPLVYPAGHVLIYKMMYWLTEGMDHVERGQVFFRYLYLL 136

Query: 128 CLLLSISSYFFSGLGKIPPVYFVLLVASKRLHSIFVLRLEFNDCLTTFMLLATI---IILQ 184
 L L ++ Y+ L +PP VL SKRLHSI+VLRLEFNDCLTTFMLLATI---IILQ
 Sbjct: 137 TLALQMACYY---LLHLPPWCVLACL SKRLHSIYVLRLEFNDCLTTFMLVTVLGAIVAS 193

Query: 185 QASSWRKDGTTIPLSVPDAADTYS LAISVKMNXXXXXXXXXXXXXXXXXCDENLIKALAPXX 244
 + K ++ L + + TYS+A+S+KMN D N+I L
 Sbjct: 194 RCHQRPKLKKS LALVI---SATYSMAVSIKMNALLYFPAMMISLFILNDANVILTLLDLV 250

Query: 245 XXXXXXXXXXXXYSFILPLHYDDQANEIRSAYFRQAFDFSRQFLYKWTNVNRFLSQETFN NV 304
 F+ Y AF+F R+F+Y+W++NW+ + +E FN+
 Sbjct: 251 AMIAWQVAVAVPFL-----RSFPQOYLHCAFNFGRKFMYQWSINWQMMDDEEAFNDK 301

Query: 305 HFHQLLFALHIITL-VLFILKFLSPKNIGKPLGRFVLDFKFWKPTLSPTNIIN-DPERS 362
 FH L H+I L LF+ ++ R + D++ L ++N +P ++
 Sbjct: 302 RFHLALLISHLIALTTLFVTRY-----PRILPDLWSSLCHPLRKNVLANANPAKT 351

Query: 363 PDFVYTVMATTNLIGVLFARSLHYQFLSWYAFSLPYLLYKARLNFIASIIIVYAAHEYCWL 422
 F V+ +N IGVLF+RSLHYQFLSWY ++LP L++ + + F I Y HE+CW
 Sbjct: 352 IPF---VLIASNFVLFVSRSLHYQFLSWYHWTLPILIFWSGMPFFVGPVIWVYLHEWCWN 408

Query: 423 VFPATEQSS 431
 +P Q+S
 Sbjct: 409 SYPPNSQAS 417

FIGURE 7 (sheet 2)

Neurospora crassa

Score = 212 bits (540), Expect = 8e-54

Identities = 140/400 (35%), Positives = 212/400 (53%), Gaps = 29/400 (7%)

Query: 35 SVFVAPLLWLADSIKVIIGTVSYTDIDFSSYMQQIFKIRQGELDYSNIFGDTGPLVYP 94
 S + P L+L D+++ +II V YT+ID+++YM+Q+ +I GE DY+ + G TGPLVYP
 Sbjct: 33 SKLIPPALFLVDALLCGLIWKVPYTEIDWAAYMEQVSQILSGERDYTKVRGGTGPLVYP 92

Query: 95 AGHVHAYSVLSWYSDGGEDVSFVQQAAGWLYLGCLLLSISSYFFSGLGKIPPVYFVLLVA 154
 A HV+ Y+ L +D G ++ QQ F LY+ L + + Y+ K PP F LL
 Sbjct: 93 AAHVYIYTGLYHLTDEGRNILLAAQQLFAGLYMVT LAVVMGCYW---QAKAPPYLFPLLT 149

Query: 155 SKRLHSIFVLRFLNDCLTTFMLLATIILQQASSWRKDGTTIPLSVPDAADTYSLAISVK 214
 SKRLHSIFVLR FNDC + I Q+ +W+ A Y+L + VK
 Sbjct: 150 SKRLHSIFVLRFCFNDCCFAVLFLWLAIFFFQR-RNWQA-----GALLYTLGLGVK 197

Query: 215 MNXXXXXXXXXXXXXXXXXCDENLIKALAPXXXXXXXXXXXXXYSFILPLHYDDQANEIRSAY 274
 M + + L F+ HY + Y
 Sbjct: 198 MTLSSLPVAVGIVLFLGSG-SFVTTLQLVATMGLVQILIGVPFL--AHYPTE-----Y 247

Query: 275 FRQAFDFSQRFLYKWTNVNWRFLSQETFNNVHFHQLLFALHIITLVLFILKFLSPKNIGK 333
 +AF+ SRQF +KWTNVNRF+ +E F + F L ALH++ L +FI +++ P K
 Sbjct: 248 LSRAFELSRQFFFKWTNVNWRFGVEIFLSKGFALTLLALHVLVLGIFITTRWIKPAR--K 305

Query: 334 PLGRFVLDFKFWKPTLS-PTNIINDPERSPDFVYTVMATTNLIGVLFARSLHYQFLSWY 392
 L + + + KP L+ P + + +P ++ T + + N +G+LFARSLHYQF ++
 Sbjct: 306 SLVQLISPVLLAGKPPLTVPEHRAAARDVTPRYIMTTILSANAVGLLFARSLHYQFYAYV 365

Query: 393 AFSLPYLLYKARLNFIASIIVYAAHEYCWLVPFATEQSSA 432
 A+S P+LL++A L+ + +++A HE+ W VFP+T SSA
 Sbjct: 366 AWSTPFLLRAGLHPVLVYLLWAVHEWAVNVPSTPASSA 405

Schizosaccharomyces pombe

Score = 176 bits (445), Expect = 8e-43

Identities = 132/390 (33%), Positives = 194/390 (49%), Gaps = 35/390 (8%)

Query: 42 LWLADSIKVIIGTVSYTDIDFSSYMQQIFKIRQGELDYSNIFGDTGPLVYPAGHVHAY 101
 L L + + II V YT+ID+ +YM+Q+ GE DY ++ G TGPLVYP GHV Y
 Sbjct: 30 LLLLEIPFVFAIISKVPYTEIDWIAVMEQVNSFLLGERDYKSLVGCTGPLVYPGGHVFLY 89

Query: 102 SVLSWYSDGGEDVSFVQQAAGWLYLGCLLLSISSYFFSGLGKIPPVYFVLLVASKRLHSI 161
 ++L + +DGG ++ Q F ++Y + +I Y F + + P +VLL+ SKRLHSI
 Sbjct: 90 TLLYYLTDGGTNIVRAQYIFAFVYW--ITTAIVGYLFK-IVRAPFYIYVLLILSKRLHSI 146

Query: 162 FVLRLFNDCLTTFMLLATIILQQASSWRKDGTTIPLSVPDAADTYSLAISVKMNXXXXX 221
 F+LRLFNDC + L + I+ W + A+ S+A SVKM+
 Sbjct: 147 FILRLFNDCGFNS-LFSSLFILSSCKKKWVR-----ASILLSVACSVKMSSLLYV 194

Query: 222 XXXXXXXXXXXXCDENLIKALAPXXXXXXXXXXXXXYSFILPLHYDDQANEIRSAYFRQAFDF 281
 L++ L P + + + +Y+ QAFDF
 Sbjct: 195 PAYLVL-----LLQILGPCKTWMHIFVIIIVQILFSIPF----LAFWSYWTQAFDF 242

Query: 282 SRQFLYKWTNVNWRFLSQETFNNVHFHQLLFALHIITLVLFILKFLSPKNIGKPLGRFVLD 341
 R F YKWTNVNRF+ + F + F + LH+ LV F K + + P
 Sbjct: 243 GRAFDYKWTNVNRFIPRSIFESTSFSTSILFLHVALLVAFCKHWNKLSRATP----- 295

Query: 342 IFKFWKPTLSPTNIINDPERSPDFVYTVMATTNLIGVLFARSLHYQFLSWYAFSLPYLLY 401
 F L+ + +P+F++T +AT+NLIG+L ARSLHYQF +W+A+ PYL Y

FIGURE 7 (sheet 3)

Sbjct: 296 -FAMVNSMLTLKPLPKLQLATPNFIFTALATSNLIGILCARSLHYQFYAWFAWYSPYLCY 354

Query: 402 KARLNFIASIIIVYAAHEYCWLVPATEQSS 431
+A I ++ EY W VFP+T+ SS

Sbjct: 355 QASFPAPIVIGLWMLQEYAWNVPSTKLSS 384
Arabidopsis thaliana

Score = 164 bits (415), Expect = 2e-39
Identities = 131/391 (33%), Positives = 194/391 (49%), Gaps = 29/391 (7%)

Query: 42 LWLADSIIVIKVIIGTVSYTDIDFSSYMQQIFKIRQGELDYSNIFGDTGPLVYPAGHVHAY 101
L LAD+I++ +II V YT ID+ +YM Q+ GE DY N+ GDTGPLVYPAG ++ Y
Sbjct: 39 LILADAILVALIIAYVPYTKIDWDAYMSQVSGFLGGERDYGNLKGDTGPLVYPAGFLYVY 98

Query: 102 SVLSWYSDGGEDVSFVQQAAGWLYLGLLLSISSYFFSGLGKIPPVYFVLLVASKRLHSI 161
S + + G +V Q FG LY+ L + + Y + + +P LL SKR+HSI
Sbjct: 99 SAVQNLTTGG--EVYPAQILFGVLYIVNLGIVLIIYVKTDV--VPWWALSLCLSKRIHSI 154

Query: 162 FVLRLFNDCLTTFMLLATIIILQQASSWRKDGTTIPLSVPDAAADTYSLSVSKMNXXXXX 221
FVLRLFNDC L+ A++ + +RK + + +S A+SVKMN
Sbjct: 155 FVLRLFNDCFAMTLHASMALFL----YRKWHLGMLV-----FSGAVSVKMNVLLEYA 202

Query: 222 XXXXXXXXXXXCDENLIKALAPXXXXXXXXXXXXXYSFILPLHYDDQANEIRSAYFRQAFDF 281
N+I ++ F++ +Y AFD
Sbjct: 203 PTLRLRLKAM--NIIGVVSALAGAALQILVGLPFLITYPV-----SYIANAFDL 251

Query: 282 SRQFLYKWTNVNWRFLSQETFNNVHFHQLLFALHIITLVLFILKFLSPKNIGKPLGRFVLD 341
R F++ W+VN++F+ + F + F L H+ LV F + K+ G +G
Sbjct: 252 GRVFIHFWSVNFKFVPERVSVFVSKEFAVCLLIAHLFLLVAFV-NYKWCKHEGGIIGFMRSR 310

Query: 342 IFKFWKP-TLSPTNIINDPERSPDFVYTVMATTNLIGVLFARSLHYQFLSWYAFSLPYLL 400
F P +LS +++ + + V T M N IG++FARSLHYQF SWY +SLPYLL
Sbjct: 311 HFFLTLPSSLSFSDVSASRIITKEHVVTAMFVGNFIGIVFARSLHYQFYSWYFSLPYLL 370

Query: 401 YKARLNFIASIIIVYAAHEYCWLVPATEQSS 431
++ +I++ E CW V+P+T SS
Sbjct: 371 WRTFPPTWLRRLIMFLGIELCWNVYPSTPSSS 401

FIGURE 8

K. lactis ALG3

TTTGTTTACAAGCTGATACCAACGAACATGAATACACCGGCAGGTTTACT
GAAGATTGGCAAAGCTAACCTTTTACATCCTTTTACCGATGCTGTATTCAG
TGCGATGAGAGTAAACGCAGAACAAATTGCATACATTTTACTTGTTACCA
ATTACATTGGAGTACTATTTGCTCGATCATTACACTACCAATTCCTATCTT
GGTACCATTTGGACGTTACCAAGTACTATTGAATTGGGCCAATGTTCCGTATC
CGCTATGTGTGCTATGGTACCTAACACATGAGTGGTGCTGGAACAGCTAT
CCGCCAAACGCTACTGCATCCACACTGCTACACGCGTGTAACACATACTG
TTATTGGCTGTATTCTTAAGAGGACCCGCAAACCTCGAAAAGTGGTGATAA
CGAAACAACACACGAGAAAGCTGAG

K. lactis Alg3p

FVYKLIPTNMNTPAGLLKIGKANLLHPFTDAVFSAMRVNAEQIAYILLVTNYI
GVLFARSLHYQFLSWYHWTLPVLLNWANVPYPLCVLWYLTHEWCWNSYPP
NATASTLLHACNTYCYWLYSZEDPQTRKVVITKQHTRKL

FIGURE 9

K. lactis ALG3 BLAST

Score	E		(bits)	Value
Sequences producing significant alignments:				
gi 586444 sp P38179 ALG3 YEAST		Dolichyl-P-Man:Man(5)GlcNAc(...)	125	1e-28
gi 984725 gb AAA75352.1		ORF 1	94	4e-19
gi 16226531 gb AAL16193.1 AF428424.1		At2g47760/F17A22.15 [A...	72	1e-12
gi 25367230 pir B84919		Not56-like protein [imported] - Ara...	72	1e-12
gi 21292031 gb EAA04176.1		agCP3388 [Anopheles gambiae str....	69	2e-11
gi 20892051 ref XP_148657.1		similar to Lethal(2)neighbour ...	65	2e-10

Alignments

S. cerevisiae

Score = 125 bits (314), Expect = 1e-28
 Identities = 60/120 (50%), Positives = 83/120 (69%), Gaps = 1/120 (0%)
 Frame = +3

Query: 66 ANLLHPFT-DAVFSAMRVNAEQIAYILLVTNYIGVLFARSLHYQFLSWYHWTLPVLLNWA 242
 ++L HP +AV +A A+ I ++L+ +N+IGVLF+RSLHYQFLSWYHWTL+L+ W+
 Sbjct: 332 SSLCHPLRKNVAVLNANP--AKTIPFVLIASNFIGVLF+SRSLHYQFLSWYHWTL+L+ W+
 Query: 243 NVPPYPLCVLWYLTHEWCWNSYPPNATASTLLHACNTYCYWLYS*EDPQTRKVVITKQHTR 422
 +P+ + +WY+ HEWCWNSYPPN+ ASTLL A NT L + + V + K H R
 Sbjct: 390 GMPFFVGPVWYVLEWCWNSYPPNSQASTLLALNTVLLLLLA-LTQLSGSVALAKSHLR 448

A. thaliana

Score = 72.0 bits (175), Expect = 1e-12
 Identities = 42/107 (39%), Positives = 57/107 (53%), Gaps = 3/107 (2%)
 Frame = +3

Query: 84 FTDVAVFSAMRVNAEQIAYILLVTNYIGVLFARSLHYQFLSWYHWTLPVLLNWANVPYPLC 263
 F+D S + + E + + V N+IG++FARSLHYQF SWY ++LP LL P L
 Sbjct: 322 FSDVSASRI-ITKEHVVTAMFVGNFIGIVFARSLHYQFYSWYFYSLLPYLLWRTFPPTWLR 380
 Query: 264 VLWYLTHEWCWNSYPPNATASTL---LHACNTYCYWLYS*EDPQTRK 395
 ++ +L E CWN YP ++S L LH WL DP K
 Sbjct: 381 LIMFLGIELCWNVPSTPSSSGLLLCLHLIILVGLWLAPSVDPYQLK 427

FIGURE 10

S. cerevisiae ALG9

ATGAATTGCAAGGCGGTAACCATTAGTTTATTACTGTTGTTATTTTAAACAAGAGT
ATATATTCAGCCGACATTCTCGTTAATTTTCAGATTGCGATGAAACTTTTAAATTATT
GGGAACCATTAAATTTATTGGTACGTGGATTGTTGTAACAAACCTGGGAATATTC
ACCCGAGTATTCTATTAGATCATGGGCTTTCTTATTACCTTTTACTGTATTCTTTA
TCCAGTAAACAAATTTACTGACCTAGAAAGTCATTGGAACCTTTTCATCACAAGA
GCATGCTTAGGCTTTTTAGTTTTATCATGGAATTTAAACTACATCGTGAAATTGC
AGGCAGCTTGGCATTGCAAATCGCAAATATTTGGATTATTTTCCAATTGTTTAAATC
CGGGCTGGTCCATGCATCTGTGGAATTATTGCCCTTCTGCCGTTGCCATGTTGTTG
TATGTAGGTGCCACCAGACACTCTCTACGCTATCTGTCCACTGGGTCTACTTCTAA
CTTTACGAAAAGTTTAGCGTACAATTTCTGGCTAGTATACTAGGCTGGCCATTTG
TTTTAATTTTAAAGCTTGCCATTATGTTTACATTACCTTTTCAACCATAGAATTATT
CTACCATCAGAACCGCATTGACTGCTGTTTGATAATTTTCATTGACTGCATTTGCT
GTGATTGTCACTGACAGTATATTTTACGGGAAGCTTGCTCCTGTATCATGGAACA
TCTTATTTTACAATGTCATTAATGCAAGTGAGGAATCTGGCCCAAATATTTTCGGG
GTTGAGCCATGGTACTACTATCCACTAAATTTGTTACTGAATTTCCCACTGCCTGT
GCTAGTTTTAGCTATTTTGGGAATTTTCCATTTGAGATTATGGCCATTATGGGCAT
CATTATTCACATGGATTGCCGTTTTCACTCAACAACCTCACAAAGAGGAAAGATT
TCTCTATCCAATTTACGGGTAAATAACTTTGAGTGCAAGTATCGCCTTTTACAAAG
TGTTGAATCTATTCAATAGAAAGCCGATTCTTAAAAAAGGTATAAAGTTGTCACT
TTTATTAATTGTTGCAGGCCAGGCAATGTCACGGATAGTGGCTTTGGTGAACAAT
TACACAGCTCCTATAGCCGTCTACGAGCAATTTTCTTCACTAAATCAAGGTGGTG
TGAAGGCACCGGTAGTGAATGTATGTACGGGACGTGAATGGTATCACTTCCCAAG
TTCTTTTCTGCTGCCAGATAATCATAGGCTAAAATTTGTTAAATCTGGATTGATG
GTCTTCTTCCAGGTGATTTTCCAGAGAGTGTTCTATTTTCAAAAAGATTAGAAGT
TTACCTAAGGGAATGAATAACAAGAATATATATGATACCGGTAAAGAGTGGCCG
ATCACTAGATGTGATTATTTTATTGACATCGTCGCCCAATAAATTTAACAAAAG
ACGTTTTCAACCTCTACATCTGATGGATAACTGGAATAAGCTGGCATGTGCTGC
ATTCATCGACCGTGAAAATTCTAAGATTTTGGGTAGAGCATTTTACGTACCGGAG
CCAATCAACCGAATCATGCAAATAGTTTTACCAAAACAATGGAATCAAGTGTACG
GTGTTTCGTTACATTGATTACTGTTTGTGTTGAAAAACCAACTGAGACTACTAATTGA

S. cerevisiae Alg9p

MNCKAVTISLLLLLFLTRVYIQPTFSLISDCDETFNYWEPLNLLVRGFGKQTWEYSPE
YSIRSWAFLLPFYCILYPVNKFTDLESHWNFFITRACLGFFSFIMEFKLHREIAGSLALQ
IANIWIIFQLFNPWFHASVELLPASAVAMLLYVGATRHSRLYLSTGSTSNFTKSLAYN
FLASILGWPFVLILSLPLCLHYLFNHRISTIRTAFCCLIFSLTAFVIVTDSIFYGKLAP
VSWNLFYNVINASEESGPNIFGVPEPWYYYPLNLLNFPLPVLVLAILGIFHLRLWPLW
ASLFTWIAVFTQOPHKEERFLYPIYGLITLSASIAFYKVLNLFNRKPILKKGIKLSVLLI
VAGQAMSRIVALVNNYTAPIAVYEQFSSLNQGQGVKAPVVNVCTGREWYHFPSSFLLP
DNHRLKFVKSGFDGLLPDGFPESGSIFKKIRTLPGKMNNKNIYDTGKEWPITRCDYFI
DIVAPINLTKDVFNPLHMDNWNKLACAAFIDGENSKILGRAFYVPEPINRIMQIVLP
KQWNQVYGVRYIDYCLFEKPTETTN

FIGURE 11

P. pastoris ALG9

TGGCCTTCCTGTCTGCTCGATACTTCCTTTTACAGTAACCAACATACATGTT
CTCCAACATGCTCTTGTATGTATTGGCCTATTCTATCTTGAGACTTGATATC
AACCTTCTATGGTATTATTTTCAGACTGTGATGAAGTGTTCAACTACTGGGA
GCCACTCAACTTCATGCTTAGAGGGTTTGGAAAACAGACTTGGGAGTATT
CTCCAGAGTATGCCATCCGATCTTGGTCCTATCTAGTGCCACTTTGGATAG
CAGGCTATCCACCATTGTTCCCTGGATATCCCTTCTTACTACTTTTTCTACTT
TTTCAGACTACTGCTGGTTATTTTTTTCATTGGTTGCAGAAGTCAAGTTGTA
CCATAGTTTGAAGAAAAATGTCAGCAGTAAGATCAGTTTCTGGTACCTTCT
ATTTACAACCGTTGCTCCAGGAATGTCTCATAGCACGATAGCCTTATTACC
ATCCTCTTTTGCTATGGTTTGTCACTTTTGCCATTAGATACGTCATTGAT
TACCTACAATTACCAACATTAATGCGCACAATCAGAGAGACTGCTGCCAT
CTCACCAGCTCACAAACAACAACACTAGCCAACCTCTCTC

P. pastoris Alg9p

WPSCLLDTSFYSNQHTCSPTCSCMYWPILSZDLISTFYGIISDCDEVFNYWEPL
NFMLRGFGKQTWEYSPEYAIRSWSYLVPLWIAGYPPLFLDIPSYFFFYFFRLLL
VIFSLVAEVKLYHSLKKNVSSKISFWYLLFTTVAPGMSHSTIALLPSSFAMVCH
TFAIRYVIDYLQLPTLMRTIRETAAISPAHKQQLANSL

FIGURE 12 (sheet 1)

P. pastoris ALG9 BLAST

Score E
Sequences producing significant alignments: (bits) Value

qi 6324110 ref NP_014180.1	catalyzes the transfer of manno...	131	1e-29
qi 21296668 gb EAA08813.1	agCP7810 [Anopheles gambiae str...	110	2e-23
qi 7019765 emb CAB75773.1	putative mannosyltransferase inv...	104	1e-21
qi 26341066 dbj BAC34195.1	unnamed protein product [Mus mu...	99	4e-20
qi 16551378 gb AAL25798.1	DIBD1 [Homo sapiens]	99	4e-20
qi 19527202 ref NP_598742.1	RIKEN cDNA 8230402H15 [Mus mus...	99	4e-20
qi 12053349 emb CAB66861.1	hypothetical protein [Homo sapi...	99	4e-20

Alignments

S. cerevisiae

Score = 131 bits (329), Expect = 1e-29
Identities = 62/141 (43%), Positives = 91/141 (64%), Gaps = 1/141 (0%)
Frame = +2

Query: 200 ISTFYGIISDCDEVFNWYWEPLNFMLRGFGKQTWEYSPEYAIRSWSYLVPLWIAGYP-PLF 376
I + +ISDCDE FNYWEPLN ++RGFGKQTWEYSPEY+IRSW++L+P + YP F
Sbjct: 21 IQPTFSLISDCDETFNYWEPLNLLVRGFGKQTWEYSPEYSIRSWAFLLPFYCILYPVNKF 80

Query: 377 LDIPSXXXXXXXXRLLLVIFSLVAEVKLYHSLKKNVSSKISFWYLLFTTVAPGMSHSTIAL 556
D+ S R L FS + E KL+ + +++ +I+ +++F PG H+++ L
Sbjct: 81 TDLESHWNFFITRACLGFFSFIMEFKLHREIAGSLALQIANIWIIFQLFNPGWGFHASVEL 140

Query: 557 LPSSFAMVCHTFAIRYVIDYL 619
LPS+ AM+ + A R+ + YL
Sbjct: 141 LPSAVAMLLYVGATRHSRLYL 161

Anopheles gambiae

Score = 110 bits (274), Expect = 2e-23
Identities = 58/130 (44%), Positives = 79/130 (60%), Gaps = 3/130 (2%)
Frame = +2

Query: 197 LISTFYGIISDCDEVFNWYWEPLNFMLRGFGKQTWEYSPEYAIRSWSYLVPLWIAGYPPPLF 376
L S Y IISDCDE +NYWEPL+++L+G G QTWEYSPE+A+RS+SY LW+ G P
Sbjct: 34 LQSALYSIISDCDETYNYWEPLHYLLKGKGFQTWEYSPEFALRSYSY---LWLHGLPAKV 90

Query: 377 LDIPS---XXXXXXXXRLLLVIFSLVAEVKLYHSLKKNVSSKISFWYLLFTTVAPGMSHST 547
L + + R LL + + E +LY L + ++ +LLF + GM S+
Sbjct: 91 LQMTDNGVLIFYFVRCLLAVTCALLEYRLYLGRKCGGGVASLWLLFQLTSAGMFISS 150

Query: 548 IALLPSSFAM 577
ALLPSSF+M
Sbjct: 151 AALLPSSFSM 160

FIGURE 12 (sheet 2)

S. pombe

Score = 104 bits (260), Expect = 1e-21
 Identities = 58/157 (36%), Positives = 85/157 (54%)
 Frame = +2

Query: 197 LISTFYGIISDCDEVFNWYWEPLNFMLRGFGKQTWEYSPEYAIRSWSYLVPLWIAGYPPLF 376
 L S + +I DCDEV+NYWEPL+++L G+G QTWEYSPEYAIRSW Y+ + G+
 Sbjct: 26 LTSASFRVIDDCDEVYNYWEPLHYLLYGYGLQTWEYSPEYAIRSWFYIALHAVPGFLARG 85

Query: 377 LDIPSXXXXXXXXRLLLVIFSLVAEVKLYHSLKKNVSSKISFWYLLFTTVAPGMSHSTIAL 556
 L + R +L FS E L ++ +N + ++ V GM ++ +
 Sbjct: 86 LGLSRLHVFYFIRGVLACFSAFCECNLILAVARNFNRAVALHLTSVLFVNSGMWSASTSF 145

Query: 557 LPSSFAMVCHTFAIRYVIDYLQPLTLMRTIRETAAIS 667
 LPSSFAM T A+ L P+ RT++ + I+
 Sbjct: 146 LPSSFAMNMVTLALS---AQLSPSTKRTVKVVSFIT 179

M. musculus

Score = 99.4 bits (246), Expect = 4e-20
 Identities = 57/143 (39%), Positives = 76/143 (53%), Gaps = 1/143 (0%)
 Frame = +2

Query: 152 SPTCSCMYWPILS*DLISTFYGIISDCDEVFNWYWEPLNFMLRGFGKQTWEYSPEYAIRSW 331
 +P S + +LS L + ISDCDE FNYWEP ++++ G G QTWEYSP YAIRS+
 Sbjct: 55 APEGSTAFKCLLSARLCAALLSNISDCDETFNYWEPHTYLIYKGFGQTWEYSPVYAIRSY 114

Query: 332 SY-LVPLWIAGYPPLFLDIPSXXXXXXXXRLLLVIFSLVAEVKLYHSLKKNVSSKISFWYL 508
 +Y L+ W A + L R LL S V E+ Y ++ K +S L
 Sbjct: 115 AYLLLHAWPAAFHARILQTNKILVFYFLRCLLAFVSCVCELYFYKAVCKKFGHLHVSRRML 174

Query: 509 LFTTVAPGMSHSTIALLPSSFAM 577
 F ++ GM S+ A LPSSF M
 Sbjct: 175 AFLVLSTGMFCSSSAFLPSSFCM 197

H. sapiens

Score = 99.4 bits (246), Expect = 4e-20
 Identities = 56/143 (39%), Positives = 76/143 (53%), Gaps = 1/143 (0%)
 Frame = +2

Query: 152 SPTCSCMYWPILS*DLISTFYGIISDCDEVFNWYWEPLNFMLRGFGKQTWEYSPEYAIRSW 331
 +P S + +LS L + ISDCDE FNYWEP ++++ G G QTWEYSP YAIRS+
 Sbjct: 55 APEGSTAFKCLLSARLCAALLSNISDCDETFNYWEPHTYLIYGEFGQTWEYSPAYAIRSY 114

Query: 332 SY-LVPLWIAGYPPLFLDIPSXXXXXXXXRLLLVIFSLVAEVKLYHSLKKNVSSKISFWYL 508
 +Y L+ W A + L R LL S + E+ Y ++ K +S L
 Sbjct: 115 AYLLLHAWPAAFHARILQTNKILVFYFLRCLLAFVSCICELYFYKAVCKKFGHLHVSRRML 174

Query: 509 LFTTVAPGMSHSTIALLPSSFAM 577
 F ++ GM S+ A LPSSF M
 Sbjct: 175 AFLVLSTGMFCSSSAFLPSSFCM 197

FIGURE 13

S. cerevisiae ALG12

ATGCGTTGGTCTGTCCCTTGATACAGTGCTATTGACCGTGATTTCCCTTTCATCTAAT
CCAAGCTCCATTACCAAGGTGGAAGAGAGTTTAAATATTCAAGCCATTCATGAT
ATTTTAACCTACAGCGTATTTGATATCTCCAATATGACCACTTGAAATTTCTGG
AGTAGTCCCTAGAACATTCGTTGGTGCTGTGATTATTGCAATGCTTTCGAGACCTT
ATCTTTACTTGAGTTCTTTGATCCAACTTCCAGGCCTACGTCTATAGATGTTCAA
TTGGTCGTTAGGGGGATTGTTGGCCTCACCAATGGGCTTTCTTTTATCTATTTAAA
GAATTGTTTGCAAGATATGTTTGATGAAATCACTGAAAAGAAAAAGGAAGAAAA
TGAAGACAAGGATATATACATTTACGATAGCGCTGGTACATGGTTTTCTTTATTTT
TAATTGGCAGTTTCCACCTCATGTTCTACAGCACTAGGACTCTGCCTAATTTTGTC
ATGACTCTGCCTCTAACCAACGTCGCATTGGGGTGGGTTTTATTGGGTGCTTATAA
TGCAGCTATATTCCTATCTGCGCTCGTGGCAATTGTATTTAGACTGGAAGTGTGAG
CTCTCAGTGCTGGTATTGCTCTATTTAGCGTCATCTTCAAGAAGATTTCTTTATTC
GATGCTATCAAATTCGGTATCTTTGGCTTGGGACTTGGTTCGCCATCAGTATCAC
CGTTGATTCATATTTCTGGCAAGAATGGTGTCTACCTGAGGTAGATGGTTTCTTGT
TCAACGTGGTTGCGGGTTACGCTTCCAAGTGGGGTGTGGAGCCAGTTACTGCTTA
TTTCACGCATTACTTGAGAATGATGTTTATGCCACCAACTGTTTTACTATTGAATT
ACTTCGGCTATAAATTAGCACCTGCAAAATTAATAAATTGTCTCACTAGCATCTCTT
TTCCACATTATCGTCTTATCCTTTCAACCTCACAAGAATGGAGATTCATCATCTA
CGCTGTTCCATCTATCATGTTGTCTAGGTGACAGGAGCAGCACATCTATGGGAG
AATATGAAAGTAAAAAAGATTACCAATGTTTTATGTTTGGCTATATTGCCCTTATC
TATAATGACCTCCTTTTTCATTTCAATGGCGTTCTTGTATATATCAAGAATGAATT
ATCCAGGCGCGAGGCTTTAACTTCTTTTAATGACATGATTGTGGAATAAATAT
TACAAACGCTACAGTTCATATCAGCATACTCCTTGCATGACAGGTGTCACCTTAT
TTGGTGAATTGAACTACGGTGTGTACGGCATCAATTACGATAAGACTGAAAATAC
GACTTTACTGCAGGAAATGTGGCCCTCCTTTGATTTCTTGATCACCCACGAGCCA
ACCGCCTCTCAATTGCCATTCGAGAATAAGACTACCAACCATTGGGAGCTAGTTA
ACACAACAAAGATGTTTACTGGATTTGACCCAACCTACATTAAGAACTTTGTTTT
CCAAGAGAGAGTGAATGTTTTGTCTCTACTCAAACAGATCATTTTCGACAAGACC
CCTACCGTTTTTTTTGAAAGAATTGACGGCCAATTCGATTGTTAAAAGCGATGTCTT
CTTCACCTATAAGAGAATCAAACAAGATGAAAAAACTGATTGA

S. cerevisiae Alg12p

MRWSVLDTVLLTVISFHLIQAPFTKVEESFNIQAIHDILTYSVFDISQYDHLKFPGVVP
RTFVGAVIIAMLSRPYLYLSSLIQTSRPTSIDVQLVVRGIVGLTNGLSFIYLNCLQDM
FDEITEKKKEENEDKDIYTYDSAGTWFLFLIGSFHLMFYSTRTLPNFVMTLPLTNVAL
GWVLLGRYNAAIFLSALVAIVFRLEVSALSAGIALFSVIFKKISLFDIAKFGIFGLGLGS
AISITVDSYFWQEWCLPEVDGFLFNVAAGYASKWGVPEPVTAYFTHYLRMMFMPPTV
LLNLYFGYKLAPAKLKIVSLASLFHIIVLSFQPHKEWRFIYAVPSIMLLGATGAAHLW
ENMKVKKITNVLCAILPLSIMTSFFISMAFLYISRMNYPGGEALTSFNDMIVEKNITN
ATVHISIPPCMTGVTLFGELNYGVYGINYDKTENTLLQEMWPSFDLITHEPTASQLP
FENKTTNHVELVNTTKMFTGFDPTYIKNFVFQERVNVL SLLKQIIFDKTPTVFLKELT
ANSIVKSDVFFTYKRIKQDEKTD

FIGURE 14

P. pastoris ALG12

TCGGTCGAGAATGATAACTGAAGAACTCAAAATCTCTCACACTTTTCATCGT
TACTGTACTGGCAATCATTGCATTTTCAGCCTCATAAAGAATGGAGATTTAT
AGTTTACATTGTTCCACCACTTGTCATCACCATATCTACAGTACTTGCACA
ACTACCCAGGAGATTACAAATCGTCAAAGTTGCTGTTTTTCTCCTAAGTTT
CGGCTCTTTGCTCATATCCCTGTCGTTTCTTTTCATCTCATCGTATAACTAC
CCTGGGGGTGAAGCTTTACAGCATTGAACGAGAACTCCTTCTACTGGA
CCAAAGTTCCCTACCTGTTGATATTAAGGTTTCATATGGATGTCCCTGCATG
CATGACTGGGGTGACTTTATTTGGTTACTTGGATAACTCAAAATTGAACAA
TTTAAGAATTGTCTATGATAAAACAGAAGACGAGTCGCTGGACACAATCT
GGGATTCTTTCAATTATGTCATCTCCGAAATTGACTTGGATTCTTCGACTG
CTCCCAAATGGGAGGGGGATTGGCTGAAGATTGATGTTGTCCAAGGCTAC
AACGGCATCAATAAACAATCTATCAAAAATACAATTTTCAATTATGGAAT
ACTTAAACGGATGATAAGAGACGCAACCAAACCTTGATGTTGGATTATTC
GTACGGTCTTTCGATCCTTCATAAAATTTGATGATAAATTATTCATTTATG
AGAGGAGCAGTCAAACCTGAAAATATATACCTCATTTGTTCAATTTGGTGT
AAAGAGTGTGGCGGATAGACTTCTTGTAATCAGGAAAGCTACAATTCCA
ATTGCTGCAAAAAATACCAATGCCCATAA

P. pastoris Alg12p

RMITEELKISHTFIVTVLAIIAFQPHKEWRFIVYIVPPLVITISTVLAQLPRRFTIV
KVAVFLLSFGSLLISLSFLFISSYNYPGGEALQHLNEKLLLLDQSSLPVDIKVH
MDVPACMTGVTLFGYLDNSKLNLRIVYDKTEDESLDTIWDSFNYVISEIDLD
SSTAPKWEGDWLKIDVVQGYNGINKQSIKNTIFNYGILKRMIRDATKLDVGFI
RTVFRSFIKFDDKLFYERSSQ

FIGURE 15 (sheet 1)

P. pastoris ALG12 BLAST

		Score	E
Sequences producing significant alignments:		(bits)	Value
gi 1302525 emb CAA96310.1	ORF YNR030w [Saccharomyces cerev...]	102	5e-21
gi 19112221 ref NP_595429.1	putative involvement in cell w...	56	5e-07
gi 15864569 emb CAC83681.1	putative dolichyl-p-man: Man7G1...	53	4e-06
gi 13129114 ref NP_077010.1	dolichyl-p-mannose:Man7GlcNAc2...	53	4e-06
gi 22266724 gb AAM94900.1 AF311904.1	membrane protein SB87 ...	53	4e-06
gi 18478284 emb CAD22101.1	putative mannosyltransferase [M...	52	8e-06

Alignments

S. cerevisiae

Score = 102 bits (255), Expect = 5e-21
 Identities = 74/258 (28%), Positives = 121/258 (46%), Gaps = 19/258 (7%)

Query: 8 RMITEELKISHTFIVTVLAIIAFQPHKEWRFIVYIVPPLVITISTVLAQLPRRFTIVKVA 187
 ++ +LKI + + +++FQPHKEWRFI+Y VP +++ +T A L + K+
 Sbjct: 302 KLAPAKLKIVSLASLFIHIIIVLSFQPHKEWRFIIYAVPSIMLLGATGAHLWENMKVKKIT 361

Query: 188 -----VXXXXXXXXXXXXXXXXXXNYPGGEALQHLNEKLLLLDQSSLPVDIKVHMD 346
 + NYPGGEAL N+ ++ + VH+
 Sbjct: 362 NVLCLAILPLSIMTSFFISMAFLYISRMNYPGGEALTSFNDMIV---EKNITNATVHIS 417

Query: 347 VPACMTGVTLFGYLDNSKLNLRIVYDKTEDES-LDTIWDSFNVI-----SEIDLDS 505
 +P CMTGVTLFG L+ I YDKTE+ + L +W SF+++I S++ ++
 Sbjct: 418 IPPCMTGVTLFGELNYGVY---IN YDKTENTLLQEMWPSFDFLITHEPTASQLPFENK 474

Query: 506 TAPKWECDWLKIDVVQGYNGINKQSIKNTIFN-----YGILKRMIRDATKLDVGFIRTVF 670
 T WE ++ + + G + IKN +F +LK++I D K F++ +
 Sbjct: 475 TTNHWE---LVNTTKMFTGFDPTYIKNFVFQERVNVLSLLKQIIFD--KTPTVFLKELT 528

Query: 671 RSFIKFDKLFYIYERSSQ 724
 + I D F Y+R Q
 Sbjct: 529 ANSIVKSDVFFTYKRIKQ 546

S. pombe

Score = 56.2 bits (134), Expect = 5e-07
 Identities = 46/152 (30%), Positives = 62/152 (40%), Gaps = 11/152 (7%)

Query: 65 IIAFQPHKEWRFIVYIVPPLVITISTVLAQL-----PRRFTIVKVAVXXXXXXXXXX 220
 + +F HKEWRFI+Y + P S + A L + F I+++
 Sbjct: 295 VYSFLGHKEWRFIIYSI-PWFNAASAIGASLCFNASKFGKKIFEILRLMFFSGIIFGFIG 353

Query: 221 XXXXXXXXXXXNYPGGEALQHLNEKLLLLDQSSLPVDIKVHMDVPACMTGVTLFGYLDNSK 400
 Y YPGG AL L E + VHMDV CMTG+T F L +
 Sbjct: 354 SSFLLYVFQYAYPGGLALTRYE-----IENHPQVSVHMDVYPCMTGITRFSQLPS-- 404

FIGURE 15 (sheet 2)

Query: 401 LNNLRIVYDKTEDESL---DTIWDSFNYVISE 487
YDKTED + F+Y+I+E
Sbjct: 405 -----WYYDKTEDPKMLNSLSLFISQFDYLITE 431

Homo sapiens

Score = 53.1 bits (126), Expect = 4e-06
Identities = 41/149 (27%), Positives = 68/149 (45%), Gaps = 6/149 (4%)

Query: 59 LAIIAFQPHKEWRFIVYIVPPLVITISTVLAQLPRR-----FTIVKVAVXXXXXXXXXX 220
+A+ + PHKE RFI+Y P L IT + + L + + V
Sbjct: 299 MALYSLLPHKELRFIIYAFPMNLNITAARGCSYLLNNYKKSPLYKAGSLLVIGHLVVNAAY 358

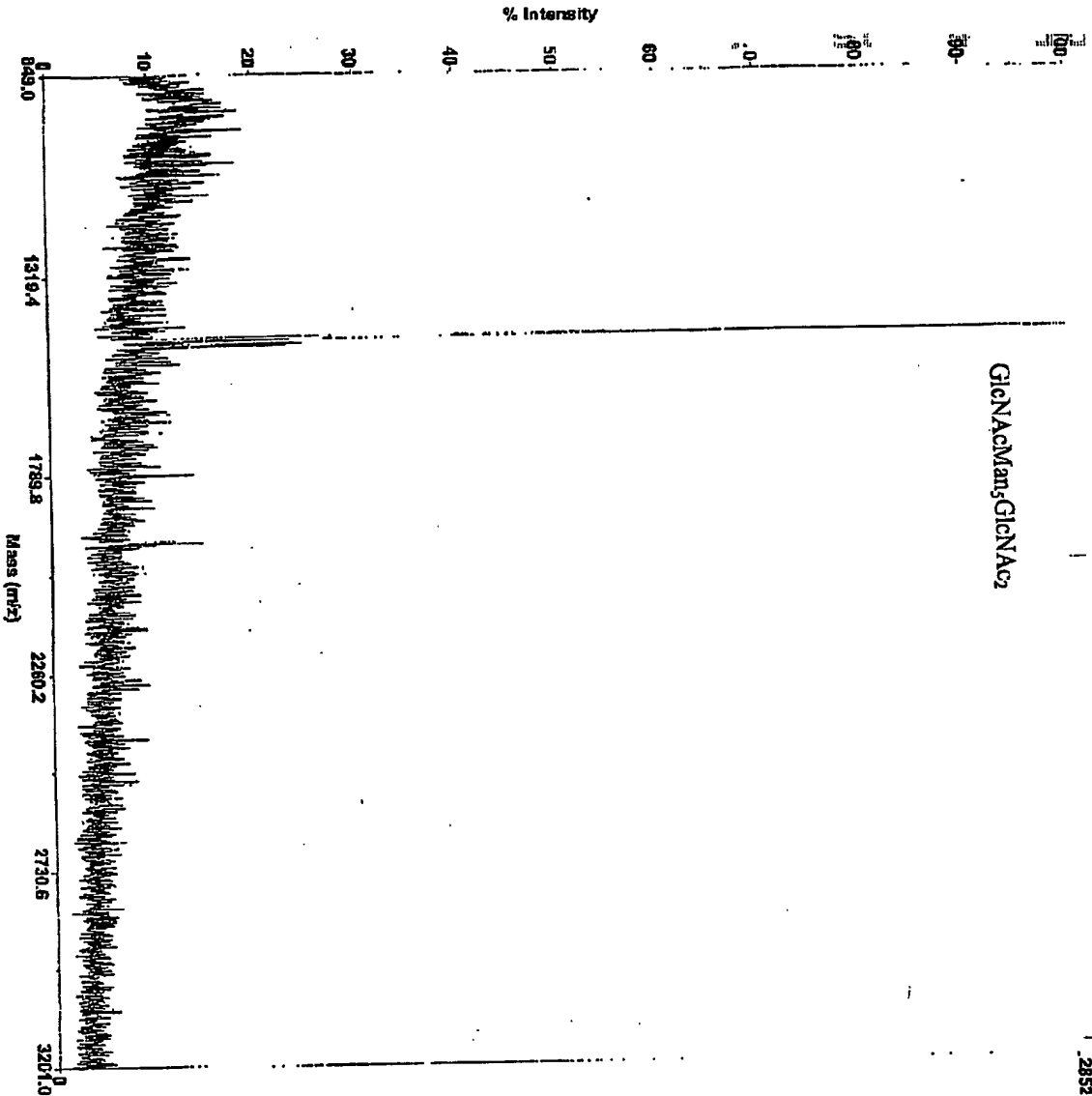
Query: 221 XXXXXXXXXXXYNYPGGEALQHLNEKLLLLDQSSLPVDIKVHMDVPACMTGVTLFGYLDNSK 400
+NYPGG A+Q L++ L+ Q+ D+ +H+DV A TGV+ F ++++
Sbjct: 359 SATALYVSHFNYPGGVAMQRLHQ--LVPPQT---DVLLHIDVAAAQTGVSRLQVNSAW 412

Query: 401 LNNLRIVYDKTEDESLDTIWDSFNYVISE 487
YDK ED T ++ +++ E
Sbjct: 413 R-----YDKREDVQPGTGMLAYTHILME 435

Applied Biosystems Voyager System 6235

Voyager Spec #1[BP = 1463.3, 2852]

FIGURE 16



Mode of operation: Linear
 Extraction mode: Delayed
 Polarity: Positive
 Acquisition control: Manual
 Accelerating voltage: 20000 V
 Grid voltage: 90%
 Guide wire on: 0.1%
 Extraction delay time: 125 nsec

Acquisition mass range: 850 -- 3200 Da
 Number of laser shots: 200/spectrum
 Laser intensity: 2786
 Laser Rep Rate: 20.0 Hz
 Calibration type: Default
 Calibration matrix: a-Cyano-4-hydroxycinnamic acid
 Low mass gate: 800 Da

Digitizer start time: 19.965
 Bin size: 0.5 nsec
 Number of data points: 37310
 Vertical scale: 500 mV
 Vertical offset: 0%
 Input bandwidth: 500 MHz

Sample well: 22
 Plate ID: 100 WELL PLATE
 Serial number: 6235
 Instrument name: Voyager-DE PRO
 Plate type filename: C:\VOYAGER\100 well plate.pl
 Lab name: PE Biosystems

Absolute x-position: 8212.98
 Absolute y-position: 35573.8
 Relative x-position: -454.524
 Relative y-position: -1213.7
 Shots in spectrum: 200
 Source pressure: 6.934e-007
 Mirror pressure: 8.616e-008
 TCS pressure: 0.01188
 TIS gate width: 10
 TIS flight length: 689

Printed: 10:47, December 18, 200

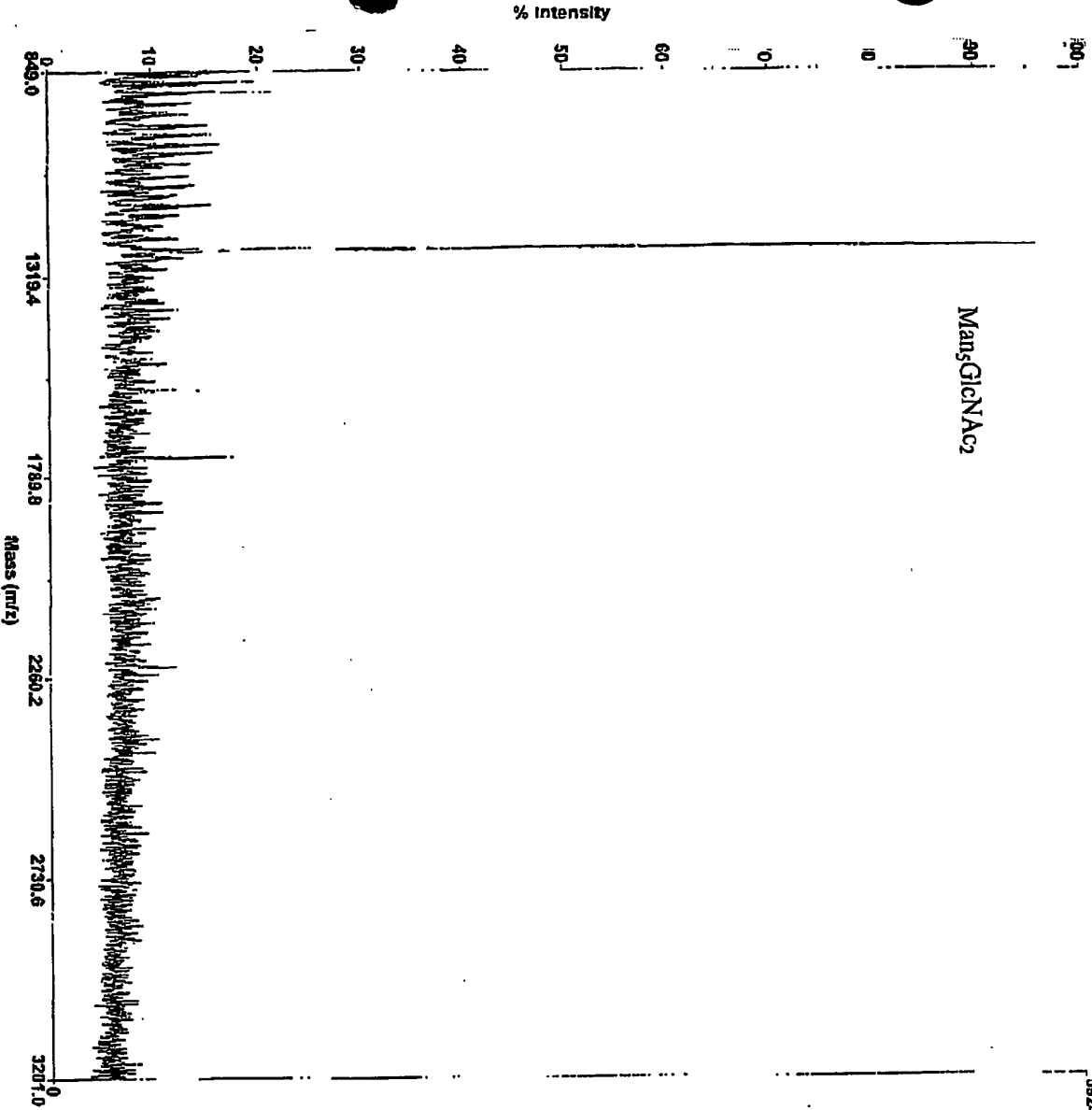
D:\Data\HuffmanData for burn\Other samples\NOV 2002\11-8-02\JN 10-31-02 IV B2 no enzymes 0001.dat

Applied Biosystems Voyager System 6235

Voyager Spec #1[BP = 1259.4, 5924]

PCT/US02/41510

WO 03/056914



Mode of operation: Unear
Extraction mode: Delayed
Polarity: Positive
Acquisition control: Manual

5924.0
Accelerating voltage: 20000 V
Grid voltage: 93%
Guide wire O: 0.1%
Extraction delay time: 125 nsec

Acquisition mass range: 850 - 3200 Da
Number of laser shots: 200/spectrum
Laser intensity: 2886
Laser Rep Rate: 20.0 Hz
Calibration type: Default
Calibration matrix: a-Cyano-4-hydroxydynamic a
Low mass gate: 800 Da

Digitizer start time: 19.855
Bin size: 0.5 nsec
Number of data points: 37310
Vertical scale: 500 mV
Vertical offset: 0%
Input bandwidth: 500 MHz

Sample well: 32
Plate ID: 100 WELL PLATE
Serial number: 6235
Instrument name: Voyager-DE PRO
Plate type filename: CAVOYAGER100 well plate.p
Lab name: PE Biosystems

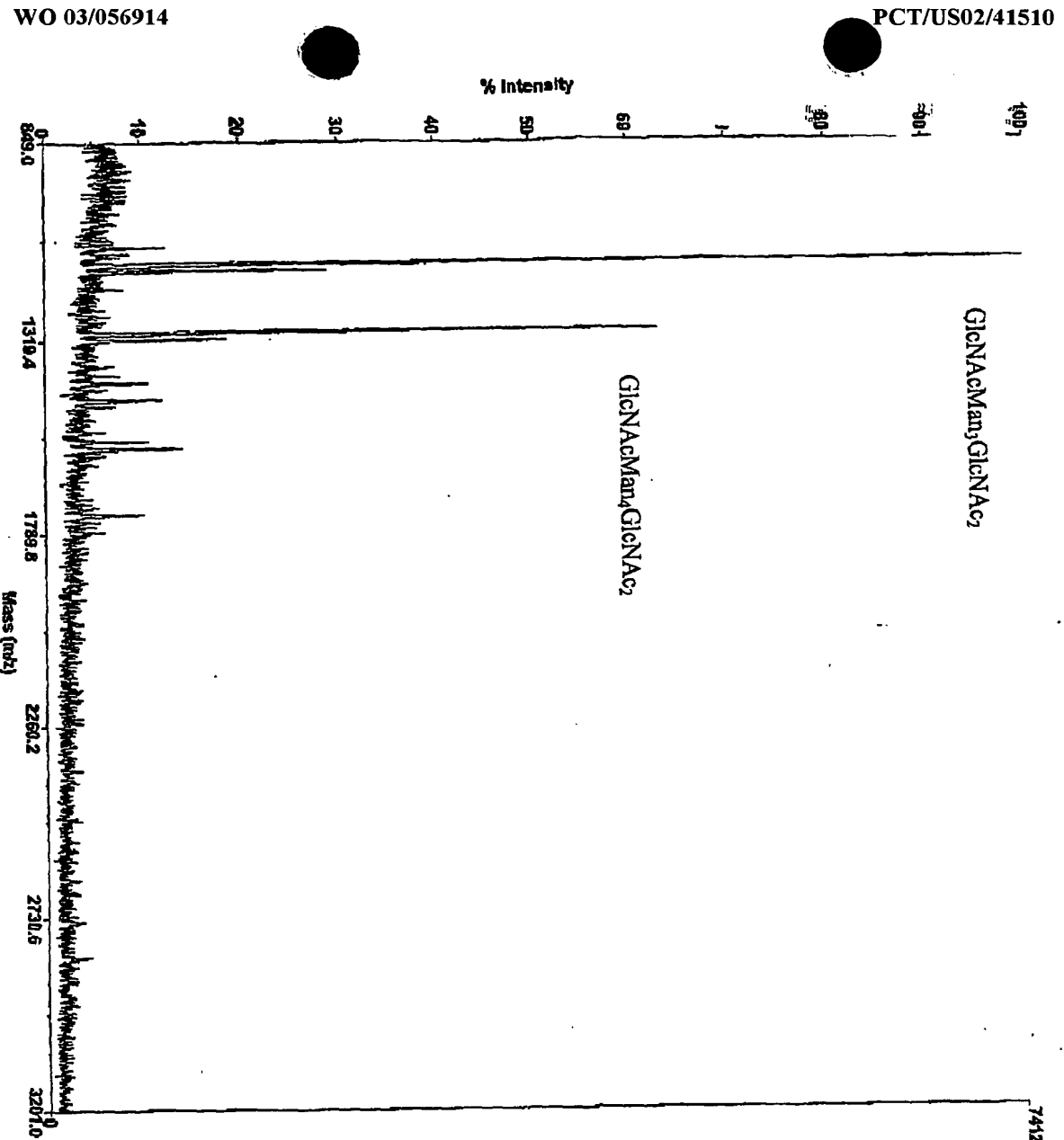
Absolute x-position: 6613.36
Absolute y-position: 30806.1
Relative x-position: -54.1439
Relative y-position: -1261.39
Shot in spectrum: 200
Source pressure: 7.709e-007
Mirror pressure: 9.387e-008
TC2 pressure: 0.01262
TIS gate width: 10
TIS flight length: 689

FIGURE 17

Applied Biosystems Voyager system v2.3.3

Voyager Spec #11BP = [1137.8, 7412]

FIGURE 18



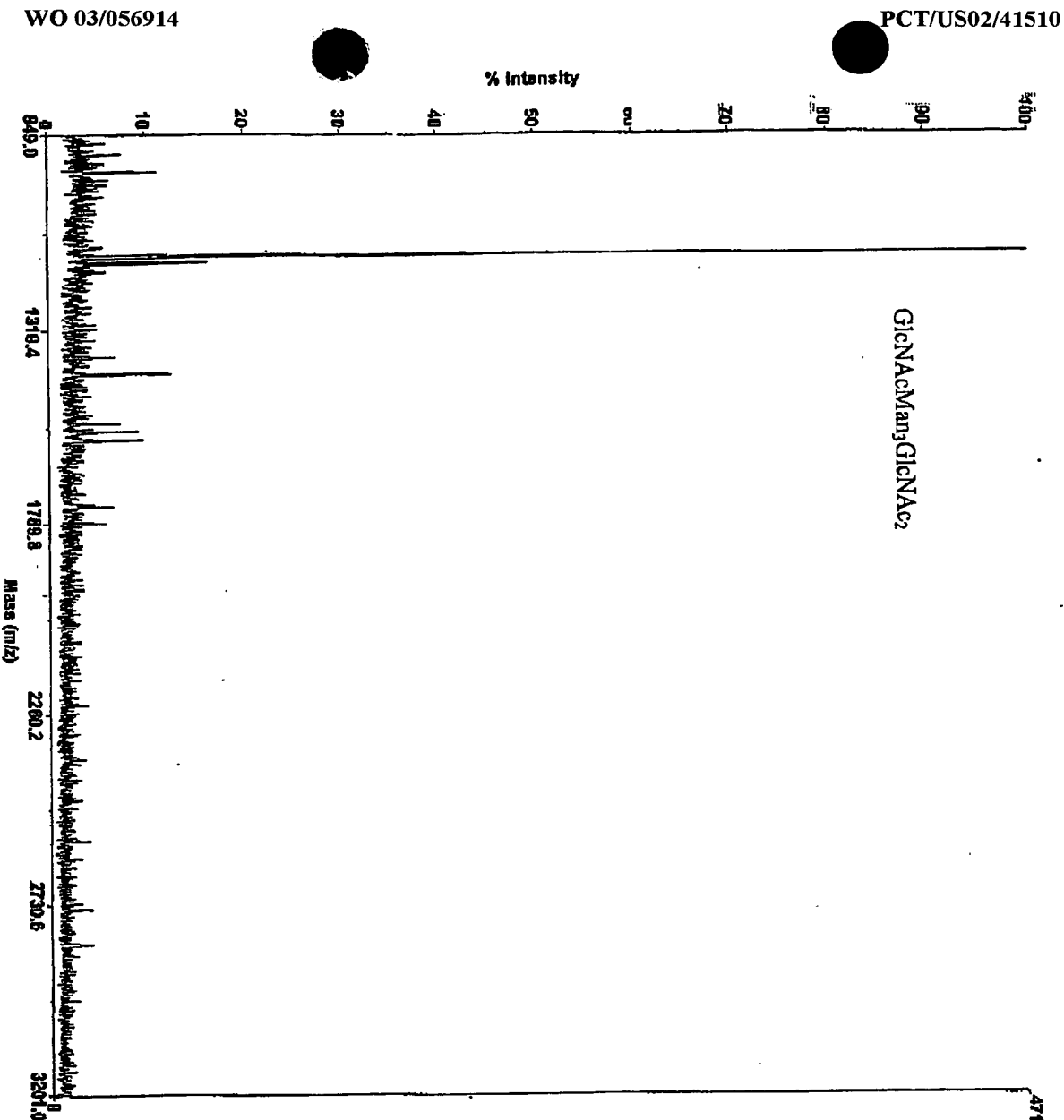
Acquired: 10:16:00, October 13, 2002
Data: Data\HuganData for Run\Other samples\OCT 2002\10-13-02\Rob-3.DMZ.dat

Mode of operation:	Linear
Extraction mode:	Delayed
Polarity:	Positive
Acquisition control:	Manual
Accelerating voltage:	20000 V
Grid voltage:	93%
Guide wire on:	0.1%
Extraction delay time:	725 nsec
Acquisition mass range:	850 - 3200 Da
Number of laser shots:	200/spectrum
Laser intensity:	2988
Laser Rep Rate:	20.0 Hz
Calibration type:	Default
Calibration method:	a-Cyano-4-hydroxycinnamic a
Low mass gate:	800 Da
Digitizer start time:	19.855
Bin size:	0.5 msec
Number of data points:	37310
Vertical scale:	500 mV
Vertical offset:	0%
Input bandwidth:	500 MHz
Sample well:	03
Plate ID:	100 WELL PLATE
Serial number:	8235
Instrument name:	Voyager-DE PRO
Plate type filename:	C:\VOYAGER\100 well plate.
Lab name:	PE Biosystems
Absolute x-position:	12810.8
Absolute y-position:	48689.9
Relative x-position:	1163.37
Relative y-position:	-637.62
Snobs in spectrum:	200
Source pressure:	1.781e-008
Mirror pressure:	1.525e-007
TOF pressure:	0.01459
TIS gate width:	10
TIS flight length:	889

Applied Biosystems Voyager System 6235

Voyager Spec #1[BP = 1139.5, 4716]

FIGURE 19



Acquired: 15:39:00, October 15, 2002
D:\Data\In\uanData for burn\Other samples\OCT 2002\10-16-02\RD 10-16-02 3 1,2dlgeet_000A.dat

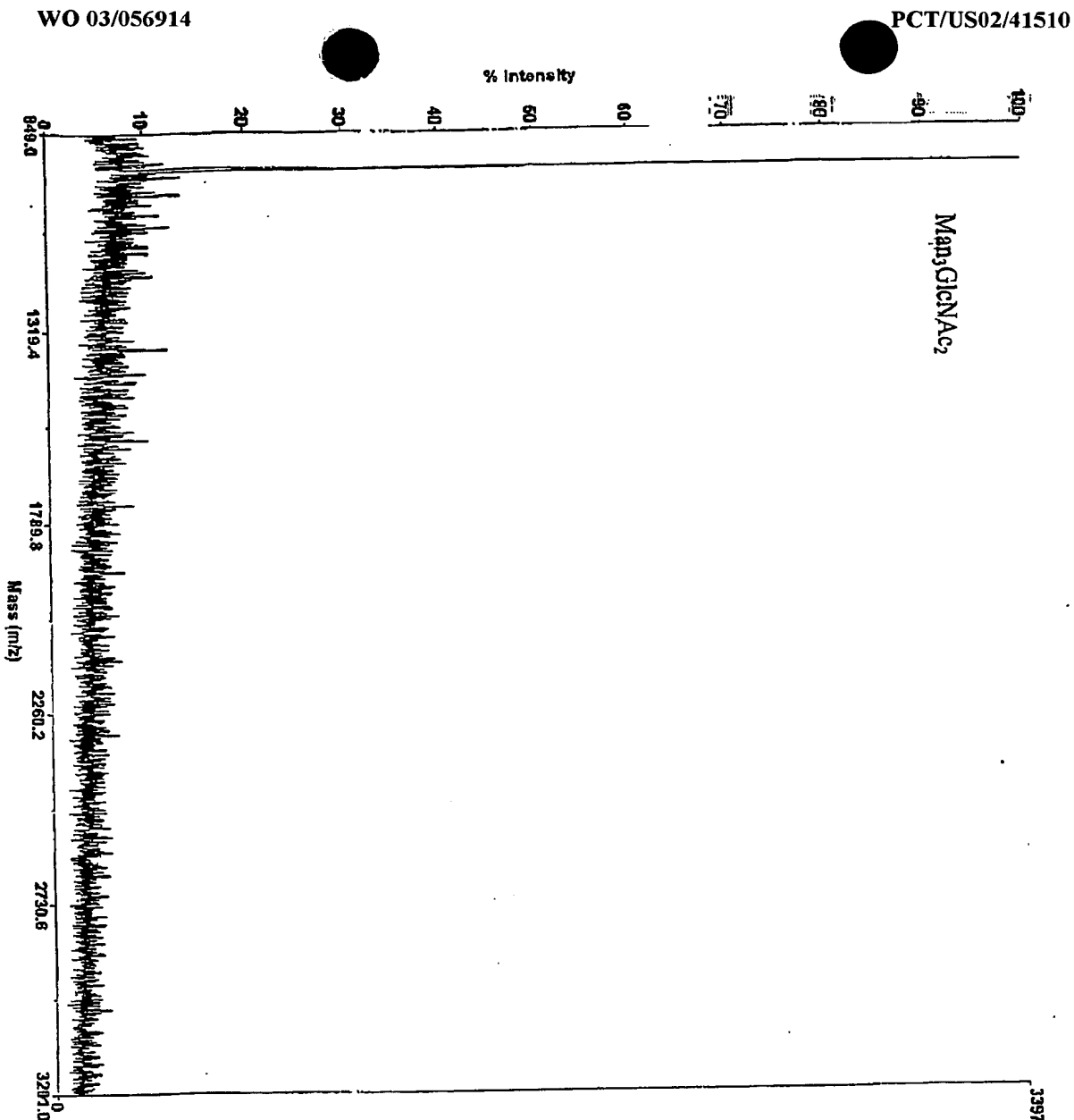
Mode of operation: Linear
Extraction mode: Delayed
Polarity: Positive
Acquisition control: Manual
Accelerating voltage: 20000 V
Grid voltage: 99%
Guide wire U: 0.1%
Extraction delay time: 125 nsec
Acquisition mass range: 850 - 3200 Da
Number of laser shots: 2000/spectrum
Laser intensity: 2766
Laser Rep Rate: 20.0 Hz
Calibration type: Dabell
Calibration matrix: a-Cyano-4-hydroxymethyl
Low mass gate: 800 Da
Digitizer start time: 19.995
Bin size: 0.5 nsec
Number of data points: 37310
Vertical scale: 500 mV
Vertical offset: 0%
Input bandwidth: 500 MHz
Sample well: 24
Plate ID: 100 WELL PLATE
Serial number: 6235
Instrument name: Voyager-DE PRO
Plate type filename: C:\VOYAGER\100 well plate.f
Lab name: PE Biosystems
Absolute x-position: 18177.2
Absolute y-position: 37530.3
Relative x-position: 1349.67
Relative y-position: 382.61
Shots in spectrum: 200
Source pressure: 1.390e-006
Mirror pressure: 1.285e-007
TCD pressure: 0.01248
TIS gate width: 10
TIS flight length: 689

Printed: 12:15, December 17, 2001

Applied Biosystems Voyager System 6235

Voyager Spec # [BP = 935.7, 3397]

FIGURE 20.



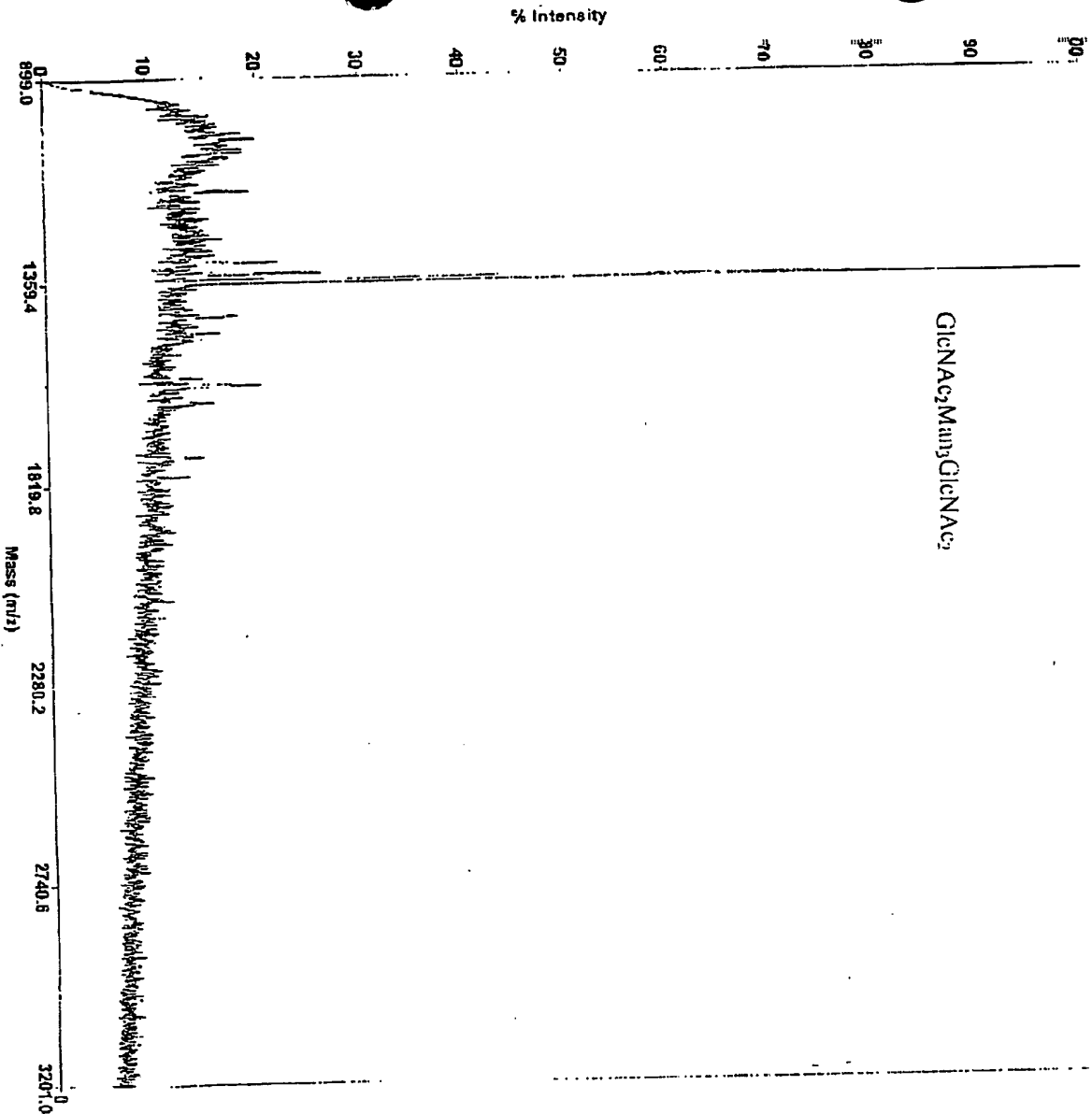
Mode of operation: Linear
 Extraction mode: Delayed
 Polarity: Positive
 Acquisition control: Manual
 Accelerating voltage: 20000 V
 Grid voltage: 93%
 Guide wire On: 0.1%
 Extraction delay time: 125 nsec
 Acquisition mass range: 650 - 3200 Da
 Number of laser shots: 200/spectrum
 Laser intensity: 2586
 Laser Rep Rate: 20.0 Hz
 Calibration type: Default
 Calibration matrix: a-Cyano-4-hydroxycinnamic a
 Low mass gate: 800 Da
 Digitizer start time: 19.955
 Bin size: 0.5 nsec
 Number of data points: 37310
 Vertical scale: 500 mV
 Vertical offset: 0%
 Input bandwidth: 500 MHz

Sample well: 22
 Plate ID: 100 WELL PLATE
 Serial number: 6235
 Instrument name: Voyager-DE PRO
 Plate type/Name: C:\VOYAGER\100 well plate.f
 Lab name: PE Biosystems

Absolute x-position: 5518.88
 Absolute y-position: 37692.8
 Relative x-position: -1148.82
 Relative y-position: 545.303
 Shots in spectrum: 200
 Source pressure: 6.934e-007
 Mirror pressure: 8.468e-008
 TCC pressure: 0.01269
 TIS gate width: 10
 TIS flight length: 689

Applied Biosystems Voyager System 6235

Voyager Spec #1 [BP = 1356.8, 5565]



Mode of operation: Linear
Extraction mode: Delayed
Polarity: Positive
Acquisition card: Automatic

FIGURE 21

5565.2 Acquisition card: Automatic

Accelerating voltage: 20000 V
Gain voltage: 93%
Guide wire ID: 0.1%
Extraction delay time: 125 nsec

Acquisition mass range: 900 - 3200 Da
Number of laser shots: 100/spectrum
Laser intensity: 2882
Laser Rep Rate: 20.0 Hz
Calibration type: Default
Calibration method: a-Cyano-4-hydroxynaphthoic acid
Low mass gate: 875 Da

Digitizer start time: 20.5305
Bin size: 0.5 nsec
Number of data points: 38159
Vertical scale: 500 mV
Vertical offset: 0%
Input bandwidth: 500 MHz

Laser control: Manual
Sample positioning: Automated
Random Pattern: Edge biased
Auto storage mode: Accumulate all passing

Min intensity: 250
Max intensity: 0
Resolution: 0
Signal-to-noise: 3

Sample well: 91
Plate ID: 100 WELL PLATE
Serial number: 6235
Instrument name: Voyager-DE PRO
Plate type filename: C:\VOYAGER\100 well plate.p
Lab name: PE Biosystems

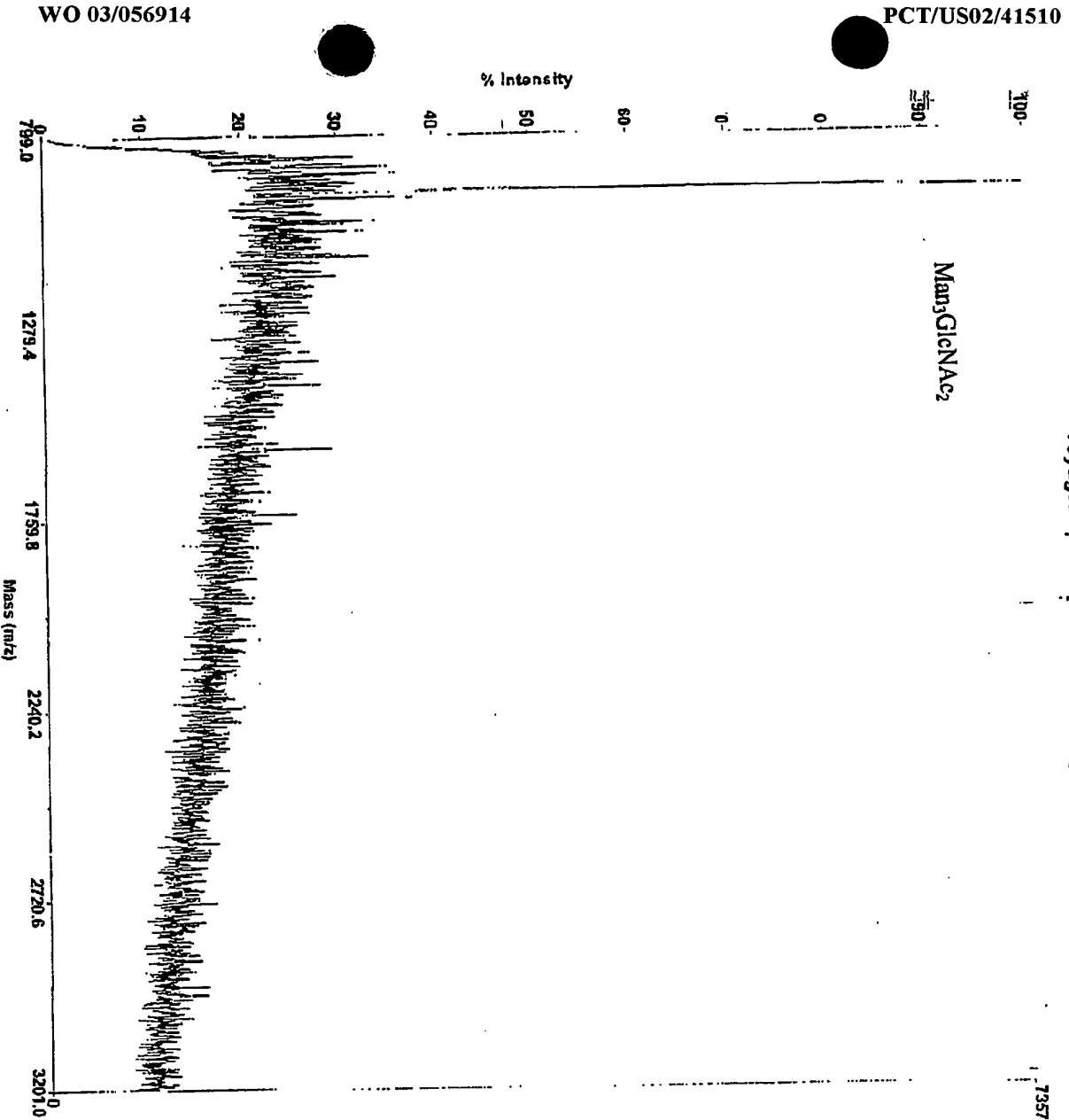
Absolute x-position: 1190.1
Absolute y-position: 2397.25
Relative x-position: -397.403
Relative y-position: 809.751
Shots in spectrum: 590
Source pressure: 9.938e-007
Mirror pressure: 9.246e-008
TIC2 pressure: 0.0122
TIS gate width: 10
TIS flight length: 689

Acquired: 12:57:00, October 28, 2002
Sample Description: Comments:
DataPath\Hq\uanData for burn\Screen data\October 2002\UN Media d\gca 10-23-02 BP8-JN LA 30-32 IV 10-24-02_91_0001.dat

Applied Biosystems Voyager System 6235

Voyager Spec #1 [BP = 934.5, 7357]

FIGURE 22



Mode of operation: Linear
Extraction mode: Delayed
Polarity: Positive
Acquisition control: Manual

Accelerating voltage: 19000 V
Grid voltage: 94%
Guide wire D: 0.15%
Extraction delay time: 175 nsec

Acquisition mass range: 800 -- 3200 Da
Number of laser shots: 150/spectrum
Laser intensity: 3052
Laser Rep Rate: 20.0 Hz
Calibration type: Default
Calibration matrix: a-Cyano-4-hydroxycinnamic A
Low mass gate: 800 Da

Digitizer scan time: 19.8945
Bin size: 0.5 nsec
Number of data points: 38535
Vertical scale: 500 mV
Vertical offset: 0%
Input bandwidth: 500 MHz

Sample well: 91
Plate ID: 100 WELL PLATE
Serial number: 6235
Instrument name: Voyager-DE PRO
Plate type filename: C:\VOYAGER\100 well plate.p
Lab name: PE Biosystems
Absolute x-position: 1843.77
Absolute y-position: 305.377
Relative x-position: 256.268
Relative y-position: -1281.12
Shots in spectrum: 150
Source pressure: 6.071e-007
Mirror pressure: 8.165e-008
TIC2 pressure: 0.01198
TIS gate width: 10
TIS flight length: 689

WO 03/056914

PCT/US02/41510

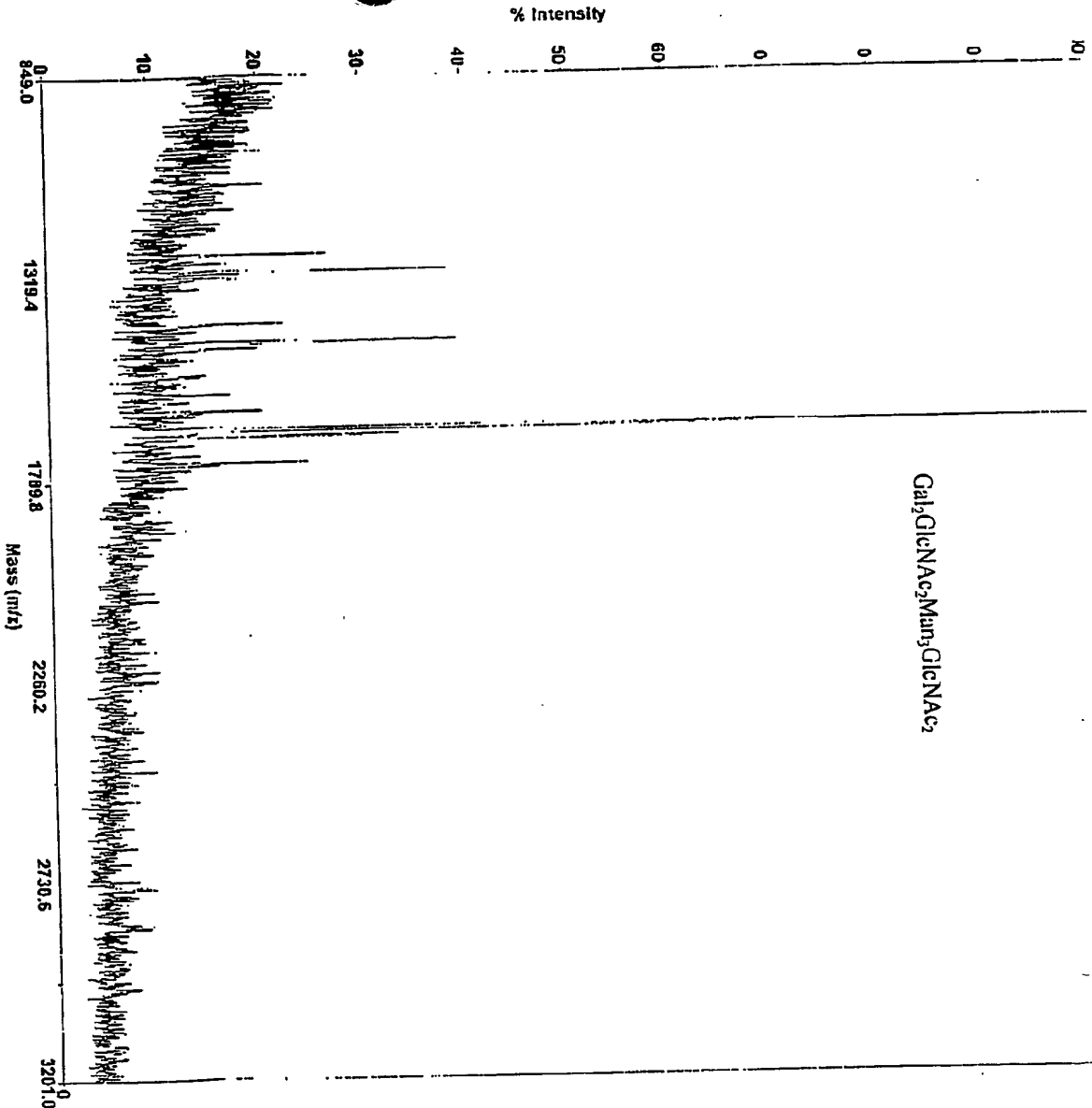
AcqRef: 08:30:00, October 30, 2002
D:\Data\HijuanData for BurnScreen data\October 2002\B-JN LA 30-32 10-24-02 1V\B-JN LA 30-32 10-24-02_91Hexo digest_0002.dai

Printed: 10:59, December 18, 200

Applied Biosystems Voyager System 6235

Voyager Spec #1[BP = 1666.9, 3309]

FIGURE 23.



Mode of operation: Linear
 Extraction mode: Delayed
 Polarity: Positive
 Acquisition control: Manual

Accelerating voltage: 20000 V
 Grid voltage: 93%
 Guide wire D: 0.1%
 Extraction delay time: 125 µsec

Acquisition mass range: 850 - 3200 Da
 Number of laser shots: 2000/spectrum
 Laser intensity: 2986
 Laser Rep Rate: 20.0 Hz
 Calibration type: Default
 Calibration matrix: α-Cyano-4-hydroxymethyl
 Low mass gate: 800 Da

Digitizer start time: 19.955
 Bin size: 0.5 µsec
 Number of data points: 37310
 Vertical scale: 500 mV
 Vertical offset: 0%
 Input bandwidth: 500 MHz

Sample well: 77
 Plate ID: 100 WELL PLATE
 Serial number: 6235
 Instrument name: Voyager-DE PRO
 Plate type filename: C:\VOYAGER\100 well plate.p
 Lab name: PE Biosystems

Absolute x-position: 33013.7
 Absolute y-position: 12200.9
 Relative x-position: 945.594
 Relative y-position: 513.403
 Shots in spectrum: 200
 Source pressure: 5.860e-007
 Mirror pressure: 7.263e-008
 TCO pressure: 0.01159
 TIS gate width: 10
 TIS flight length: 689

Acquired: 13:05:00, December 07, 2002

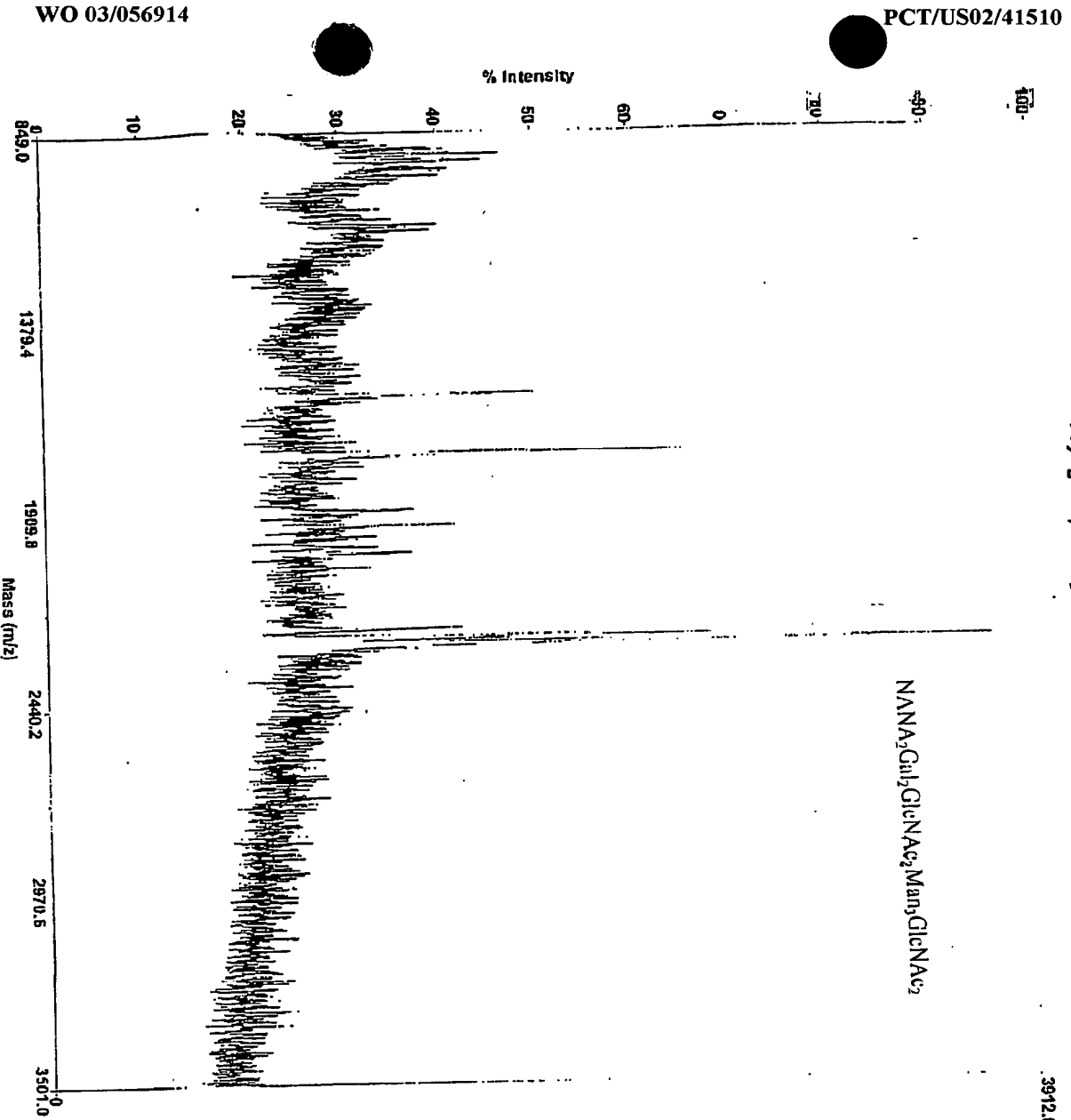
Filename: \\falcon\workspace\sam\test12-7-02\Bob.PBP6-5+Gal_T_0001.dat

Printed: 11:05, December 18, 2002

Applied Biosystems Voyager System 6235

Voyager Spec #1[BP = 2264.5, 3912]

FIGURE 24



Mode of operation: Linear
Extraction mode: Delayed
Polarity: Negative
Acquisition control: Manual
3912.0 Acquisition control:

Accelerating voltage: 20000 V
Grid voltage: 93%
Guide wire to: 0.1%
Extraction delay time: 175 nsuc

Acquisition mass range: 850 -- 3500 Da
Number of laser shots: 200/spectrum

Laser intensity: 3365
Laser Rep Rate: 200 Hz
Calibration type: Default
Calibration matrix: a-Cyano-4-hydroxycinnamic ac
Low mass gate: 800 Da

Digitizer start time: 19.958
Bin size: 0.5 usuc
Number of data points: 40049
Vertical scale: 500 mV
Vertical offset: 0%
Input bandwidth: 500 MHz

Sample well: 42
Plate ID: 100 WELL PLATE
Serial number: 6235
Instrument name: Voyager-DE PRO
Plate type filename: C:\VOYAGER\100 well plate.pl
Lab name: PE Biosystems

Absolute x-position: 7823.04
Absolute y-position: 28538.8
Relative x-position: 1155.54
Relative y-position: -448.927
Shots in spectrum: 200
Source pressure: 7.54e-007
Mirror pressure: 8.456e-008
TC2 pressure: 0.01302
TIS gate width: 10
TIS flight length: 689

FIGURE 25

S. cerevisiae ALG6

ATGGCCATTGGCAAAGGTTACTGGTGAACAAACCAGCAGAAGAATCATT
TTATGCTTCTCCAATGTATGATTTTTTGTATCCGTTTAGGCCAGTGGGGAA
CCAATGGCTGCCAGAATATATTATCTTTGTATGTGCTGTAATACTGAGGTG
CACAATTGGACTTGGTCCATATTCTGGGAAAGGCAGTCCACCGCTGTACG
GCGATTTTGAGGCTCAGAGACATTGGATGGAAATTACGCAACATTTACCG
CTTTCTAAGTGGTACTGGTATGATTTGCAATACTGGGGATTGGACTATCCA
CCATTAACAGCATTTCATTTCGTACCTTCTGGGCCTAATTGGATCTTTTTTCA
ATCCATCTTGGTTTGCCTAGAAAAGTCACGTGGCTTTGAATCCCCCGATA
ATGGCCTGAAAACATATATGCGTTCTACTGTCATCATTAGCGACATATTGT
TTTACTTTTCTGCAGTAATATACTTTACTAAGTGGCTTGGTAGATATCGAA
ACCAGTCGCCCATAGGACAATCTATTGCGGCATCAGCGATTTTGTTC AAC
CTTCATTAATGCTCATTGACCATGGGCACTTTCAATATAATTCAGTCATGC
TTGGCCTTACTGCTTATGCCATAAATACTTATTAGATGAGTATTATGCTA
TGGCGGCCGTTTGT TTTGTCTATCCATTTGTTTTAAACAAATGGCATTGTA
TTATGCACCGATTTTTTTTTGCTTATCTATTAAGTCGATCATTGCTGTTCCCC
AAATTTAACATAGCTAGATTGACGGTTATTGCGTTTGCAACACTCGCAACT
TTTGCTATAATATTTGCGCCATTATATTTCTTGGGAGGAGGATTAAAGAAT
ATTCACCAATGTATTCACAGGATATTCCTTTTGCCAGGGGGCATCTTCGAA
GACAAGGTTGCTAACTTCTGGTGCGTTACGAACGTGTTTGTA AAAATACAA
GGAAAGATTCACTATACAACA ACTCCAGCTATATTCATTGATTGCCACCGT
GATTGGTTTCTTACCAGCCATGATAATGACATTACTTCATCCCAAAAAGCA
TCTTCTCCCATACGTGTTAATCGCATGTTTCGATGTCCTTTTTTCTTTTAGC
TTTCAAGTACATGAGAAA ACTATCCTCATCCCACTTTTGCCTATTACACTA
CTCTACTCCTCTACTGATTGGAATGTTCTATCTCTTGTAAGTTGGATAAAC
AATGTGGCTTTGTTTACGCTATGGCCTTTGTTGAAAAAGGACGGTCTTCAT
TTACAGTATGCCGTATCTTTCTTACTAAGCAATTGGCTGATTGGAAATTC
AGTTTTATTACACCAAGGTTCTTGCCAAAATCTTTAACTCCTGGCCCTTCT
ATCAGCAGCATCAATAGCGACTATAGAAGAAGAAGCTTACTGCCATATAA
TGTGGTTTGGAAAAGTTTATCATAGGAACGTATATTGCTATGGGCTTTTA
TCATTTCTTAGATCAATTTGTAGCACCTCCATCGAAATATCCAGACTTGTG
GGTGTTGTTGAACTGTGCTGTTGGGTTCAATTTGCTTTAGCATATTTTGGCTA
TGGTCTTATTACAAGATATTCATTCCGGTAGCAAATCCATGAAGGACTTG
TAG

S. cerevisiae ALG6p

MAIGKRLLVNKP AEESFYASPMYDFLYPFRPVGNQWLPEYIIFVCAVILRCTIG
LGPYSGKGSPLYGD FEAQRHWMEITQHLPLSKWYWYDLQYWGLDYPPLTA
FHSYLLGLIGSFFNPSWFALEKSRGFESPDNGLKTYMRSTVIISDILFYFPAVITY
FTKWLGRYRNQSPIGQSIAASAILFQPSLMLIDHGHFQYNSV MLGLTAYAINN
LLDEYYAMAAVCFVLSICFKQMALYYAPIFFAYLLSRSLFPKFNIARLTVI AF
ATLATFAIIFAPLYFLGGGLKNIHQCIHRIFPFARGIFEDKVANFWCVTNV FVK
YKERFTIQQLQLYSLIATVIGFLPAMIMTLLHPKKHLLPYVLIACSMSFFLFSFQ
VHEKTILIPLLPITLLYSSTDWNVLSLVSWINNVALFTLWPLLKKDGLHLQYA
VSFLLSNWLIGNFSFITPRFLPKSLTPGPSISSINSDYRRRSLLPYNV VWKSFIIGT
YIAMGFYHFLDQFVAPPSKYPDLWVLLNCAVGFICFSIFWLWSYYKIFTS GSK
SMKDL

FIGURE 26

P. pastoris ALG6

ATGCCACATAAAAGAACGCCCTCTAGCAGTCTGCTGTATGCAAGAATTCC
AGGGATCTCTTTTGAAAACCTCTCCGGTGTTTGATTTTTTGTCTCCTTTTGGA
CCCGCTCCTAATCAATGGGTAGCACGATACATCATCATCTTTGCAATT
CTCATCAGATTGGCAGTTGGGCTGGGCTCCTATTCCGGCTTCAACACCCCT
CCAATGTATGGGGATTTTGAAGCTCAGAGGCATTGGATGGAAATTACTCA
GCATTTATCCATAGAAAAATGGTACTTCTACGACTTGCAATATTGGGGGCT
TGACTATCCTCCCTTGACAGCCTTTCATTCATACTTCTTTGGCAAATTAGGC
AGCTTCATCAATCCAGCATGGTTTGCTTTAGACGTCTCCAGAGGGTTTGAA
TCAGTGGATCTAAAATCGTACATGAGGGCGACCGCAATTCTCAGTGAGCT
GTTATGTTTTATTCCAGCTGTCATTTGGTATTGTCGTTGGATGGGACTTAAC
TACTTCAATCAAAACGCCATTGAGCAAACATAATAGCGTCTGCTATTCTT
TTCAATCCATCTTTAATTATCATAGATCATGGCCACTTCCAGTACAACCTCA
GTTATGCTAGGTTTTGCTTTATTATCCATATTAAATCTGTTGTACGATAATT
TTGCATTAGCGGCTATTTTTTTTCGTTCTTTCAATAAGCTTTAAGCAAATGGC
TCTCTATTATAGCCCCATCATGTTTTTTTACATGCTGAGTGTGAGTTGTTGG
CCTTTGAAAAACTTCAACTTGTTGAGATTGGCTACTATCAGTATTGCAGTA
CTCTTGACTTTTGCAACTCTATTACTGCCTTTTGTATTAGTAGATGGGATGT
CACAAATTGGCCAAATATTATTCAGAGTTTTCCCGTTTTCAAGAGGCTTGT
TTGAGGATAAGGTGGCCAACTTTTGGTGTACAACGAATATACTGGTAAAG
TACAAACAGTTATTCAGTACAAAACCCTTACTAGGATATCGCTAGTAGC
AACTTTGATTGCAATTAGTCCGTCTTGCTTCATCATTTTTACTCACCCAAAG
AAGGTTTTACTACCGTGGGCTTTTGCTGCTTGCTCTTGGGCGTTCTATCTTT
TCTCTTTCCAAGTCCACGAGAAATCAGTTTTTAGTTCATTGATGCCTACCA
CTCTATTACTGGTAGAAAAAGACTTGGACATCATCTCAATGGTCTGCTGGA
TTTCTAATATTGCCTTCTTCAGCATGTGGCCTCTATTAAAAAGAGACGGGC
TGGCTTTGGAATATTTTGTCTTGGGAATATTGAGTAATTGGCTGATTGGAA
ACCTCAATTGGATTAGTAAATGGCTTGTCCTCCAGTTTCCTGATTCCAGGGC
CTACTCTCTCAAAAAAGTTCCTAAAAGAGATACTAAAACAGTTGTTTCA
ACTACTGGTTTTTGGGGGTCAGTAACATTTCGTTTCATACCTCGGAGCTACA
GTTATCCAGTTCGTAGATTGGCTGTACCTTCCACCTGCCAAGTATCCAGAT
TTGTGGGTTATTTTGAACACTACATTGTCGTTTGCTTGTTTCGGGTGTTTT
GGCTATGGATTAACATACTGTACATTTTGCGTGATTTTAAGCTTAAAG
ATGCTTAG

P. pastoris Alg6

MPHKRTPSSSLYARIPGISFENSPVFDLSPFGPAPNQWVARYIIIFAILRLAV
GLGSYSGFNTPPMYGDFEAQRHWMETQHLSIEKWYFYDLQYWGLDYPPLT
AFHSYFFGKLGSFINPAWFALDVSRGFESVDLKSVMRATAILSELLCFIPAVIW
YCRWMGLNYFNQNAIEQTIIASAILFNPSLIIDHGHFQYNSVMLGFALLSILNL
LYDNFALAAIFFVLISISFKQMALYYSPIMFFYMLSVCWPLKNFNLLRLATISI
AVLLTFATLLLPFVLVDGMSQIGQILFRVFPFSRGLFEDKVANFWCTTNILVK
YKQLFTDKTLTRISLVATLIAISPSCFIIFTHPKKVLLPWAFACSWAFYLFQ
VHEKSVLVPLMPTTLLLVEKDLDIISMVCWISNIAFFSMWPLLKRDGLALEYF
VLGILSNWLIGNLNWISKWLVPFLIPGPTLSKKVPKRDTKTVVHTHWFWGS
VTFVSYLGATVIQFVDWLYLPPAKYPDLWVILNTTSLFACGLFWLWINYNL
YILRDFKLKDA

FIGURE 27 (sheet 1)

P. pastoris ALG6 BLAST

Score E
Sequences producing significant alignments: (bits) Value

gi 1420090 emb CAA99190.1	ORF YOR002w [Saccharomyces cerev...]	489	e-137
gi 7490584 pir T40396	glucosyltransferase - fission yeast ...	369	e-101
gi 19921070 ref NP_609393.1	CG5091-PA [Drosophila melanoga...]	47	4e-64
gi 15240920 ref NP_198662.1	glucosyltransferase-like prote...	244	3e-63
gi 7019325 ref NP_037471.1	dolichyl-P-Glc:Man9GlcNAc2-PP-d...	238	2e-61
gi 12002040 gb AAG43163.1	AF063604_1 brain my046 protein [H...]	236	7e-61
gi 1176671 sp Q09226	ALG6 CAEEL Probable dolichyl pyrophosph...	222	9e-57
gi 21302638 gb EAA14783.1	agCP4617 [Anopheles gambiae str....]	219	8e-56
gi 5441788 emb CAB46771.1	probable glucosyltransferase [Sc...]	192	1e-47
gi 13129070 ref NP_076984.1	hypothetical protein MGC2840 s...	112	1e-23
gi 2996578 emb CAA12176.1	glucosyltransferase [Homo sapiens]	112	1e-23
gi 20835439 ref XP_131506.1	similar to Dolichyl pyrophosph...	104	3e-21

Alignments

S. cerevisiae

Score = 489 bits (1259), Expect = e-137
Identities = 274/530 (51%), Positives = 358/530 (67%), Gaps = 5/530 (0%)

Query: 20 SFENSPVDFLSPFGPAPNQWVXXXXXXXXXXXXXXXXXVGLGSYSGFNTPPMYGDFEQRH 79
SF SP++DFL PF P NQW+ +GLG YSG ++PYGDFEQRH
Sbjct: 16 SFYASPMYDFLYPFRPVGNQWLPEYIIFVCAVILRCTIGLGPYSGKGSPLYGDFEQRH 75

Query: 80 WMEITQHLSEIKWYFYDLQYWGLDYPPLTAFHSYFFGKLGSFINPAWFALDVSRGFESVD 139
WMEITQHL + KQY+YDLQYWGLDYPPLTAFHSY G +GSF NP+WFAL+ SRGFES D
Sbjct: 76 WMEITQHLLPSKQWYDYDLQYWGLDYPPLTAFHSYLLGLIGSFFNPSWFALEKSRGFESPD 135

Query: 140 --LKSVMRATAILSELLCFIPAVIWCYRWMGLNYFNQNAIEQTIIASAILFNPSLIIDH 197
LK+YMR+T I+S++L + PAVI++ +W+G Y NQ+ I Q+I ASAILF PSL++IDH
Sbjct: 136 NGLKTYMRSTVIISDILFYFPAVIYFTKWLQ-RYRNQSPIGQSIASAILFQPSLMLIDH 194

Query: 198 GHFQYNSVMLGFALLSILNLLYDNFALAAIFFVLSISFKQMALYYSPIMFFYMLSVSCWP 257
GHFQYNSVMLG +I NLL + +A+AA+ FVLSI FKQMALYY+PI F Y+LS S
Sbjct: 195 GHFQYNSVMLGLTAYAINNLLDEYYAMAACFVLSICFKQMALYYAPIFFAYLLSRSL- 253

Query: 258 LKNFNLLRLATISIAVLLTFATLLLP-FVLVDGMSQIGQILFRVFPFSRGLFEDKVANFW 316
FN+ RL I+ A L TFA + P + L G+ I Q + R+PPF+RG+FEDKVANFW
Sbjct: 254 FPKFNARLTVIAFATLATFAIIFAPLYFLGGGLKNIHQCIHRIFPFARGIFEDKVANFW 313

Query: 317 CTTNVLVKYQLFTDKTLTRISLVATLIAISPCFIIFTHPKKVLLPWAFACSWAFYLF 376
C TN+ VKYK+ FT + L SL+AT+I P+ + HPKK LLP+ ACS +F+LF
Sbjct: 314 CVTNVFKYKERFTIQQLQLYSLIATVIGFLPAMIMTLLHPKKHLLPYVLIACMSFFLF 373

Query: 377 SFQVHEKSXXXXXXXXXXXXXKOLDIISMVCWISNIAFFSMWPLLKRDGLALEYFVLGI 436
SFQVHEK+ D +++S+V WI+N+A F++WPLLK+DGL L+Y V +
Sbjct: 374 SFQVHEKTILIPLLPITLLYSSTDWNVLSLVSWINNVALFTLWPLLKDGHLQYAVSFL 433

Query: 437 LSNWLIGNLNLWISKWLVPFLIPGPTLSKKVPKRDTKTVVHTHWFGSVTFVSYLGVATVI 496
LSNWLIGN ++I+ +P L PGP++S +++++ + W S +Y+
Sbjct: 434 LSNWLIGNFSFITPRFLPKSLTPGPSISSINSYRRRSLLPYNVWVKSFIIGTYIAMGFY 493

FIGURE 27 (sheet 2)

Query: 497 QFVDWLYLPPAKYPDLWVILNTTSLFACGLFWLWINYNLYILRDFKLKD 546
 F+D PP+KYPDLWV+LN + F CF +FWLW Y ++ +KD
 Sbjct: 494 HFILDQFVAPPSKYPDWVLLNCAVGFCFSIFWLWSYKIFTSGSKSMKD 543

S. pombe

Score = 369 bits (946), Expect = e-101
 Identities = 228/513 (44%), Positives = 315/513 (61%), Gaps = 35/513 (6%)

Query: 21 FEN-SPVDFLSPFGPAPNQWVXXXXXXXXXXXXXXXXXVGLGSYSGFNTPPMYGDFAQRH 79
 FEN +PV F+S F ++++ + +G YSG+NTPPMYGDFAQRH
 Sbjct: 5 FENGAPVQQFVSRFRSYSSKFLFFPCLIMSLVFMQWLISIGPYSGYNTPPMYGDFAQRH 64

Query: 80 WMEITQHLSEIKWYFYDLQYWGLDYPPLTAFHSYFFGKLGS-FINPAWFALDVSRGFESV 138
 WME+T H + +WYF DLQ+WGLDYPPLTA+ S+FFG +G F NP WFA SRGFES+
 Sbjct: 65 WMEITLHTPVSQWYFRDLQWGLDYPPLTAYVSWFFGIIGHYFFNPEWFADVTSRGFESL 124

Query: 139 DLKSYMRTAILSELLCFIPAVIWCYCRMGLNYFNQNAIEQTIIASAILFNPSLIIIDHG 198
 +LK +MR+T I S LL +P +++Y +W N +++ +LF P+L++IDHG
 Sbjct: 125 ELKLFMRSTVIASHLLILVPPLMFYSKWSRRI--PNFVDRNASLIMVLFQPALLLIDHG 182

Query: 199 HFQYNSVMLGFALLSILNLLYDNFALAAIFFVLSISFKQMALYYSPIMFFYMLSVSCWPL 258
 HFQYN VMLG + +I NLL + + A FF L+++FKQMALY++P +FFY+L P
 Sbjct: 183 HFQYNCVMLGLVMAIANLLKNQYVAATFFFCALATFKQMALYFAPPIFFYLLGTCVKPK 242

Query: 259 KNFNLLRLATISIAVLLTFATLLLPFVLVDGMSQIGQILFRVFPFSRGLFEDKVFANFWCT 318
 F+ R +S+ V+ TF+ +L P++ +D + + QIL RVFFF+RGL+EDKVFANFWCT
 Sbjct: 243 IRFS--RFILLSVTTVFTFSLILFPWIYMDYKTLPLQILHRVFPFARGLWEDKVFANFWCT 300

Query: 319 TNILVKYKQLFTDKTLTRISLVATLIAISPSCFIIFTHPKKVLLPWAFACSWAFYLFSS 378
 N + K +++FT L ISL+ TLI+I PSC I+F +P+K LL FA+ SW F+LFSS
 Sbjct: 301 LNTVFKIREVFTLHQLQVISLIFTLISILPSCVILFLYPRKRLALGFASASWGFFLFSS 360

Query: 379 QVHEKSXXXXXXXXXXXXXKDLDIISMVCWISNIAFFSMWPLLKRDGLALEYFVLGILS 438
 QVHEKS ++ + +N+A FS+WPLLK+DGL L+YF L ++
 Sbjct: 361 QVHEKSVLLPLPTSILLCHGNITTKPWIALANNLAVFSLWPLLKKDGLGLQYFTLVLMW 420

Query: 439 NWLIGNLNWISKWLVPFLIPGPTLSKKVPRDRTKTVVHTHWFWGSVTFVSYLGVATVIQF 498
 NW IG++ SK ++ F + Y+G VI
 Sbjct: 421 NW-IGDMVVFSSKNVLF--IQLSFYVGMIVILG 451

Query: 499 VDLYLPPAKYPDLWVILNTTSLFACGLFWLW 531
 +D PP++YPDLWVILN TLSFA F +LW
 Sbjct: 452 IDLFIPPSRYPDWVILNVTLFAGFTIYLW 484

D. melanogaster

Score = 247 bits (630), Expect = 4e-64
 Identities = 175/490 (35%), Positives = 267/490 (54%), Gaps = 55/490 (11%)

Query: 57 VGLGSYSGFNTPPMYGDFAQRHWMEITQHLSEIKWYF----YDLQYWGLDYPPLTAFHS 112
 + L SYSGF++PPM+GD+EAQRHW EIT +L++ +WY DLQYWGLDYPPLTA+HS
 Sbjct: 19 ISLYSYSGFDSPPMHGDYEAQRHWQEITVNLAVGEWYTNSSNNDLQYWGLDYPPLTAYHS 78

Query: 113 YFFGKLGSFINPAWFALDVSRGFESVDLKSVMRTAILSELLCFIPAVIWCYCRMGLNYF 172
 Y G++G+ I+P + L SRGFES + K +MRAT + +++L ++PA++ + +
 Sbjct: 79 YLVGRIGASIDPRFVELHKSRRGFESKEHKRMRATVVSADVLIYLPAMLLLAYSLDKAFR 138

FIGURE 27 (sheet 3)

Query: 173 NQNAIEQTIIASAILFNPSLIIIDHGHFQYNSVMLGFALLSILNLLYDNFALAAIFFVLS 232
 + + + + +A P +ID+GHFQYN++ LGFA ++I +L F AA FF L+
 Sbjct: 139 SDDKLFLEFTLVAAY---PGQTLIDNGHFQYNNISLGFAAFAIAAILRRRFYAAAFFFTLA 195

Query: 233 ISFKQMALYYSPIMFFYMLSVCWPLKNFN--LLRLATISIAVLLTFATLLLPFVLVDGM 290
 +++KQM LY+S + FF L C K+F + ++ I+ VL TFA L +P+ + +
 Sbjct: 196 LNYKQMELYHS-LPFFAFLGECVSQKSFASFIAEISRIA AAVVLGTFAILWVPW--LGSL 252

Query: 291 SQIGQILFRVFPFSRGLFEDKVANFWCTTNILVKYKQLFTDKTLTRISLVATLIAISPSC 350
 + Q+L R+FP +RG+FEDKVAN WC N++ K K+ ++ + + + TLIA P+
 Sbjct: 253 QAVLQVLHRLFPVARGVFEDKVANVWCAVNVVWKLKKHISNDQMALVCIACLTIASLPTN 312

Query: 351 FIIFTHPKKVLLPWAFACSWAFYLFQVHEKSXXXXXXXXXXXXXKDLDIISMVCW- 409
 ++F V A S AF+LFSFQVHEK+ + + CW
 Sbjct: 313 VLLFRRTNVGFLLENTSLAFFLFSFQVHEKTILLTALPA-----LFLKWCWP 362

Query: 410 -----ISNIAFFSMWPLLKRDLALEYFVLGILSNWLIGNLNWISKWLVPSPFLIPGPTLS 464
 + FSM PLL RD L + V + + + + SK LS
 Sbjct: 363 DEMILFLEVTVFSMLPLLARDELLVPAVVATVAFHLIFKCFDSKSK-----LS 410

Query: 465 KKVPKRDTKTVVHTHWFWSVTFVSYL GATVIQFVDWLYLP-PAKYPDLWVILNTTLSFA 523
 + P + + + +S + A+ L +P P KYPDLW ++ + S
 Sbjct: 411 NEYPLKYIANI-----SQILMISVVVAS-----LTVPAPTKYPDLWPLIISVTSCG 456

Query: 524 CFGLEFWLWIN 533
 F LF+LW N
 Sbjct: 457 HFFLEFLWGN 466

A. thaliana

Score = 244 bits (622), Expect = 3e-63

Identities = 187/488 (38%), Positives = 248/488 (50%), Gaps = 39/488 (7%)

Query: 62 YSGFNTPPMYGDFAQRHWMEITQHLSIEKWY----FYDLQYWGLDYPPLTAFHSYFFGK 117
 YSG PP +GDFAQRHWMEIT +L + WY + DL YWGLDYPPLT+ SY G
 Sbjct: 61 YSGAGIPPKFGDFAQRHWMEITTNLPVIDWYRNGTYNDLTWGLDYPPLTAYQSYIHGI 120

Query: 118 LGSFINPAWFALDVSRGFESVDLKS YMRATAILSELLCFIPAVIWIYCRWMGLNYFNQNAI 177
 F NP AL SRG ES K MR T + S+ F PA +++ N
 Sbjct: 121 FLRFFNPESVALLSSRGHESYLGKLLMRWTVLSSDAFIFFPALFFVLVYHRNRTRGGKS 180

Query: 178 EQTIIASAILFNPSLIIIDHGHFQYNSVMLGFALLSILNLLYDNFALAAIFFVLSISFKQ 237
 E + IL NP LI+IDHGHFQYN + LG + +I +L ++ L + F L++S KQ
 Sbjct: 181 EVAWHIAMILLNPCLILIDHGHFQYNCISLGLTVGAIAAVLCESEVLTCTVLFSLALSHKQ 240

Query: 238 MALYYSPIMFFYMLSVCWPLKNFNLLRLATISIAVLLTFATLLLPFVLVDGMSQIGQIL 297
 M+ Y++P F ++L C K+ +L + + IAV++TF P+ V + +L
 Sbjct: 241 MSAYFAPAFFSHLLG-KCLRRKS-PILSVIKGLGIAVIVTFVIFWWPY--VHSLDDFLMVL 296

Query: 298 FRVFPFSRGLFEDKVANFWCTTNILVKYKQLFTDKTLTRISLVATLIAISPSCFIIFTHP 357
 R+ PF RG++ED VANFWCTT+IL+K+K LFT ++L ISL AT++A PS P
 Sbjct: 297 SRLAPFERGIYEDYVANFWCTTSILIKWKNLFTTQSLKSISLAATILASLPSMVQQLSP 356

Query: 358 KKVLLPWAFACSWAFYLFQVHEKSXXXXXXXXXXXXXKDLDIISMVCWISNIAFFS 417
 + S AFYLFQVHEKS L + ++ A FS
 Sbjct: 357 SNEGFLYGLLNSSMAFYLFQVHEKSILMPFLSATLLA---LKLDPHFHSHLTYALFS 412

FIGURE 27 (sheet 4)

Query: 418 MWPLLKR DGLALEYFVLGILSNWLI---GNLWISKWLVP SFL---IPGPTLSKKVPKRD 471
M+PLL RD L + Y L L + GN + I K V F PG
Sbjct: 413 MFPLLCRDKLLIPYLTLSFLFTVIYHSPGNHHAIQKTDVSFFSFKNFPGYVF----- 464

Query: 472 TKT VVH THWFWGSVTFVS YLGATVIQFVDWLYLPPAKYPDLWVILNTTLSFACFGLFWLW 531
++ TH+F V V YL PP KYP L+ L L F+ F +F +
Sbjct: 465 ---LLRTHFFISVVLHVLYLTIK-----PPQKYPFLFEALIMILCFSYFIMFAFY 511

Query: 532 INYNLYIL 539
NY + L
Sbjct: 512 TNYTQWTL 519

FIGURE 28

K. lactis ALG6

ATCTCTGTTTCAACAGCTCTTGCATTCATTGGTTCTTTCGGTCCAATCTATA
TCTTTGGAGGATACAAGAAGCTTAGTGCAATCAATGCACAGGATTTTTCCAT
TTGCCAGGGGTATCTTTGAAGATAAAGTTGCGAATTTTTGGTGCGTTTCTA
ATATTTTCATCAAATATAGAAATCTATTCCTCAGAAAGGATCTTCAATTAT
ACTCATTACTCGCAACAGTTATTGGGCTTTTACCATCATTATTATAACAT
TTTTATACCCGAAGAGACATTTACTACCATATGCTTTGGCCGCATGTTTCA
TGTCATTCTTCTTATTAGCTTCCAGGTTTCATGAAAAGACAATCTTATTAC
CTTTACTTCCTATTACACTCTTGTACACGTCAAGAGATTGGAATGTTCTAT
CATTGGTTTGTGTTGGATTAAACAACGTGGCATTGTTTACACTCTGGCCATTAC
TGAAAAAGGACAATCTAGTATTGCAATATGGAGTCATGTTTCATGTTTAGC
AATTGGTTGATCGGTAACCTTCAGTTTCGTACACACCACGCTTCCTCCCAAAA
TTTTTGACACCAGGGCCATCCATCAGTGATATAGATGTTGATTATAGACGG
GCAAGTTTACTACCCAAGAGCCTAATATGGAGATTAATCATTGTTGGCTCA
TATATTGCAATGGGGATTATTCATTTTCTAGACTATTACGTCTCCCCGCCA
TCAAAATACCCTGATTTATGGGTGCTTGCCAATTGTTTCTTGGGCTTCTCA
TGTTTTGTGACATTTTGGATATGGAACAATTATAATTATTCGAAATGAGAA
ACAGCACTTTGCAAGATTTA

K. lactis Alg6p

ISVSTALAFIGSFGPIYIFGGYKNLVQSMHRIFPFARGIFEDKVANFWCVSNIFIK
YRNLFTQKDLQLYLLATVIGLLPSFIITFLYPKRHLLPYALAACSMSFFLFSFQ
VHEKTILLPLLITLLYTSRDWNVLSLVCWINNVALFTLWPLLKKDNLVLQYG
VMFMFSNWLIGNFSFVTPRFLPKFLTPGPSISDIDVDYRRASLLPKSLIWRLIIV
GSYIAMGIIHFLDYYVSPPSKYPDLWVLANCSLGFSCFVTFWIWNNYNYSKZE
TALCKI

FIGURE 29 (sheet 1)

K. lactis ALG6 BLAST

Score E
Sequences producing significant alignments: (bits) Value

gi 1420090 emb CAA99190.1	ORF YOR002w [Saccharomyces cerev...]	392	e-108
gi 7490584 pir T40396	glucosyltransferase - fission yeast ...	187	2e-46
gi 15240920 ref NP_198662.1	glucosyltransferase-like prote...	117	2e-25
gi 7019325 ref NP_037471.1	dolichyl-P-Glc:Man9GlcNAc2-PP-d...	103	2e-21
gi 12002040 gb AAG43163.1 AF063604.1	brain my046 protein [H...]	102	8e-21
gi 19921070 ref NP_609393.1	CG5091-PA [Drosophila melanoga...]	101	1e-20

Alignments

S. cerevisiae

Score = 392 bits (1006), Expect = e-108
Identities = 182/280 (65%), Positives = 218/280 (77%), Gaps = 1/280 (0%)
Frame = +1.

Query: 1 ISVSTALAFIGSFGPIYIFGG-YKNLVQSMHRIFPPFARGIFEDKVANFWCVSNIFIKYRN 177
I+ +T F F P+Y GG KN+ Q +HRIFPPFARGIFEDKVANFWCV+N+F+KY+
Sbjct: 265 IAFATLATFAIIFAPLYFLGGGLKNIHQCIHRIFPPFARGIFEDKVANFWCVTNVFKYKE 324

Query: 178 LFTQKDLQLYSLLATVIGLLPSFIITFLYPKRHLLPYALAAACSMSSFFLFSFQVHEKXXXX 357
FT + LQLYSL+ATVIG LP+ I+T L+PK+HLLPY L ACSMSFFLFSFQVHEK
Sbjct: 325 RFTIQQLQLYSLIATVIGFLPAMIMTLLHPKKHLLPYVLIACSMSSFFLFSFQVHEKTILI 384

Query: 358 XXXXXXXXYTSRDWNVLSLVSWINNVALFTLWPLLKKDNLVLQYGVMMFMSNWLIGNFSF 537
Y+S DWNVLSLV WINNVALFTLWPLLKKD L LQY V F+ SNWLIGNFSF
Sbjct: 385 PLLPITLLYSSTDWNVLSLVSWINNVALFTLWPLLKKDGLHLQYAVSFLLSNWLIGNFSF 444

Query: 538 VTPRFLPKFLTPGPSISDIDVDYRRASLLPKSLIWRLLIIVGSYIAMGIIHFLDYVSPPS 717
+TPRFLPK LTPGPSIS I+ DYRR SLLP +++W+ I+G+YIAMG HFLD +V+PPS
Sbjct: 445 ITPRFLPKSLTPGPSISSINDYRRSLLPYNVVWKSFIIGTYIAMGFYHFLDQFVAPPS 504

Query: 718 KYPDLWVLNCSLGFSCFVTFWIWNNYXLFEMRNSTLQDL 837
KYPDLWVL NC++GF CF FW+W+ Y +F + +++DL
Sbjct: 505 KYPDLWVLLNCAVGFCFSIFWLWSYKIFTSGSKSMKDL 544

S. pombe

Score = 187 bits (475), Expect = 2e-46
Identities = 106/280 (37%), Positives = 150/280 (53%), Gaps = 1/280 (0%)
Frame = +1

Query: 1 ISVSTALAFIGSFGPIYIFGGYKNLV-QSMHRIFPPFARGIFEDKVANFWCVSNIFIKYRN 177
+SV+ F P +I+ YK L+ Q +HR+PPFARG++EDKVANFWC N K R
Sbjct: 251 LSVTVVFTFSLILFP-WIYMDYKTLPLQILHRVFPFARGLWEDKVANFWCTLNVTVEKIRE 309

Query: 178 LFTQKDLQLYSLLATVIGLLPSFIITFLYPKRHLLPYALAAACSMSSFFLFSFQVHEKXXXX 357
+FT LQ+ SL+ T+I +LPS +I FLYP++ LL A+ S FFLFSFQVHEK
Sbjct: 310 VFTLHQLQVISLIFTLISILPSCVILFLYPRKRLALGFASASWGFFLFSFQVHEKSVLL 369

FIGURE 29 (sheet 2)

Query: 358 XXXXXXXXYTSRDWNVLSLVCWINNVALFTLWPLLKKDNLVLQYGVMMFMSNWLIGNFSF 537
 + + NN+A+F+LWPLLKKD L LQY + + NW
 Sbjct: 370 PLLPTSILLCHGNITTKPWIALANNLAVFSLWPLLKKDGLGLQYFTLVLMWNW----- 422

Query: 538 VTPRFLPKFLTPGPSISDIDVDYRRASLLPKSLIWRLLIIVGSYIAMGIIHFLDYVSPPS 717
 I D+ V K++++R I + Y+ M +I +D ++ PPS
 Sbjct: 423 -----IGDMVV-----FSKNVLFERFIQLSFYVGMIVILGIDLFIPPPS 460

Query: 718 KYPDLWVLANCSLGFSCFVTFWIWNNYXLFEMRNSTLQDL 837
 +YPDLWV+ N +L F+ F T ++W L + + DL
 Sbjct: 461 RYPDLWVILNVTLSFAGFFTIYLWTLGRLLHISSKLSTDL 500

A. thaliana

Score = 117 bits (292), Expect = 2e-25
 Identities = 81/240 (33%), Positives = 120/240 (50%), Gaps = 2/240 (0%)
 Frame = +1

Query: 85 MHRIFPFARGIFEDKVANFWCVSNIFIKYRNLFQKDLQLYSLLATVIGLLPSFIITFLY 264
 + R+ PF RGI+ED VANFWC ++I IK++NLFT + L+ SL AT++ LPS + L
 Sbjct: 296 LSRLAPFERGIYEDYVANFWCTTSILIKWKNLFTTQSLKSISLAATILASLPSMVQQILS 355

Query: 265 PKRHLLPYALAACSMSFFLFSFQVHEKXXXXXXXXXXXXXYTSRDWNVLSLVCWINNVALF 444
 P Y L SM+F+LFSFQVHEK + L + ALF
 Sbjct: 356 PSNEGFLYGLNSSMAFYLFQVHEKSILMPFLSATLLALKLPDHFHSHLTY---ALF 411

Query: 445 TLWPLLKKDNLVLQYGVMMFMSNWLIGNFSFVTPRFLPKFLTPG--PSISDIDVDYRRAS 618
 +++PLL +D L++ Y + SF+ F + +PG +I DV +
 Sbjct: 412 SMFPLLCRDKLLIPYLT-----SFL---FTVIYHSPGNHAIQKTDVSFFSFK 457

Query: 619 LLPKSLIWRLLIIVGSYIAMGIIHFLDYVSPPSKYPDLWVLANCSLGFSCFVTFWIWNNY 798
 P + L+ +I++ ++H L + PP KYP L+ L FS F+ F + NY
 Sbjct: 458 NFPGYVF--LLRTHFFISV-VLHVLYLTIKPPQKYPFLFEALIMILCFSYFIMFAFYTN 514

H. sapiens

Score = 103 bits (258), Expect = 2e-21
 Identities = 78/266 (29%), Positives = 123/266 (46%), Gaps = 3/266 (1%)
 Frame = +1

Query: 7 VSTALAFIGSFGPIYI--FGGYKNLVQSMHRIFPFARGIFEDKVANFWCVSNIFIKYRN 180
 V A + SF ++ F + +Q + R+FP RG+FEDKVAN WC N+F+K +++
 Sbjct: 232 VKLACIVVASFVLCWLPFFTEREQTLQVLRRLFPVDRGLFEDKVANIWCSFNVFLKIKDI 291

Query: 181 FTQKDLQLYSLLATVIGLLPSFIITFLYPKRHLLPYALAACSMSFFLFSFQVHEKXXXXX 360
 + + S T + LLP+ I L P + L +C++SFFLFSFQVHEK
 Sbjct: 292 LPRHIQLIMSFCTFLSLLPACIKLILQPSSKGKFTLVSCALSFFLFSFQVHEKSILLV 351

Query: 361 XXXXXXXXYTSRDWNVLSLVCWINNVALFTLWPLLKKDNLVLQYGVMMF-FSNWLIGNFSF 537
 + + + W V+ F++ PLL KD L++ V M F + +FS
 Sbjct: 352 SLPVCLVLS----EIPFMSTWFLLVSTFMSMLPLLKDELLMPSVVTMAFFIACVTSFSI 407

Query: 538 VTPRFLPKFLTPGPSISDIDVDYRRASLLPKSLIWRLLIIVGSYIAMGIIHFLDYVSPPS 717
 + SIS V S I + + + S I M ++ + + PP
 Sbjct: 408 FEKTSEELQLKSFSIS---VRKYLPCFTFLSRIIQYLF LISVITMVLTLTMTVTLDP 464

Query: 718 KYPDLWVLANCSLGFSCFVTFWIWNN 795
 K PDL+ + C + F+ F ++ N
 Sbjct: 465 KLPDLFSVLVCFVSCLNLFLLVYFN 490

FIGURE 30

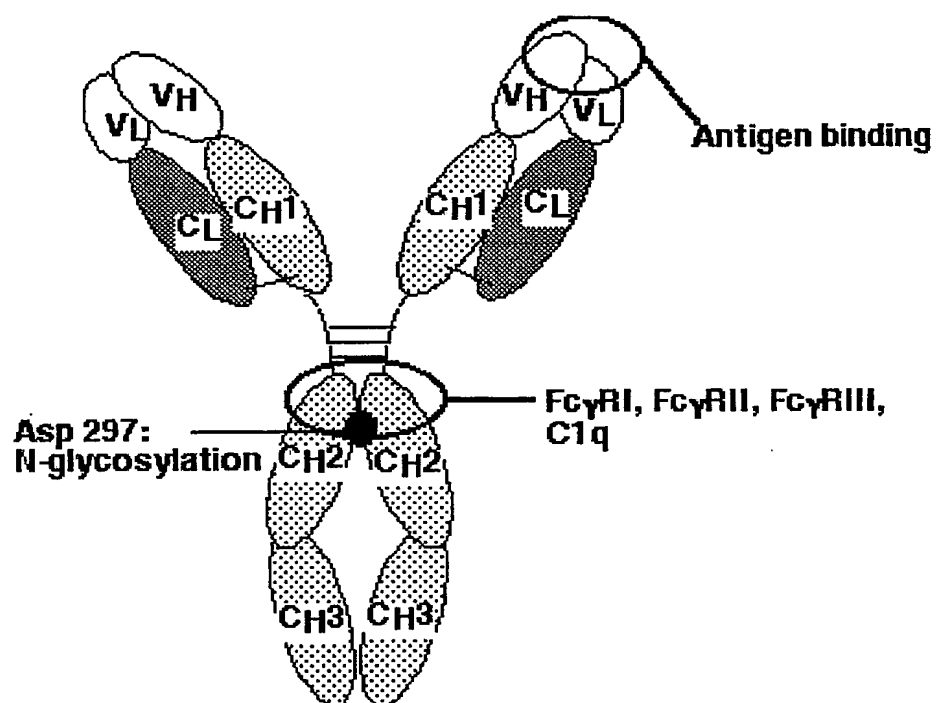


FIGURE 31

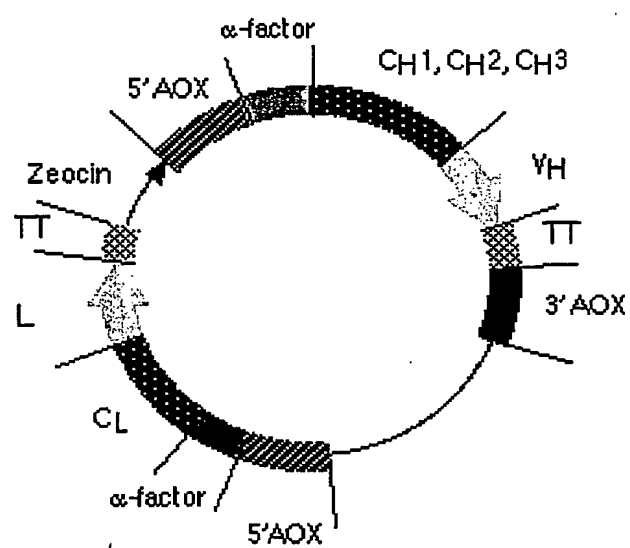


FIGURE 34 (sheet 2)

TCATTCAAACCTGAAAACAAAACAGGAAGAGGGAATTGAGCCAATTGGGAAGGACTTT
GGGGCCGATCCTAAACCAATTAATTTATTTATTTGGGAGGATGGGGGCGGGCTCGGG
AGGGAGGAGAGGGGTTGAACAGTTTCCTTTTGTTCCTCACTGTTAATTCGCCCACCT
TCGGGCCCTTCTTGTCTGCAGCGCCAAGCAGGGTGCAGAGGGGCTGTGGCTTGCTT
GAGGGGCCACTGTGGGGCTTCACTCCTGGTCACAGGTGGCAGCAGAGAAAAGAGATG
TCTATAAGCAGGGGGATGTAGCTCAGTTTGTAGAATGCTTGCATAGCATAAATGAAG
TCCTGGGTTCATCCCCAGCACCATATAAATGCAGGTAAGAAACAGAGTCAGGAGGA
CCAAGCATTCTCCTTGGCTACATAACAAAAGCAAGGCCTTTGTCCCCATGTCTTGGC
TACAAGAGACCCTATCTCAGAAAATTGTGGGGGGGAGGGGGGGGAAATGGCCTTGA
AAACACAGCCAGTCAGTGTCACTGCATTGCCAGAACTGGTGGATCCCAGGTGTGCTT
GGCAGATAACAGCTAAAAGGCACATAACCTTGGTGGGGAAATAAATGCCTGTGGTGT
CCTGAGGGCCCCACCAAGTTCCAAAAA

>gi|18997007|gb|AAL83249.1|AF474154_1 N-
acetylglucosaminyltransferase V [Mus musculus]

MAFFSPWKLSSQKLGFFLVTFGFIWGMMLLHFTIQQRTQPESSSMLREQILDLSKRY
IKALAEENRDVVDGPYAGVMTAYDLKKTAVLLDNILQRIKLESKVDNLVNGTGAN
STNSTTAVPSLVSLEKINVADIINGVQEKCVLPMDGYPHCEGKI KWMKDMWRSDPC
YADYGV DGTSCSFFIYLSEVENWCPRLPWRANKPYEEADHNSLAEIRTD FNILY GMM
KKHEEFRWMRLRIRRMADAWIQAIKSLAEKQNLKRRKKILVHLGLLT KESGFKIA
ETAFSGGPLGELVQWSDLITSLYLLGHDIRISASLAE LKEIMKKVVGNRSGCPTVGD
RIVELIYIDIVGLAQFKKTLGPSWVHYQCMLRVLDSFGTEPEFNHAS YAQSKGHKTP
WGKWNLPQQFYTMFPHTPDNSFLGFVVEQHLNSSDIHHINEIKRQNQSLVYGKVDS
FWKNKKIYLDIIHTYMEVHATVYGSSTKNIPSYVKNHGILSGRDLQFLLRETKLFVG
LGFPYEGPAPLEAIANGCAFLNPKFNPPKSSKNTDFFIGKPTLRELTSQHPIAEVFI
GRPHVWTVDLN NREEVEDAVKAILNQKIEPYMPYEFTCEGMLQRINAFIEKQDFCHG
QVMWPPLSALQVKLAEPGQSCQVCQESQLICEPSFFQHLNKEKD LLYKVTCQSSE
LYKDILVPSFYPKSKHCVFQGD LLLFSCAGAHPTHQRI CPCRDFIKQVALCKDCL

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US02/41510

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : A01K 67/027; C12N 9/10, 1/04, 1/16, 1/18

US CL : 800/13-18; 435/193, 252.3, 254.11, 254.51, 254.21, 254.23, 254.3, 254.4, 254.6, 254.7, 325, 463

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 800/13-18; 435/193, 252.3, 254.11, 254.51, 254.21, 254.23, 254.3, 254.4, 254.6, 254.7, 325, 463

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
EAST, CAPLUS, BIOSIS, EMBASE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	MIELE, R. G. et al. Glycosylation of Asparagine-28 of Recombinant Staphylokinase with High-Mannose-Type Oligosaccharides Results in a Protein with Highly Attenuated Plasminogen Activator Activity. Journal of Biological Chemistry. 19 March 1999, Volume 274, No. 12, pages 7769-7776.	1-60
A	MARTINET, W. et al. Modification of the Protein Glycosylation Pathway in the Methylophilic Yeast Pichia Pastoris. Biotechnology Letters. December 1998, Volume 20, No. 12, pages 1171-1177.	1-60
A	MARAS, M. et al. In Vitro Conversion of the Carbohydrate Moiety of Fungal Glycoproteins to Mammalian-Type Oligosaccharides. Journal of Biochemistry. 1997., Volume 249, pages 701-707.	1-60
A	RAJU, T. S. et al. Species-Specific Variation in Glycosylation of IgG: Evidence for Species-Specific Sialylation and Branch-Specific Galactosylation and Importance for Engineering Recombinant Glycoprotein Therapeutics. Glycobiology. 2000, Volume 10, No. 5, pages 477-486.	1-60



Further documents are listed in the continuation of Box C.



See patent family annex.

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"E" earlier application or patent published on or after the international filing date	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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"O" document referring to an oral disclosure, use, exhibition or other means	"&" document member of the same patent family
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Date of the actual completion of the international search

15 May 2003 (15.05.2003)

Date of mailing of the international search report

09 JUN 2003

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Telephone No. 703-308-0196

INTERNATIONAL SEARCH REPORT

PCT/2/41510

C. (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	OMTVEDT, L. A. et al. Glycosylation of Immunoglobulin Light Chains Associated with Amyloidosis. <i>Amyloid: International Journal of Experimental and Clinical Investigation</i> . 2000, Volume 7, pages 227-244.	1-60
A	FUKUTA, K. et al. Comparative Study of the N-Glycans of Human Monoclonal Immunoglobulins M Produced by Hybridoma and Parental Cells. <i>Archives of Biochemistry and Biophysics</i> . 01 June 2000, Volume 378, No. 1, pages 142-150.	1-60
A	TAKAHASHI, N. et al. Comparative Structural Study of the N-Linked Oligosaccharides of Human Normal and Pathological Immunoglobulin G. <i>Biochemistry</i> , 1987, Volume 26, pages 1137-1144.	1-60
A	KATO, J. et al. Nucleotide Sequence of a Regulatory Region Controlling Alginate Synthesis in <i>Pseudomonas aeruginosa</i> : Characterization of the AlgR2 Gene. <i>Gene</i> . 1989, Volume 84, pages 31-38.	1-60
A	DUMAN, J. G. et al. O-Mannosylation of <i>Pichia Pastoris</i> Cellular and Recombinant Proteins. <i>Biotechnology and Applied Biochemistry</i> . 1998, Volume 28, pages 39-45.	1-60
A	MIELE, R. G. et al. Glycosylation Properties for the <i>Pichia Pastoris</i> -Expressed Recombinant Kringle 2 Domain of Tissue-Type Plasminogen Activator. <i>Biotechnology and Applied Biochemistry</i> . 1997, Volume 25, pages 151-1157.	1-60
A	TREMBLAY, L. O. et al. Cloning and Expression of a Specific Human Alpha 1,2-Mannosidase that Trims Man9GlcNAc2 to Man8GlcNAc2 Isomer B during N-Glycan Biosynthesis. <i>Glycobiology</i> . 1999, Volume 9, Number 10, pages 1073-1079.	1-60
A	VAZQUEZ-REYNA, A. B. et al. Biosynthesis of Glycoproteins in <i>Candida Albicans</i> : Biochemical Characterization of a Soluble Alpha-Mannosidase. <i>FEMS Microbiology Letters</i> . 1993, Volume 106, pages 321-326.	1-60
A	RUNGE, K. W. et al. A New Yeast Mutant in the Glucosylation Steps for the Asparagine-Linked Glycosylation Pathway. <i>Journal of Biological Chemistry</i> . 25 November 1986, Volume 261, No. 33, pages 15582-15590.	1-60
A	SUZUKI, C. Immunochemical and Mutational Analyses of P-Type ATPase Spf1p Involved in the Yeast Secretory Pathway. <i>Bioscience Biotechnology Biochemistry</i> . 2001, Volume 65, Number 11, 2405-2411.	1-60
A	AEBI, M. et al. Cloning and Characterization of the ALG3 Gene of <i>Saccharomyces Cerevisiae</i> . <i>Glycobiology</i> . 1996, Volume 6, No. 4, pages 439-444.	1-60

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